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(54) Title: GENES FOR ALTERING MITOCHONDRIAL FUNCTION AND FOR HYBRID SEED PRODUCTION

(57) Abstract: The present invention relates to isolated nucleic acid molecules which restore fertility to cytoplasmic male sterile plants and modify expression of toxic mitochondria proteins by the plant. The present invention also relates to methods of identifying a candidate plant suitable for breeding with a cytoplasmic male sterile plant and methods of identifying a candidate gene restoring fertility in plants by analyzing for the candidate plant and candidate gene, respectively, for the presence of the nucleic acid molecule of the present invention. Also disclosed are methods of producing hybrid plant seed, methods of directing gene expression to plant mitochondria, and method of expressing a gene preferentially in roots of a plant. Promoters and terminators from plant genes which restore fertility to cytoplasmic male sterile plants and modify expression of toxic mitochondria proteins are also disclosed. Finally, methods of producing plants with a cytoplasmic male sterile plant restoration system are disclosed.

## GENES FOR ALTERING MITOCHONDRIAL FUNCTION AND FOR HYBRID SEED PRODUCTION

[0001] This application claims the benefit of U.S. Patent Application Serial No. 60/347,996, filed January 10, 2002, which is hereby incorporated by reference in its entirety.

[0002] This invention arose out of research sponsored by the USDA NRI (Grant No. 98-35300-6171). The U.S. Government may have certain rights in this invention.

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### FIELD OF THE INVENTION

[0003] The present invention relates to improving productivity or usefulness of plants by altering mitochondrial gene expression and to the production of hybrid seed. Specifically, the present invention relates to the use of genes that affect mitochondrial gene expression, some of which ameliorate male sterility and others which cause male sterility or altered floral development. The invention also provides a method of facilitating the identification of genes with similar functions in other plant species.

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### BACKGROUND OF THE INVENTION

[0004] A widely used method for producing hybrid seeds involves crossing a cytoplasmic male sterile (CMS) plant line with a fertile plant line. Typically, the fertile line contains a fertility restorer gene in its nuclear genome, so that all of the progeny are male fertile. All seeds collected from a CMS plant must result from cross-pollination. However, the hybrid seed so generated will itself be male sterile unless the male parent has brought a nuclear fertility-restorer gene into the next generation. The fertility of the progeny is important for productivity in plant varieties where self-pollination is responsible for production of the desirable crop. For example, a fruit crop of a self-pollinated species requires male fertility, while an ornamental species will produce attractive flowers or plant morphology even when no pollen is produced.

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[0005] While a number of naturally occurring CMS/restorer systems exist and are currently in use for hybrid seed production, there are a number of crop species which lack known CMS and fertility restorer genes. For example, a hybrid seed of tomato is typically made by hand emasculation of plants to be used as female parents.

5 This hand-made method of cross-pollination is quite labor intensive and cost-prohibitive for many crops. In addition, certain naturally occurring CMS/restorer systems have some drawbacks. For example, corn plants carrying the CMS-T cytoplasm are more susceptible to a blight disease.

[0006] Fertility restorer genes that have been particularly useful for hybrid 10 seed production are active as single dominant alleles at a locus, though multigenic systems are sometimes used. A *Petunia* fertility restorer locus termed *Rf* is known to be effective with no additional helper genes to restore fertility (Edwardson et al., "Fertility Restoration in Cytoplasmic Male Sterile *Petunia*," *J. Hered.*, 58:195-196 (1967); Izhar, "Cytoplasmic Male Sterility in Petunia. III. Genetic Control on 15 Microsporogenesis and Male Fertility Restoration," *J. Hered.*, 69:22-26 (1978)).

[0007] Nuclear fertility restoration genes confer normal pollen development upon plants carrying sterility-encoding mitochondria. The mitochondrial genes responsible for causing the male sterility have been identified in a number of species, including *Petunia*, maize, *Brassica*, and common bean. The expression of these 20 CMS-encoding mitochondrial genes is affected by the nuclear restorer genes, as shown for *Rf* in *Petunia* (Pruitt et al., "Cytochrome Oxidase Subunit II Sequences in *Petunia* Mitochondria: Two Intron-Containing Genes and an Intron-Less Pseudogene Associated With Cytoplasmic Male Sterility," *Curr. Genet.*, 16:281-91 (1989); Nivison et al., "Identification of a Mitochondrial Protein Associated With 25 Cytoplasmic Male Sterility in *Petunia*," *Plant Cell*, 1:1121-30 (1989); Nivision et al., "Sequencing, Processing, and Localization of the *Petunia* CMS-Associated Mitochondrial Protein," *Plant J.*, 5:613-623 (1994); Hanson et al., "Mitochondrial Gene Organization and Expression in Petunia Male Fertile and Sterile Plants," *J. Hered.*, 90:362-368 (1999)); *RfI* in CMS-T maize (Dewey et al., "Novel 30 Recombinations in the Maize Mitochondrial Genome Produce a Unique Transcriptional Unit in the Texas Male-Sterile Cytoplasm," *Cell*, 44:439-49 (1986); Wise et al., "Mitochondrial Transcript Processing and Restoration of Male Fertility in T-Cytoplasm Maize," *J. Hered.*, 90:380-385 (1999); Kennell et al., "Influence of

Nuclear Background on Transcription of a Maize Mitochondrial Region Associated With Texas Male Sterile Cytoplasm," Mol. Gen. Genet., 210:399-406 (1987); Kennell et al., "Initiation and Processing of *atp6*, T-*urf13*, and *ORF221* Transcripts From Mitochondria of T Cytoplasm Maize," Mol. Gen. Genet., 216:16-24 (1989)); *Rfp1* and 5 *rfp1* in *Brassica* (Singh et al., Suppression of Cytoplasmic Male Sterility by Nuclear Genes Alters Expression of a Novel Mitochondrial Gene Region," Plant Cell, 3:1349-1362 (1991); Singh et al., "Nuclear Genes Associated With a Single Brassica CMS Restorer Locus Influence Transcripts of Three Different Mitochondrial Gene Regions," Genetics, 143:505-516 (1996)); restorers in radish (Krishnasamy et al., 10 "Organ-Specific Reduction in the Abundance of a Mitochondrial Protein Accompanies Fertility Restoration in Cytoplasmic Male-Sterile Radish," Plant Molec. Biol., 26:935-946 (1994)); restorers in sunflower (Horn et al., "A Mitochondrial 16 kDa Protein is Associated With Cytoplasmic Male Sterility in Sunflower," Plant Molec. Biol., 17:29-36 (1991); Laver et al., "Mitochondrial Genome Organization and 15 Expression Associated With Cytoplasmic Male Sterility in Sunflower (*Helianthus annuus*)," Plant J., 1:185-193 (1991); Monéger et al., "Nuclear Restoration of Cytoplasmic Male Sterility in Sunflower is Associated With the Tissue-Specific Regulation of a Novel Mitochondrial Gene," EMBO J., 13:8-17 (1994); Smart et al., "Cell-Specific Regulation of Gene Expression in Mitochondria During Anther 20 Development in Sunflower," Plant Cell, 6:811-825 (1994)); restorers in rice (Akagi et al., "A Unique Sequence Located Downstream From the Rice Mitochondrial *atp6* May Cause Male Sterility," Curr. Genet., 25:52-58 (1994); Kadowaki et al., "A Chimeric Gene Containing the 5' Portion of *atp6* is Associated With Cytoplasmic Male Sterility of Rice," Mol. Gen. Genet., 224:10-16 (1990)); and *Fr2* in broad bean 25 (Chase, "Expression of CMS-Unique and Flanking Mitochondrial DNA Sequencs in *Phaseolus vulgaris*," L. Curr. Genet., 25:245-251 (1993); He et al., "Pollen Fertility Restoration by Nuclear Gene *Fr* in CMS Bean: Nuclear-Directed Alteration of a Mitochondrial Population," Genetics, 139:995-962 (1995)). The expression of various nuclear restorer genes has been reported to be either enhanced in reproductive 30 tissue, as in the case of sunflower, or, as in the case of *Petunia*, expressed in both vegetative and reproductive tissues. Thus, different fertility restorer genes carry different promoters and nuclear expression regulatory elements which may confer very limited tissue-specific expression or very broad expression in the plant.

[0008] Reduction in the amount of the protein product of the CMS-encoding gene is the usual effect of these restorers whose target mitochondrial genes are known. These genes may possibly act by affecting the transcription or translation rate, the transcript or protein stability, processing, splicing, etc. Alleles of some 5 restorer genes may up-regulate while others may down-regulate the expression of particular mitochondrial genes. Fertility restorer genes and their alleles or homologous counterparts in other species may thus be extremely valuable in engineering the expression of genes introduced into higher plant mitochondria.

[0009] The cloning and sequencing of the restorer gene *Rf2* in maize has been 10 reported in Cui et al., "The *rf2* Nuclear Restorer Gene of Male-Sterile T-Cytoplasm Maize," Science, 272:1334-1336 (1996) and U.S. Patent No. 5,981,833 to Wise et al. This restorer gene acts in conjunction with a second required gene, *Rf1*, the gene that reduces the amount of the toxic protein, to restore fertility to plants carrying the maize 15 CMS-T cytoplasm (Dewey et al., "Novel Recombinations in the Maize Mitochondrial Genome Produce a Unique Transcriptional Unit in the Texas Male-Sterile Cytoplasm," Cell, 44:439-49 (1986); Dewey et al., "A Mitochondrial Protein Associated With Cytoplasmic Male Sterility in the T Cytoplasm of Maize," Proc. Natl. Acad. Sci. USA, 84:5374-78 (1987); Wise et al., "*Urf13-T* of T Cytoplasm Maize Mitochondria Encodes a 13kD Polypeptide," Plant Mol. Biol., 9:121-26 20 (1987)). Plants of genotype *Rf1Rf2*, though sterile, have greatly reduced amounts of the URF13 protein. In contrast, sterile plants of genotype *rf1Rf2* have abundant amounts of the URF13 protein. The *Rf2* gene is, thus, unusual in that no effect on the expression of the maize T-CMS-associated protein, URF13, has been detected. The sequence of the gene bore out the absence of observable effect on mitochondrial gene 25 expression; according to sequence analysis, *Rf2* is apparently an aldehyde dehydrogenase (Liu et al., "Mitochondrial Aldehyde Dehydrogenase Activity is Required for Male Fertility in Maize," The Plant Cell, 13:1063-1078 (2001)). It has been proposed that *Rf2* acts by compensating for a metabolic defect caused by the low levels of the URF13 protein that remain despite the presence of *Rf1*, the gene that 30 reduces the amount of the toxic protein (Dewey et al., "A Mitochondrial Protein Associated With Cytoplasmic Male Sterility in the T Cytoplasm of Maize," Proc. Natl. Acad. Sci. USA, 84:5374-78 (1987)) and also alters the T-*urf13* transcript profile (Kennell et al., "Influence of Nuclear Background on Transcription of a Maize

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Mitochondrial Region Associated With Texas Male Sterile Cytoplasm," Mol. Gen. Genet., 210:399-406 (1987)).

- [0010] An abnormal recombinant mitochondrial gene in *Petunia* CMS lines (termed *pcf*) has been genetically correlated with CMS (Young et al., "A Fused 5 Mitochondrial Gene Associated With Cytoplasmic Male Sterility is Developmentally Regulated," Cell, 50:41-49 (1987)). Because plant mitochondrial RNA is edited from C to U in some locations, the edited RNA sequence for the *pcf* gene has been determined, allowing the prediction of the *pcf*-encoded protein (Wintz et al., "A Termination Codon is Created by RNA Editing in the *Petunia* Mitochondrial *atp9* 10 Gene Transcript," Curr. Genet., 19:61-64 (1990); Sutton et al., "Editing of Pre-mRNAs Can Occur Before *cis*- and *trans*-Splicing in *Petunia* Mitochondria," Mol. Cell Biol., 11:4274-4277 (1991); Nivision et al., "Sequencing, Processing, and Localization of the *Petunia* CMS-Associated Mitochondrial Protein," Plant J., 5:613-623 (1994); Hanson et al., "Mitochondrial Gene Organization and Expression in 15 Petunia Male Fertile and Sterile Plants," J. Hered., 90:362-368 (1999)). Antibodies to synthetic peptide sequences have revealed the presence of a 19.5 kD PCF protein located in both the membrane and soluble fraction of mitochondria (Nivison et al., "Identification of a Mitochondrial Protein Associated With Cytoplasmic Male Sterility in *Petunia*," Plant Cell, 1:1121-30 (1989)). The PCF protein is processed 20 from a longer precursor protein and is entirely encoded by the *urfS* region of the *pcf* gene (Nivision et al., "Sequencing, Processing, and Localization of the *Petunia* CMS-Associated Mitochondrial Protein," Plant J., 5:613-623 (1994)). The PCF protein is strongly expressed in sporogenous cells of premeiotic petunia anthers in CMS lines, but undetectable in CMS-*Rf* lines (Conley et al., "Tissue-Specific Protein Expression 25 in Plant Mitochondria," Plant Cell, 6:85-91 (1994)). Abnormalities in *Petunia* pollen development are first observed in meiosis, and by the developmental stage where fertile plants are releasing pollen, CMS anthers are hollow shells (Conley et al., "Effects of *Petunia* Cytoplasmic Male Sterile (CMS) Cytoplasm on the Development of Sterile and Fertility-Restored *P. parodii* Anthers," Am. J. Bot., 81:630-640 (1994)). 30 It is evident that the *pcf* gene product is disrupting mitochondrial function, leading to death of the sporogenous cells, though the exact mechanism at the molecular level is not known.

[0011] In maize T, *Petunia*, rice, and *Brassica* Pol CMS systems, particular transcripts of CMS-associated genes have been reported to be altered in restored lines (Pruitt et al., "Transcription of the *Petunia* Mitochondrial CMS-Associated *pcf* Locus in Male Sterile and Fertility-Restored Lines," *Mol. Gen. Genet.*, 227:348-355 (1991); 5 Dewey et al., "Novel Recombinations in the Maize Mitochondrial Genome Produce a Unique Transcriptional Unit in the Texas Male-Sterile Cytoplasm," *Cell*, 44:439-49 (1986); Kennell et al., "Initiation and Processing of *atp6*, T-*urf13*, and *ORF221* Transcripts From Mitochondria of T Cytoplasm Maize," *Mol. Gen. Genet.*, 216:16-24 (1989); Kennell et al., "Influence of Nuclear Background on Transcription of a Maize 10 Mitochondrial Region Associated With Texas Male Sterile Cytoplasm," *Mol. Gen. Genet.*, 210:399-406 (1987); Singh et al., "Suppression of Cytoplasmic Male Sterility by Nuclear Genes Alters Expression of a Novel Mitochondrial Gene Region," *Plant Cell*, 3:1349-1362 (1991); Singh et al., "Nuclear Genes Associated With a Single Brassica CMS Restorer Locus Influence Transcripts of Three Different Mitochondrial 15 Gene Regions," *Genetics*, 143:505-516 (1996); Wise et al., "Mitochondrial Transcript Processing and Restoration of Male Fertility in T-Cytoplasm Maize," *J. Hered.*, 90:380-385 (1999)). In *Brassica*, the presence of either one of two restorer genes results in monocistronic transcripts of *atp6*, instead of the dicistronic *orf224/atp6* transcripts found in CMS lines (Singh et al., "Suppression of Cytoplasmic Male 20 Sterility by Nuclear Genes Alters Expression of a Novel Mitochondrial Gene Region," *Plant Cell*, 3:1349-1362 (1991)). A UG-rich sequence appears to be the target of the *Brassica* restorer alleles (Singh et al., "Nuclear Genes Associated With a Single *Brassica* CMS Restorer Locus Influence Transcripts of Three Different Mitochondrial Gene Regions," *Genetics*, 143:505-516 (1996)). In *Petunia*, *pcf* 25 transcripts with 5' termini at -121 are specifically reduced in restored lines (Pruitt et al., "Transcription of the *Petunia* Mitochondrial CMS-Associated *pcf* Locus in Male Sterile and Fertility-Restored Lines," *Mol. Gen. Genet.*, 227:348-355 (1991)), while transcripts terminating at -266 and -522 remain at normal levels. In maize T cytoplasm, a sequence unlike either the *Brassica* restorer target or the *Petunia* -121 30 transcript terminus is the putative recognition signal for the *Rf1* gene (Dill et al., "*Rf8* and *Rf\** Mediate Unique T-*urf13*-Transcript Accumulation, Revealing a Conserved Motif Associated With RNA Processing and Restoration of Pollen Fertility in T- cytoplasm Maize," *Genetics*, 147:1367-1379 (1997)).

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[0012] The steady-state amounts of the *Petunia pcf*-encoded protein and the maize *urf13*-encoded protein decrease greatly in restored lines compared to unrestored lines (Nivison et al., "Identification of a Mitochondrial Protein Associated With Cytoplasmic Male Sterility in *Petunia*," Plant Cell, 1:1121-30 (1989); Dewey et al., 5 "Novel Recombinations in the Maize Mitochondrial Genome Produce a Unique Transcriptional Unit in the Texas Male-Sterile Cytoplasm," Cell, 44:439-49 (1986); Wise et al., "Urf13-T of T Cytoplasm Maize Mitochondria Encodes a 13kD Polypeptide," Plant Mol. Biol., 9:121-26 (1987)). Abundance of CMS-associated proteins is also reduced in sunflower and radish (Horn et al., "A Mitochondrial 16 10 kDa Protein is Associated With Cytoplasmic Male Sterility in Sunflower," Plant Mol. Biol., 17:29-36 (1991); Laver et al., "Mitochondrial Genome Organization and Expression Associated With Cytoplasmic Male Sterility in Sunflower (*Helianthus annuus*)," Plant J., 1:185-193 (1991); Krishnasamy et al., "Organ-Specific Reduction in the Abundance of a Mitochondrial Protein Accompanies Fertility Restoration in 15 Cytoplasmic Male-Sterile Radish," Plant Mol. Biol., 26:935-946 (1994)). The mechanism behind the reduction in quantity of CMS-associated proteins in restored lines is not understood. For example, absence of transcripts that could potentially encode the PCF protein is not the explanation; only the shortest transcript is reduced in restored lines (Pruitt et al., "Transcription of the *Petunia* Mitochondrial CMS- 20 Associated *pcf* Locus in Male Sterile and Fertility-Restored Lines," Mol. Gen. Genet., 227:348-355 (1991)).

[0013] In *Petunia* and in some other CMS/restorer systems, the abnormal gene is co-transcribed with known mitochondrial genes. One possible mechanism for CMS in *Petunia* and its restoration, which is also consistent with current data, is that the 25 restorer gene not only results in decrease in the expression of PCF, but also improves the expression of the co-transcribed genes *nad3* and *rps12* in some way. For example, it remains possible that an RNA processing event results in little translation of PCF but enhanced production of NAD3 and RPS12 protein.

[0014] In sum, with the exception of maize *Rf2*, in those systems where 30 analysis has reached the molecular level, restorer genes have been found to affect the abundance of mitochondrial-encoded DNAs, RNAs, and proteins.

[0015] Cytoplasmic male sterility/restorer systems have been proven to be an invaluable tool in the production of hybrid seeds. Despite their importance for both

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the production of major crops such as rice and sunflower and the study of organelle/nuclear interactions in plants, none of the nuclear fertility-restorer genes that reduce the expression of aberrant mitochondrial proteins have been cloned.

[0016] The present invention is directed to overcoming these deficiencies in  
5 the art.

## SUMMARY OF THE INVENTION

[0017] The present invention relates to an isolated nucleic acid molecule which restores fertility to cytoplasmic male sterile plants and modifies expression of  
10 toxic mitochondria proteins by the plant. The nucleic acid molecule encodes a protein having an amino acid sequence of SEQ ID NOs: 2, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, or 41. Alternatively, the nucleic acid molecule encodes a protein containing a motif having an amino acid sequence corresponding to any of SEQ ID NOs: 3 to 18 or an amino acid sequence identified with a METAMEME software using the amino  
15 acid sequence of SEQ ID NO: 2 as input or an amino acid sequence identified as significantly similar to SEQ ID NO: 2 using a NCBI BLAST software (threshold= E less than or equal to 15) with SEQ ID NO: 2 as input. Alternatively, the nucleic acid molecule hybridizes to a nucleotide sequence of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ  
20 ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, or SEQ ID NO: 40 under stringent conditions of a hybridization buffer containing 20% formamide in 0.9M saline/0.09M SSC buffer at a temperature of 42°C. Alternatively, the nucleic acid molecule has a nucleotide sequence of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ  
25 ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, or SEQ ID NO: 40.

[0018] Another aspect of the present invention relates to a method of identifying a candidate plant suitable for breeding with a cytoplasmic male sterile plant. The method involves analyzing the candidate plant for the presence, in its genome, of the above nucleic acid molecule of the present invention.

[0019] Yet another aspect of the present invention relates to a method of identifying a candidate gene restoring fertility in plants. The method involves analyzing the candidate gene for the presence of the above nucleic acid molecule in accordance with the present invention.

5 [0020] The present invention also relates to a method of producing hybrid plant seed. The method first involves providing a cytoplasmic male sterile plant. Next, a second plant containing the above nucleic acid molecule in accordance with the present invention is provided. Finally, the cytoplasmic male sterile plant and the second plant are bred under conditions effective to produce hybrid progeny seed  
10 which yield fertile plants.

[0021] Another aspect of the present invention relates to a method of producing plant seeds for an inbred line of plants. The method first involves providing a cytoplasmic male sterile plant. Next, a second plant containing the above nucleic acid molecule in accordance with the present invention is provided. Then, the cytoplasmic male sterile plant and the second plant are bred under conditions effective to produce hybrid progeny seed which yield fertile plants. Next, hybrid fertile plants are produced from the hybrid progeny seeds. Finally, the hybrid fertile plants and the second plant are backcrossed to produce seed which yield inbred progeny plants.  
15

[0022] Yet another aspect of the present invention relates to a method of directing gene expression to plant mitochondria. The method involves transforming a plant with a chimeric nucleic acid molecule containing a transgene operatively linked to a promoter or a terminator from a plant gene which restores fertility to cytoplasmic male sterile plants and modifies expression of toxic mitochondria proteins by the plant under conditions effective to direct expression of the transgene in the mitochondria of  
20 the transformed plant. The promoter has a nucleotide sequence of from nucleotide 1 to 1981 of SEQ ID NO: 1. The terminator has a nucleotide sequence of from nucleotide 3761 to 4593 of SEQ ID NO: 1.  
25

[0023] The present invention also relates to a promoter from a plant gene which restores fertility to cytoplasmic male sterile plants and modifies expression of  
30 toxic mitochondria proteins by the plant under conditions effective to direct expression of the transgene in the mitochondria of the transformed plant. The promoter has a nucleotide sequence of from nucleotide 1 to 1981 of SEQ ID NO: 1.

[0024] Another aspect of the present invention relates to a terminator from a plant gene which restores fertility to cytoplasmic male sterile plants and modifies expression of toxic mitochondria proteins by the plant under conditions effective to direct expression of the transgene in the mitochondria of the transformed plant. The 5 terminator has a nucleotide sequence of from nucleotide 3761 to 4593 of SEQ ID NO: 1.

[0025] Yet another aspect of the present invention relates to a nucleic acid construct. The nucleic acid construct includes: (i) a promoter or a terminator from a plant gene which restores fertility to cytoplasmic male sterile plants and modifies expression of toxic mitochondria proteins by the plant under conditions effective to direct expression of the transgene in the mitochondria of the transformed plant and (ii) 10 a nucleic acid heterologous to and operatively coupled to the promoter or the terminator. The promoter has a nucleotide sequence of from nucleotide 1 to 1981 of SEQ ID NO: 1. The terminator has a nucleotide sequence of from nucleotide 3761 to 15 4593 of SEQ ID NO: 1.

[0026] The present invention also relates to a method of expressing a gene preferentially in roots of a plant. The method involves transforming a plant with a nucleic acid construct. The nucleic acid construct includes a promoter suitable for driving expression preferentially in roots having a nucleotide sequence of from 1 to 20 1388 of SEQ ID NO: 44; a nucleic acid heterologous to the promoter, where the promoter is operatively coupled 5' to the nucleic acid to induce transcription of the nucleic acid; and a terminator having a nucleotide sequence of from nucleotide 3168 to 4016 of SEQ ID NO: 44, where the terminator is operably coupled 3' to the nucleic acid.

[0027] Another aspect of the present invention relates to a method of altering 25 plant floral morphology in ornamental plants. The method involves transforming an ornamental plant with the above nucleic acid molecule in accordance with the present invention.

[0028] Another aspect of the present invention relates to a method of producing 30 plants with a cytoplasmic male sterile plant restoration system. The method first involves transforming a first plant in its chloroplast genome with a nucleic acid which causes the plant to become male sterile. Next a second plant is transformed with the above nucleic acid molecule in accordance with the present

invention whose protein product is targeted to the chloroplast. Finally, the first and second plants are crossed to produce progeny plants possessing a cytoplasmic male sterile plant restoration system.

[0029] Another aspect of the present invention relates to a method of  
5 producing plants with a cytoplasmic male sterile plant restoration system. The  
method first involves mutagenizing a first plant having a nucleic acid which encodes a  
protein. The protein has a motif having an amino acid sequence corresponding to any  
of SEQ ID NOs: 3 to 18 or an amino acid sequence identified with a METAMEME  
software using the amino acid sequence of SEQ ID NO: 2 as input or an amino acid  
10 sequence identified as significantly similar to SEQ ID NO: 2 using a NCBI BLAST  
software (threshold= E less than or equal to 15) with SEQ ID NO: 2 as input. Next,  
the mutagenized first plant is crossed with a wild-type plant having mitochondrial  
DNA polymorphisms compared to mitochondrial DNA in the mutagenized first plant  
to produce progeny plants. Finally, it is determined if the progeny plants are fertile,  
15 whereby fertile progeny plants can be used as a fertile maintainer line, where the  
mutagenized first plant, the fertile maintainer line, and a wild-type allele present in  
the first plant before mutagenesis comprises a new cytoplasmic male sterile plant  
restoration system.

[0030] The present invention also relates to an isolated nucleic acid sequence  
20 corresponding to SEQ ID NO: 42 or SEQ ID NO: 44.

[0031] The present invention identifies nucleic acid sequences which encode  
the gene for restoration of fertility to cytoplasmic male sterile plants. This gene  
modifies the expression of the mitochondrial genome and is the first such gene  
sequence that has been identified. In petunia, the gene may be transferred to lines  
25 lacking the gene in order to restore fertility. More importantly, the gene sequence has  
characteristics that can be used to identify comparable genes from economically  
important species. Thus, the gene and the sequence information may be used to  
develop hybrid seed production systems in economically important plants.  
Furthermore, the information may be used in crop improvement by controlling  
30 mitochondrial gene expression.

**BRIEF DESCRIPTION OF THE DRAWINGS**

- [0032] Figures 1A-D show that the *Rf* locus contains two tandem mitochondrially targeted PPR motif genes. Figure 1A illustrates the genomic organization of the region containing the *Rf-PPR592* and *Rf-PPR591* genes.
- 5 Duplicated blocks are indicated by similar shading. Arrows indicate the direction of transcription. 1 and 2 show locations of the primers used to amplify the *rf-PPR592* gene from a CMS plant. Figure 1B shows a single onion epidermal cell expressing a known mitochondrially targeted green fluorescent protein (GFP) after DNA bombardment. Figure 1C shows a single onion epidermal cell transiently expressing
- 10 44 N-terminal amino acids of *Rf-PPR592* fused to GFP. Figure 1D shows a comparison of PPR motifs found in *Rf-PPR592* with the MEME-derived consensus from 1,303 PPR motifs. The 14 PPR repeats are sorted by decreasing statistical significance, with PPR 230-264 showing the highest match to the consensus motif that is generated by retaining only the amino acids that occur at least in 6 of the
- 15 14 repeats.
- [0033] Figures 2A-B show the genetic structure of the *rf-PPR592* gene. Figure 2A illustrates that a comparison of *Rf-PPR592* and *rf-PPR592* reveals a size polymorphism. The first lane was loaded with the *Rf-PPR592* PCR amplicon obtained from a restorer line (*Rf/Rf*), the adjacent lane was loaded with the *rf-PPR592* PCR amplicon obtained with the same primer pair from a CMS line (*rf/rf*). Figure 2B illustrates that a comparison of *Rf-PPR592*, *Rf-PPR591*, and *rf-PPR592* reveals five similarity blocks. For each block, (I to V), the two blocks that exhibit the greatest similarity are shown with the same shading. Overall all three sequences are greater than 90% identical at the nucleotide level except in block V, where *Rf-PPR591* exhibits only 23% identity to the other two genes. The locations of 47- and 49-nt deletions in *Rf-PPR591* and 47- and 530-nt deletions in *rf-PPR592* with respect to the *Rf-PPR592* sequence in blocks I and II are shown as lines.
- 20 25 [0034] Figures 3A-B show the expression pattern of *rf-PPR592* and *Rf-PPR592*. Figure 3A depicts the examination of floral bud RNA for expression of *rf-PPR592* and *Rf-PPR592*. RT-PCR of floral bud RNA of a CMS plant (S) with primers specific to *rf-PPR592*, and RT-PCR of floral bud RNA of an *Rf/Rf* (nontransgenic) fertile plant with primers specific for *Rf-PPR592* (R). DNA, positive

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control for the amplification where the substrate is leaf DNA from a CMS plant; M, mass markers; 0, no template added, negative control. Figure 3B depicts the examination of different tissues for expression of *rf-PPR592*. RT-PCR of RNA from different tissues of a CMS plant with primers specific to *rf-PPR592*. DNA, M, and

5 0 are same as in Figure 3A.

[0035] Figures 4A-D illustrate the restoration of fertility to CMS *Petunia* lines by transformation with a 4.6-kb genomic sequence carrying *Rf-PPR592*. Figure 4A shows the flower of *P. parodii* CMS line 3688. Figure 4B shows the regenerant carrying *Rf-PPR592*. Figure 4C shows the *P. hybrida* CMS line 2423. Figure 4D shows the regenerant carrying *Rf-PPR592*.

[0036] Figures 5A-B illustrate the cosegregation of the *Rf-PPR592* transgene, restoration of fertility, and reduction of PCF. Figure 5A shows the DNA blot hybridized with an *npt* II transgene-specific probe. Lane 1, *P. parodii* CMS line 3688; lanes 2-and 3, sterile T<sub>1</sub> progeny of transformed *P. parodii*; lanes 4-9, fertile T<sub>1</sub> progeny. Figure 5B shows the immunoblot of floral bud proteins probed with anti-PCF antibody. Lanes are as in Figure 5A.

[0037] Figures 6A-B illustrate abnormal flowers on plants obtained by introducing *Rf-PPR592* into a CMS background. Figure 6A shows a petaloid flower on a plant carrying a recombination event near the *Rf* locus, affecting the region 5' to 20 *Rf-PPR592*. Figure 6B shows an abnormal flower on a plant carrying the CMS cytoplasm and the 4.5 kb *Rf-PPR592* transgene.

[0038] Figures 7A-B show methods for creating a new CMS/restorer system.

[0039] Figure 8 shows a two-line method for hybrid rice production, using an engineered inducible restorer gene.

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## DETAILED DESCRIPTION OF THE INVENTION

[0040] The present invention relates to an isolated nucleic acid molecule which restores fertility to cytoplasmic male sterile plants and modifies expression of toxic mitochondria proteins by the plant.

30 [0041] One form of the nucleic acid molecule of the present invention is a nucleotide sequence of from nucleotide 1982 to 3760 of SEQ ID NO: 1, identified herein as *Rf-PPR592*, as follows:

1 ATATATATACAAACTGATTTCTGTCTATTGCACAGTGTATTTACATACCCCTGAAAAGGGTAGCTCCGCT  
 81 AATAATGTTATCTTACAAAAAACAATAATTTTACATAATATACAAAACCTATTGTTATGTATTGAAATAT  
 161 GATAAAAATATTGTTATTTGTAATATAGCTATTAGGTAGTCATGTGGTGAATATTCCTAAATATTACCTGAG  
 241 TCGGCCATTGGCTAAAATATTTATTTAGTCGATACCTCAAGCTGTATATCCCATAGCAGCTACACTA  
 5 321 TAGCATATCTCTATTACCTTTAGTATTGTAACCCCCAATAGTATACAAGTTGACACCGCAATAGTGTACC  
 401 CCAATGTTGTTGTTGCTATAAAATGTTAAACAAAATTGAGTGATGGTAGTGAATTTAGTGTAAAGCTGGG  
 481 TAGTTTAAACATTCTTTGAAATTGAGTCAGTCATAGTACAAAAAACTGAAATATTATGTTCTGATT  
 561 TTGGCTGGTCTCTAAAATTTGAAATGCTGGTAGTTCACTTAGCGAGGGCAAATAGACTACATGCCAAATT  
 641 TAGTTAAAAGAAGTGTGCTGGAAAGTATTGAAAGATTGACAGTGTGCTAGACAACACGTCAAATT  
 10 721 TGTAGAAAATCTAGGAAGAAATTCAAAAGCAAATATTGCTTAAGCAGTCAGTCAGTCATGCTGCCTAGGGAG  
 801 TGAGAAATGGGCATCACTATAAGATTGATTTCATTGATATTTCATTATAAACTTAAGGAAAAGTGCAGGAAA  
 881 GCCACTTTGGCTTGCCTTACCGTGAAGCTACTTCAAAGAAAAGAGCTAGTTTAGCTTTGAACTTTAAT  
 961 CATTGTGGGCCGAACCTCAGACCTTGTGGCCGAACCTCATACTACATTACAAGTAAAAATTAGTCACAGGCCACTTTA  
 1041 CCACTAGTATTGGTTGAAGTCATTTTATTGGTTACATGAGAGACCACCTTTGAACTTCATCTTGTGCGC  
 15 1121 TTGAACCTCATGCCTAAGTTAAAGTTCAACTCAATCCGTAAGGGCTGAATTAGGCATAGATGCGTAAACCTCAA  
 1201 CCTTGTGGACTGAAGTTGAACCTCGCCCTTATGGTGGCCTGAAGTTGAACCTCAATCCTGTGGCTGAACCTGTGTA  
 1281 AGTTCAACCCACAAGGATTAAAGTTCAAAATGACCTCTCAAGCAAATCTGCAAAAAAAAGTGGTCTCATGCACT  
 1361 TTTACCCATTCCAAGTAGGCTGAAGTCAGTCCAGCCCACAATTATCAAGTTCCAAAAATTTCACAATATACCTCTTA  
 1441 TCTCGGTTATGATCTTGTATGATTAGCAGGAAATGGACGGGGAAAGTGCACGAAAGACCACTTTGCCATTGGCTTTG  
 20 1521 GGTACAGGCCACTAACCAAAATTAGTTAGTTGTTGCTACTTTGCTTAAAGAGATAGAACCTCAGTCCAGAGGCCGGA  
 1601 TTGAAGTTCAGTCCTTAAAGATTGAACTTCGATCCAGTGCCATATGGACTGAAGTCAGTCAGTCAGTCCTTAAAGATGGAAC  
 1681 TCAGTCCAGGCCATGGACTGAAGTTCAATCCTAAAGATAGAACCTCAGTCCAGGGCCGTGGACTGAAGTTCAG  
 1761 TCAATTATCAGAACTTAAGTCAGTATTAGTAAAGGCCAAAGTGGTAGTATAAGACCAATAAAATAGAGGCC  
 1841 TAAAACAAATAACAGTGTAAAAGTGGCTGATGGACGAAATTCTACAAAATGGACTCGAGGTAGCAATTCAACTTCAA  
 25 1921 CCTATGGTGTAGTCGTACAATTCTCCAATCACCCCTACTAAGTGAAGTGAAGCGAAGATGATGAGAAATTGCACTGC  
 2001 GTTACTGTCATGGTAATCCCTTTCTCATTGCTTCAATTGACCCCGACATTATTCTACCAATACATGT  
 2081 TCCATTCAGTTAAAGGAATTGGGGTTCTAATGAATTGAGAATGTTAGTGTAGTGTGCTTCAAGTGTGTT  
 2161 CCGTCAAATGGTTACAACCTGGCTCTGCTCTTCTAAATTGTTGAAGCTTGGTACATATGAAGC  
 2241 ATTACTCTCTGTTCTATTGAGAATCCACAAATTACGTTACGTTGATGCTTGCCTGAGCACTGTG  
 30 2321 GTTAACAGTTGTTGCCCTATGCATCGTACCGATCTCGGATTCTGTATTAGCCATTCACTCAAGAAAGGTATTCCATA  
 2401 TAATGAAGTCACCTTACTACCTAATAAGGGACTTTGCTGAAAATAAGGTCAAAGATGCTGTTCAATTGTTCAA  
 2481 AGTTGGTGAGGGAGAATATATGTGAGCCTGATGAGTCAGTGTAGGGACGGTCATGGATGGCTTGCAAGAAGGCCAT  
 2561 ACTCAAAAGCTTTGATTGCTCCGGTAATGGAACAAGGAATTACTAACGGCGATACATGCATCTACAAACATTGTTAT  
 2641 CGATGCCCTTGCAAAGATGGGATGGCTAGATGGTGTACCGCTTGAACGAGATGAAACAAAAACATTCTCCAG  
 35 2721 ACATTATTACATACCTCATTGATCGATGGTTGGGTAAGTTAGTCAGTGGAAAAGGTTAGGACTTTGTTCTGAG  
 2801 ATGATACATCTTAATATTGATCCAGATGTGTCACCTCAACTCCGTATTGATGGACTATGCAAAGAGGGAAAGTGA  
 2881 AGATGCCGAGGAATAATGACATACATGATCGAAAAGGTGTAAGACCTAACTGAGATAACCTACAATGTTGAATGGAT  
 2961 GATATTGCTTGCCTGGTCAAATGGTAGAGCGAGGGAGAATTGGATTCATGAGATAAGGGCATTGAGCCTGATATC  
 3041 ATTAGCTATACCGCACTAATAAAAGGATACGTCGAGAAAAGGAAATTGGATAAGGCCATGCAATTGTTCTGAAATTTC  
 40 3121 TCAAAATGGATTGAAACCTAGTATTGTTACCTGCAGTGTCTCTGGTGGCTTTGAAAGTGGAGAAACTGAATGTT  
 3201 CAAAAATATTCTTGATGAGATGCAAGCTGGGGCACATACCTAATTATACACTCATTGCACTTGTGTTGGTTAT  
 3281 TTTAAGAATGGACTTGTGAGAGGGCTATGTCACACTCCATAAGTGGAAAAGGAGGAGAGAAGATACAAATATTCAAAT  
 3361 TTACACGGCTGTCATTAAAGGATGTGCAAAATGGTAAGCTCGACAAAGCTCATGCTACGTTGAGAAGCTTCCCTGA  
 3441 TAGGCTTACATCCTGATGTGATAACACACTGCAATGATTAGTGGATATTGTCAGAAGGGTTGTTAGATGAAAGCTAA  
 45 3521 GATATGCTAAGGAAATGGAGGACAATGGTTGTTGCCAGACAACCGAACATACAATGTTATTGCGGGGATTTCAG  
 3601 AAGCAGTAAAGTTAGTGAAGGCTTCTGAGGAAATAGCTGGGAAGAGCTTCTCATTGAGGCACTGTGAG  
 3681 AGTTATTGATGGATATTAGCAGAGGATCCTTGTCTTCAACATGATTCCAGAATTTCACCGGGATAATAAGAAGTGA

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3761 ATAACTTTGCACCTGTTTTTGACGATATCACCATTATTCTGCTATTCCTTCATCTAGCAAAAGAAATTGCATC
3841 CAGTGGAAATTGCGGAAGCTGAAAAAATGGCAAGAAGAACATTGCTTAAGCTTCCTGGCAAGCTTATATCGGAGGGACAT
3921 CATTTGGGTGTTGGCTCTCTCTTATCTGGAAATCAAATGTTCTGCGCTCTTAATATCAGAAACAATGTGAACCT
4001 CCATATATGTACGAGTTATAAGTTCGGAATATGATTCAATGGTTCACTATTCTATTTGATATGAAATTAATTTT
5 4081 GAGCGACCCAGTGTTGACCATTGCCAACCTCGGTTATTATATGATTGAAATTCCCTCCAATCTCCAATACTCACTTCAT
4161 TTTGTCTGTTGAATTTCATTTCTTTCTGTTACGATTGTCACTTCACCGCTTGAGTATCCATCAGGTTCCA
4241 GTTGAAGAAAGAATCATTTTGCATGACCATCATGCTTCTGAGTGCAAGATCAAGAGAGGTACTTTCTCTCAAGAA
4321 CCTCTGGTTTTAAGTGTCTGGGTCTTCAGTACTTTAAGCTATTTCTAATCCTTGAAGAGATTACATACATAT
4401 CTGTGCACTGTGTTGTTCTTTCTGGGTGATACTTGTGTTAGCTAAGGATTGAAAAGTAATTCATTTCAT
10 4481 TAGCAATAGATATGAAACAGCTTGTAAAGGACTCTGGAGTCCTAAAAATTGGCTATGCAAATAGCCTATTGCATCA
4561 ATTTGTCGTTGAATCCATGTATCATAAAAAAA

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*Rf-PPR592*, isolated from *Petunia* has an open reading frame (“ORF”) of 1779 bp, extending between nucleotides 1982-3760.

15 [0042] The nucleic acid molecule of the present invention which has the nucleotide sequence of from nucleotide 1982 to 3760 of SEQ ID NO: 1 encodes a protein or polypeptide having a deduced amino acid sequence of SEQ ID NO: 2, as follows:

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20 MMRIAVRYCLNGNPFFSFFAYSIAPRHYSTNTCSISVKGNFGVSNEFENVKCLDDAFSLFRQMVTTKPLPSAVSFS
KLLKALVHMKHYSSVVSIFREIHKLRIPVDAFALSTVVNSCLMHRTDLGFSVLAIHFKKGIPYNEVTFTTLIRGL
FAENKVKDADVHLFKKLVRENICEPDEVMYGTVMMDGLCKKGHTQKAFDLLRLMEQGITKPDTCIYNIVIDAFCKDGM
LDGATSLLNEMKQKNIPPDIITYTSILDGLGKLSQWEKVRTLPLEMIHLNIYPDVCTFNSVIDGLCKEGKVEDAEE
IMTYMIEKGVEPNEITYNVMDGYCLRQMRARRIFDSMIDKGIEPDIISYTALINGYVEKKKMDKAMQLFREIS
25 QNGLKPSIVTCVSLRLGLEVGRTECAKIFFDEMQAAGHIPNLYTHCTLLGGYFKNGLVEEAMSHFKLERREDT
NIQIYTAVINGLCKNGKLDKAHATFEKLPLIGLHPDVITYTAMISGYCQEGLLDEAKDMLRKMEDNGCLPDNRRTYN
VIVRGFFRSSKVSEMKAFLKEIAGKSFSFEAATVELLMDDIAEDPSLLNMPEFHRDNKK

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[0043] As shown in Figure 1D, most of the predicted mature protein (87%) of Rf-PPR592 consists of 14 pentatricopeptide repeat motifs (PPRs). These repeats extend from the amino acid in position 54 to the amino acid in position 544 and are organized in two sets of tandem repeats, one set containing 3 PPRs from amino acid 54 to amino acid 158, the other set containing 11 PPRs from amino acid 160 to amino acid 544. Thus, another suitable nucleic acid molecule in accordance with the present invention encodes a protein containing a motif having an amino acid sequence corresponding to any of the PPR motifs (SEQ ID NOS: 3 to 18), where SEQ ID NO: 3 is as follows:

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E E A . . L Y . . M . . . G . . P N . . T Y N A L I N A Y A K . G . .

where SEQ ID NO: 4 is as follows:

5 D . A . . . F . . M . . . G . . P D . . T Y . . L I . G L C K . G . .

where SEQ ID NO: 5 is as follows:

D D A F S L F R Q M V T T K P L P S A V S F S K L L K A L V H M K H Y  
10

where SEQ ID NO: 6 is as follows:

S S V V S I F R E I H K L R I P V D A F A L S T V V N S C C L M H R T

15 where SEQ ID NO: 7 is as follows:

D L G F S V L A I H F K K G I P Y N E V T F T T L I R G L F A E N K V

where SEQ ID NO: 8 is as follows:

20 D A V H L F K K L V R E N I C E P D E V M Y G T V M D G L C K K G H T

where SEQ ID NO: 9 is as follows:

25 Q K A F D L L R L M E Q G I T K P D T C I Y N I V I D A F C K D G M L

where SEQ ID NO: 10 is as follows:

D G A T S L L N E M K Q K N I P P D I I T Y T S L I D G L G K L S Q W  
30

where SEQ ID NO: 11 is as follows:

E K V R T L F L E M I H L N I Y P D V C T F N S V I D G L C K E G K V

35 where SEQ ID NO: 12 is as follows:

E D A E E I M T Y M I E K G V E P N E I T Y N V V M D G Y C L R G Q M

- 17 -

where SEQ ID NO: 13 is as follows:

G R A R R I F D S M I D K G I E P D I I S Y T A L I N G Y V E K K K M

5

where SEQ ID NO: 14 is as follows:

D K A M Q L F R E I S Q N G L K P S I V T C S V L L R G L F E V G R T

10 where SEQ ID NO: 15 is as follows:

E C A K I F F D E M Q A A G H I P N L Y T H C T L L G G Y F K N G L V

where SEQ ID NO: 16 is as follows:

15

E E A M S H F H K L E R R R E D T N I Q I Y T A V I N G L C K N G K L

where SEQ ID NO: 17 is as follows:

20

D K A H A T F E K L P L I G L H P D V I T Y T A M I S G Y C Q E G L L

and where SEQ ID NO: 18 is as follows:

D E A K D M L R K M E D N G C L P D N R T Y N V I V R G F F R S S K V

25

A PPR motif-containing gene can be identified if it contains the consensus sequence (SEQ ID NOs: 3 or 4) or if it is found with a MEME software (Bailey et al., "Fitting a Mixture Model by Expectation Maximization to Discover Motifs in Biopolymers," Proceedings of the Second International Conference on Intelligent Systems for Molecular Biology, pp. 28-36, AAAI Press, Menlo Park, California (1994), which is hereby incorporated by reference in its entirety). To find whether a protein has a PPR motif with the MEME software, the parameters for motif searching should be set as minimum width=35, maximum width=35. MEME (Multiple Em for Motif Elicitation) is a software tool for discovering motifs in a group of related DNA or protein sequences (Bailey et al., "Fitting a Mixture Model by Expectation Maximization to Discover Motifs in Biopolymers," Proceedings of the Second

30

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International Conference on Intelligent Systems for Molecular Biology, pp. 28-36, AAAI Press, Menlo Park, California (1994), which is hereby incorporated by reference in its entirety). MEME takes as input a group of DNA or protein sequences (the “training set”) and outputs as many motifs as requested. MEME uses statistical modeling techniques to automatically choose the best width, number of occurrences, and description for each motif. MEME represents motifs as position-dependent letter-probability matrices which describe the probability of each possible letter at each position in the pattern. Individual MEME motifs do not contain gaps. Patterns with variable-length gaps are split by MEME into two or more separate motifs.

10 [0044] Another suitable nucleic acid molecule for the present invention is a nucleic acid molecule which encodes a protein having an amino acid sequence identified with a METAMEME software using the amino acid sequence of SEQ ID NO: 2 as input. Meta-MEME is a software toolkit for building and using motif-based hidden Markov models of DNA and proteins. The input to Meta-MEME is a set of similar protein sequences, as well as a set of motif models discovered by MEME. Meta-MEME combines these models into a single, motif-based hidden Markov model and uses this model to produce a multiple alignment of the original set of sequences and to search a sequence database for homologs (Grundy et al., “Meta-MEME: Motif-based Hidden Markov Models of Biological Sequences,” Computer Applications in the Biosciences, 13(4):397-406 (1997), which is hereby incorporated by reference in its entirety).

15 [0045] Also suitable for the present invention is a nucleic acid molecule which encodes a protein having an amino acid sequence identified as significantly similar to SEQ ID NO: 2 using a NCBI BLAST software (threshold= E less than or equal to 15) with SEQ ID NO: 2 as input for comparison (Fulton et al., “Identification, Analysis, and Utilization of Conserved Ortholog Set Markers for Comparative Genomics in Higher Plants,” Plant Cell, 14:1457-1467 (2002), which is hereby incorporated by reference in its entirety).

20 [0046] Also suitable in the present invention is a nucleic acid molecule which has a nucleotide sequence of SEQ ID NO: 19, an 8.5 kb fragment containing *Rf-PPR592* capable of transforming cytoplasmic male sterile plants, as follows:

- 19 -

GGATCCAAAATTCACTAAAGGTTAACCGCGAGGAACTGAAGTGGAGAGCAATGTGGTATCTGGTCATGGAC  
GGAGTCATGGGGTATAGTGCTGGCTATTTAGCTAAGTGGAGACTGTGTGAGGAATTATTTAAAGAACATGCTAC  
TTTCTCGTCCACATAAACATGTCAAATATTTCTACTGTGATAGAGAGTCAAGGAAATAGTGTGGATTCTCCC  
AAACACAAACGATTGAGAAAAGTGAAGTGAAGGCTGAAGAGAAAAGTAAAGAGAACTGGAGCTAAGAACACAG  
5 AAGCACAAACCTATAAACATAGACACTGGCATGTCAGAAAATTAACTTCGATTCTCAGTAGAAAGACACA  
AATACATCAGTAAATTCTTAGGCTCAAGCAAGGATACTCTGGTAGAATTGCATTATACCAACATAATAAG  
CTCAAAAAAAATAACTAAGCTGCAACTAGCTACTTGGTCCGAGAAGCTTGTATCAGGAAGTCCACAGTCCAA  
AACCAAGGTAGACTATCAAGCTAGTCCCACCAGTCATTTCTTAGACTTGTCTCACGATAAACTAAGATCATT  
TTTTATGATACATGGATTCAACGACAAATTGATGCAATAGGCTATTGCATAGCCAAAATTAGGAGACTCC  
10 AGAGTCCTTACAAAGCTGTTCATATCTATTGCTAATGAAAATTACCTTTCAATCCTAGCTATAAAC  
AAAGTATCACCGAAAAAAAGAAACACATGCACAGATATGTATGAATCTCTCAAAGGATTAGAAAATAGCT  
TAAAAGTACTGAAAGAACCCAGAACACTAAAAACCAAGAGGTTCTAGAGAGAAAAGTACCTCTTGATCTG  
CACTCAGAAAGCATGATGGTCATGGCAAAATGATTCTTCAACTGGAACCTGATGGACTCAAGGCAGTGA  
AAATGACAATCGTAACAGAAAAAGAAAATTGAAAATTCAACAAGACAAAATGAAGTGAATGGAGATTGGAG  
15 GGAATTCAATCATATAAAACGAAGGTAGGCAATGGCAACACTGGTCGCTCAAAATTAAATTCCATATCAA  
AATAGAATACTGAAACCATTGAAATCATATTCCGAAACTTATAACTCGTACATATGGGAGTTCACATTGTTCT  
GATATTAAGAGCGCAGAACATTGATTCCAAGATAAGAAGAGAGGCCAAACACCAAAATGATGTCCTCCGAT  
ATAAGCTTGCAGGAAAGCTTAAGCAATGTTCTTGCATTTTCACTGGCAATTCCACTGGATGCAATT  
TCTTGTCAAGATGAAAGGAAATAGCAGAATAATGGTGTATCGTCAAAAAACAGGTGCAAAAGTTATTCACT  
20 TCTTATTATCCGGTGAATTCTGGAATCATGTTAAGCAAAGAAGGATCCTCTGCTATAATATCCATCAATAACTC  
TACAGTAGCTGCCTCAAATGAGAAGCTTCCCAGCTATTCTTCAGAAAAGCCTTCATTCACACTTACTG  
CTTCTGAAAATCCCCACAATAACATTGTATGTCGGTTGTCTGGCAACACCCATTGTCCTCCATTCTTCA  
GCATATCTTAGCTCATCTAACACCTCTTGACAATATCCACTAATCATTGAGTGTATGTTATCACATCAGG  
ATGTAAGCCTATCAAGGGAAAGCTCTCAAACGTAGCATGAGCTTGTGAGCTTACCAATTGGCACAATCCATTA  
25 ATGACAGCGTGTAAATTGAATATTGTATCTTCTCTCCCTTCAACTTATGGAAGTGTGACATAGCCTCTT  
CAACAAGTCATTCTAAAATAACCAAGCAAAGTCAATGAGTGTATAAATTAGGTATGTCCCCCGAGCTG  
CATCTCATCAAAGAATATTTCGACATTCACTGAGCTTCAACTTCAAAAGACCACGCAAGAGAACACTGCAGGTA  
ACAATACTAGGTTCAATCCATTGAGAAATTTCACGAAACAATTGCAATGCCCTATCCATTTCCTTCG  
CGTATCCATTATTAGTGCAGGTTAGCTAATGATATCAGGCTCAATGCCCTATCTATCATGGAATCAAATTCT  
30 CCTCGCTTACCCATTGACCACGCAAGCAATATCCATTACCAATTGTAGGTTATCTCATTAGGTTCTACA  
CCTTTTCGATCATGTATGTCATTATTCCTCGGCATCTCAACTTCCCTCTTGCAAGTCCATCAATGACGG  
AGTGAAGGTGCACACATCTGGATAAATATTAGATGTATCATCTCAAGGAACAAAGCTCAACCTTCCACTG  
ACTTAACCTACCAAACCATCGATCAATGAGGTATATGTAATAATGTCGGAGGAATGTTTTGTTCTACCTCG  
TTCAAAAGGCTGGTAGCAGCATCTAGCATCCCATTGCAAAAGGCATCGATAACAATGTTGAGATGCATGTAT  
35 CGGGCTTAGTAATTCTGTTCCATTAACCGGAGCAATCAAAGCTTTGAGTATGCCCTCTGCAAGCCC  
ATCCATGACCGTCCCACATGACTTCATCAGGCTCACATATATTCTCCCTACCAACTTTGAAACAAATGAACA  
GCATCTTGACCTTATTCAGCAAAAGTCCCTTATTAAGGTAGTAAAGGTGACTTCATTATGGAATACCTT  
TCTTGAAGTGAATGGCTAATACAGAAAATCCGAGATCGGTACGATGCATAAGGCAACAATGTTAACCAAGCT  
CAAGGCAGAACATCAACAGGAATCGTAATTGTCGGATTCTGCAAAAGTAAACACAGAAGAGTAATGCTTC  
40 ATATGTACCAAGCTTCAACAATTAGAGAAAGAGACAGCAGAAGGAAGAGGCTTAGTTGTAACCATTGACGGA  
ACAAACTGAAAGCATCATCTAACACACTAACATTCTCAAATTCAATTAGAAACCCAAAATTCCCTTAACGAAAT  
GGAACATGTATTGGTAGAATAATGTCGGGTGCAATTGAATAAGCAAAGAATGAGAAAAAGGGATTACATTGAGA  
CAGTAACGCACTGCAATTCTCATCATTCGCTTCACTTAGTAGGGTGATTGGAAGAATTGTACGACTAT  
GACACCATAGGTTGAAGTGAATTGCTACCTCGAGTCCATTGAGAAATTGTCATCAGCCACTTTAACAC

- 20 -

TGTTATTTAGTTAGGCCTCTATTTTATTGGCTTATACTAACCACTTTGGGCCCTTACTAAATAAAACTGA  
 CTTAAGTTCTGATAATTGACTGAACCTCAGTCATACGGCCCTGGACTGAAGTTCTATCTTAAGGATTGAACCT  
 CAGTCCCATATGGCTCTGGACTGAAGTCCATCTTAAGGACTTGACTGAACCTCAGTCATATGGCACTGGATCGAA  
 GTTCAATCTTAAGGACTGAACCTCAATCCGGCTCTGGACTGAAGTTCTATCTCTTAAGCAAAGTAGGCCACAA  
 5 ACTAAATATTTGGTATTAGTGGCCTGTACCCAAAGACCAATGGCAAAAGTGGTCTTCGTGACTTCCCCGTCC  
 ATTTGCTAAATCATAACAAAGATCATAACCGAGATAAGGAGGTATATATTGTGAAATTGGTGAACCTGAATAA  
 TTGTGGGCTGAACCTCAGCCTACTTGCATGGTAAAGTGCATGAGAGACCCTTTTGCAAGATTTGCT  
 TGAGAGGTCACTTTGAAACTTAAATCCTGTGGTGAACCTCACACAAGTCAGCCCACAAGGATTGAAGTTC  
 AACTCAGGCCACCAAAAGGGCGAAGTTCAACTCAGTCCACAAGGTTGAAGTTACGCATCTATGCCTAAAAAT  
 10 TCAGCCCTTACGGATTGAAGTTGAACCTAAACTTAGGCATGAAGTTCAAGCGCACAAAGATTGAAGTTCCAAA  
 AGTGGTCTCTCATGTAACCAATAAAAGATGACTCAAACCAAATACTAGTGGTAAAGTGGCTGTGAGCTAA  
 TTTTTACTTGTGAATGTATGAAGTTGGCCACAAGGTCTGAAGTTGGCCACAATGATTAAAGTTCCAAAAAA  
 GCTAAAAAAACTAGCTCTTTCTTGAAAGTAGCTCACCGTAAAGGCAAGGCCAAAAGTGGCTTCTGCAC  
 TTTCCCTTAAGTTATAATGAATAAAATATGAATGGAAATAACATCTTATAGTGTATGCCCTACACTCCCTAAG  
 15 GCAGCATTGTCTTGACTGCCTTGTAAAGCAATATTGCTTGAATTCTCCACATTTCTACATAATT  
 TGACGTGTTGCTAGGCAACACTGTCCGTACAATCTTCTGAATACTTCCCAGACAACACTTCTTTAACGTA  
 AAATTGGCCATGTAGTCTATTGCCCCCTCGCTAAATGAAAAGTGCAGCATTCAAAATTTAGAAGGACAGC  
 CAAAATCAAGAAACATATAATATTCAGTTGGTACTATGACTTGAACATAATTCAAAAGTGGTTAAACATTT  
 AAAACTACCCAGCTTACACTAAAAAATTACACTACCATCACTCAAATTGTTAAACATTTATAGCCACACAAC  
 20 CACAACATTGGGTACACTATTGGGTGTCAAACCTGTATACTATTGGGGGTATACGAATAACTAAAAAGTAAATA  
 ATAGAAGATATCGTATAGGTATACTGCGCTATGGATATAACAGCTTGGAGTATATGCGACTATAAAATAAA  
 TTTTAGCCAATGGCGACTCAGGTAATATTAGGAAATATTACACCACATGACTACCTAATAGCTATATTA  
 CAAAAATAACAATATTATCATATTACAATACATAACAATGAGTTGTATATTATGTAACCAAAAGTGGTT  
 GTTATTGGTAAAGATAACATTATTAGCGGAGCTACCCCTTTCAAGGGTATGTAATATAACACTGTGCAAAT  
 25 AGACAGAAAAATCAGTTGTATATATATCAGATATTGATCCCCCTCATTTCGTATGTTACTTTTA  
 TATTATATATCCCTTAGTAAAAACTGGCTCCGCCACTGCCAGTAAGGTAGTATTAGTTGGCTCGCTCAATAA  
 AGTAACATCTACGTTATTTCATCAACATTAAAAGGAAGATTCACTATCCACATAGGCATCATCATTATCAA  
 AGAATATCAGTCATACATTGTATATATAACTTCTCAAATAACTAATTAAAGTACATTAAAAGG  
 AAGATTCACTATCCTTTAATATTCTGTTATTACTTGTGACTCCTTAGTAAAAACTGGCTC  
 30 CGCCACTACCAGTGTAAATTAAATTGCGTCCCTCACTAAAGTAACACCTATAATTAAATTTCATGAAGTCA  
 GAGTTAGCATTGGAAAGGGATATAAGCACATGCATTGTGTATATATATAACTTGCTCAAATAAAACTAACTTAA  
 AATGAAATTTCATTTCCTAGTACAATGAACATGCTCAATGCGTAATTAGTTGAGGTGGCTATATGAATATG  
 TTATTAATTGAAAGCAAACATAAAACTGATAGAAGAATTGACCTAAAATTGAACTTGAGCTGCTTCAGT  
 TACTATCTCATTTCACTATATATGTGTATCAGCTAATTCTATGATTAATTAAACAAATTGTAAGTATTAAAC  
 35 AAAATAACGAATAAAATGGAAAATAAGTACTTGATGAACGTAGGGCCGGAGTTGGCCGAGGTGACCGGAGACAAT  
 GGAAAGCAGAGTTACTATTGGACTAAATGCCACAAAAGAATCATTGTTTACAATGTAGCAAGTTGGCACGA  
 TTATGATTCTGACACAATAGCCACATTATAGAAAGATAATGTGGCACTAATGAGGTAAATTTCATTATGGAATGA  
 TAACAAACAAATAAGTACACGATTAAAACAAACTGAAAGGGTTGCGCGATAATTAAATCATTGTTTACGAAAA  
 TACTCATCAAATCAAATATTATCTGCCCTGCATGTAAGTTCTATTTACTCGCTTGCCTAAATTATG  
 40 AAAAATTTACTCACACCGGATATATACCTAACCATAGCAATTAACTATGTAATGTGATGTAATGAAGAGAGA  
 ATGTCTATTAATTAAATTATGGCTAAGTGTAGTGTATTGTAACAAATGACGTGCAATTGTTGATTAGACACT  
 TACAAAAATACCCACGAAATCTAAATAATTACAGCCACTATCCACTACTTCAAATATTATCTGGCCTACCCATT  
 AAAATTTACTCACTACCCCTCCAGACTTATATTATAAGGTATAAAAGGTAAACAAATAATAATGGCCT  
 CCAGACTTTATACCATAATTGCAAGCCTAAAGGTACACCTATAACAAAGGTACACAAATAATAATGGGT

ATGGTGGGTAATACTTTAGTTTATGGGTAGAGTAATTTAATGGGTATGACTTGAAATACTTAAATTCA  
 TGGGTATAGAGTGTAAAATTCTTGTGATTGGGTATATACACCCGATGTGGGTAAAGTACTTGCTAATTTG  
 CCCTAAGGTAATAATAGACTTATAGTATAAGGAAATACGCAGTGTGATAGATACTTGAATTTCATGAGTAATG  
 TAATTTAATAGGTATAGTAAGGTAATACTTATATTCCAAGGGTACACATTGAAAGCTCAATATTTATT  
 5 CCAAAGAATAAGAGACCAAACAATGTGTTGAGATTATTACTTGTGTCACAAACTAAAAAGAAAACCTTAG  
 AAGTCTAAATTACAATACTTAACATGCATTTACGAATAAAATATCACAAACACTATTAGAGATAATGT  
 CGTGGATGATGTTAACATATTGACTACACAACCCATTGTACAATAATTGAAGCATGTATATGCACGACCAAGA  
 CTCCATCATCATAGATCAAATGAATGTTATTTAATGCATGAAACCTAAGTAGAACATTTATGCCCTAATGAAC  
 AAAACCAAGCAAAAGATACTACTTGTGCAATTGAATGAATTCTACCGTATATACTAATATACACCAGAGGTT  
 10 AGTTAACACTTGGAACTTCAAAAGGTGTACAACCAGAGTTCCCTACATTGATGGTTCTTCATTCACCA  
 ACTGATAAAATGAAGGCTGGTATAGTCTACAAATCCCTAGTCCCTGTGAACTTGCATCCCTCTAGCTACATGC  
 AGAACATGTCCTTAGATCCCAGGTGTATTGCCATTGCCACTGAACATGGAGGACAATGTATAATTGTCCTC  
 CTCACCCATTGCACATACTCTGTCATTGCTGCACATCTACATGCCCTTCTGAATATTCTCTGAGTCACAAAG  
 CATCATGAGACATCTGCAAGTATTTGAATGGGATAATCACATTCCAAATCGAAAGGTTCTGTCTTAACAAAG  
 15 TCAAGCTGCATCTGACAAAGAGACTCGTTGATGAAATGCCACATAAGAGCACATGCAAACCAAGTTGTTAAA  
 GCATTGTACACATATAACCTACTTCTGATGTGAATAAAAGAAAGTCGATCAATGACAGAGGAAACAGTCACATCTAT  
 AACACAAAAAAATCTTTCTTAAAGTACGACCAACATAGATAACTATAATTCCCGTAGATGTCAAACACTCTTATT  
 GAATAAAAATAAACAAACATGTACTATTGCTCATTATCCGACTGTACAAGGATTTCTTAATGATGGTATAA  
 TAGGAGCAATCCCTTATGACAGATGCACTAATTGTTGGGTGCATATTCAATGCAAGACTGTGGGTATATA  
 20 ATCTAAAATATCATTCAAATCAAACCTGGGACGATTGAGAGAAGATTAGCATGGCCTCTGCACAAGGATGACAG  
 CATAAATCGAGAAATGTTCAAATAAGGAAATATATATATTACCTGTTCAATTGCCATAGTTCTAAAGAAGTT  
 TTGGCAGTTAAAGTATTAATAGTTACCTGTTGATTGCTGACAGGTTAGCCTGGGGTGTCTGGGACGGACCTG  
 TGATTATTCTGCTAATCTCCTGTATATTGCAATGTGCAGTTAATCCAGTGCACCTTGCCTGTTATGGATGG  
 ATCC

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**[0047]** Another suitable nucleic acid molecule in accordance with the present invention is isolated from rice and identified herein as *Rhpr1*, which has a nucleotide sequence of SEQ ID NO: 20, as follows:

30 ATGGCGCGCCCGTCCCTACCCGCCCGCGCGGGTGGCGGGCGGCGTCCCACGCTCGGAGGGCTCGATCCAAG  
 GGCGAGGAGGCCGCGGGGGCCAGTGGCGCCGAGGACGCACGCCACGTGTTGACCAATTGCTCCGGCGTGGCAG  
 GGGCGCCTCGATCTACGGCTGAAACCGCCCTCGCCGACGTGCGCGTACAGCCCCGCGGCCCGTGTCCC  
 TACAACCGCATGGCCCGAGCCGGCGCCGCAAGGTAACCTCCACCGTGCACACCTATGCCATCCTCATCGGCTGCT  
 GCTGCCGTGCGGGCCGCTTGGACCTCGGTTCGCGGCCCTGGCAATGTCGTCAAGAAGGGATTAGAGTGGATGC  
 35 CATCACCTTCACTCCTGCTCAAGGCCCTGTGCGACAAGAGGACGAGCGACGCAATGGACATAGTGCTCC  
 AGAATGACCGAGCTCGGCTGCATACCAAGATGTCTCTCCTACAATAATCTTCTCAAGGGTCTGTGATGAGAAC  
 GAAGCCAAGAAGCTCTCGAGCTGCTGCACATGATGGCTGATGATCGAGGAGGAGGTAGCCCACCTGATGTGGTGC  
 GTATAACACTGTCCTCAATGGCTCTCAAAGAGGGGATTGACACAAAGCTTACAGTACATACCATGAAATGCTG  
 GACCGGGGGATTTACCAAGATGTTGACCTACAGCTCTATTATTGCTGCCATTGCAAGGCTCAAGCTATGGACA  
 40 AAGCCATGGAGGTACTTAAACACCATGTTAAGAATGGTGTGATGCCTGACATATAATAGTATTCTGCA  
 TGGATATTGCTCTTCAGGGCAGCCAAAGAGGCTATTGGAACACTCAAAAGATGCGCAGTGTGGCGTCGAACCA  
 AATGTTGTTACTTATAGTCAGTGAATTATCTTGCAAGAATGGAAGATCCACCGAAGCTAGAAAGATTTCG  
 ATTCTATGACCAAGAGGGCCCTAGAGCCTGATATTGCTACCTATCGTACCCCTGCTCAGGGGTATGCTACCAAAGG

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AGCCCTGTTGAGATGCATGCTCTGGATTGATGGTACCAAATGGTATCCAACCGGATCATCATGTATTCAAC
ATTCTAATATGTGCATACGCTAAACAAGAGAAAGTAGATCAGGCAATGCTTGTATTCAAGAAAATGAGGCAGCATG
GATTGAATCCGAATGTAGTGTGCTATGGAACAGTATAGATGACTTCAAGTCAGGCAGTGAGATGATGCTAT
GCTTATTTGAGCAGATGATCGATGAAGGACTAACCCCTAACATTATTGTGTATAACCTCCCTAACATGGCTG
5   TGCACCTGTGACAAATGGGACAAGGCTGAAGAGTTAACATTCTGAAATGTGGATCGAGGCATCTGTCTGAACACTA
TTTCTTAATTCAATAATTGACAGTCATTGCAAAGAAGGGAGGGTATAGAATCTGAAAAACTCTTGACTTGAT
GGTACGAATTGGTGTGAAGCCGATATCATTACGTACAATACTCATCGATGGATGCTGCTAGCTGGTAAGATG
GATGAAGCAACGAAGTTACTTGCCAGCATGGTCTAGTTGGGTGAAACCTGATATTGTTACCTATGGCACCTTGA
TTAATGGCTACTGTAGAGTTAGCAGGATGGATGACGCATTAGCTTTCAAAGAGATGGTGGAGCAGTGGTGTAG
10  TCCTAATATTATTACGTATAACATAATTCTGAAAGGTTATTACATACCAGAAGAACTGCTGCTGCAAAAGAACTC
TATGTCAGTATTACCAAAAGTGGAACACAGCTGAACTTAGCACGTACAACATAATCCTTCATGGACTTGC
15  ACAATCTCACTGACGAGGCACTTCGAATGTTAGCAGGATTTACAGCTGGAGACTAGGACTTT
TAACATTATGATTGGTGCCTTACTTAAATGTGGAAGAATGGATGAAGCTAAGGATTGTTGCTGCTCACTCGGCT
AACCGTTAGTGCAGATGTTAGGACCTACAGTTAATGGCAGAAATCTTATAGACCAGGGTCGCTAGAAGAAT
20  TGGATGATCTATTCTTCATGGAGGAGAATGGCTGTTCCGCCACTCCGCATGCTAAATTCCATTGTTAGGAA
ACTGTTACAGAGGGGTGATATAACCAGGGCTGGCACTTACCTGTTCATGATTGATGAGAACGACTTCCCTCGAA
GCATCCACTGCTTCCCTCTGTTAGAATCTCCCCAATCGCTGGAGCAAATATCAAGAATATCACACTTGCTG
TAAATTGAAATTAAATTAAAGCAGCCAAATGCACCTGTGAGTTAGGCCAAAGTGGTCCAAATCTGCCTAAACC
25  TGGCACAAATTGGTCGGTAGTGTGCGCACAGTTCACTTATCGCGCGGGTTATCGCCTTACCGCGGGGTACG
ACGGTTACCGCACTACCGCAGGGTGACGGTAACCCGGCCAAACGATAAGGTAAACCTGGTCGACAAATTGG
CCCACCGACCAGTTATCGCCTACCGCGGGATGCCTCAGTAGGACCTAG

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*Rhpr1* is a rice homolog of the *Petunia Rf-PPR592* gene.

[0048] The nucleic acid molecule of the present invention which has the  
25 nucleotide sequence of SEQ ID NO: 20 encodes a protein or polypeptide having a deduced amino acid sequence corresponding to SEQ ID NO: 21 as follows:

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MARRVPTRPRGGGGGGVPRSEGSIQGRGGRAGGSGAEDARHVFDELLRRGRGASIYGLNRALADVARHSPAAVSR
YNRMARAGAGKVTPTVHTYAILIGCCCRAGRDLGFAALGNVVKKGFRVDAITFTPLLKGLCADKRTSDAMDIVLR
30 RMTELGCIPDVFSYNNLLKGLCDENRSQEAELELLHMMADDRGGSPPDVSYNTVLNGFFKEGDSDKAYSTYHEML
DRGILPDVVTYSSIIAALCKAQAMDKAMEVLNTMVKNVGMPDCMTYNSILHGYCSSQPKEAIGTLKKMRSDGVEP
NVVTYSSLNYLCKNGRSTEARKIFDSMTKRGLEPDIATYRTLLQGYATKGALVEMHALLDLMVRNGIQPDHHVFN
IILICAYAKQEVDQAMLVFSKMRQHGLNPNVVCYGTIDVLCKSGSVDDAMLYFEQMIDEGLTPNIIVYTSLIHGL
CTCDKWDKAELILEMLDRGICLNТИFFNSIIDSHCKEGRVIESEKLFDFLMVRIGVKPDIITYNTLIDGCCLAGKM
35 DEATKLLASMVSVGVKPDIVTYGTLINGYCRVSRMDDALALFKEMVSSGVSPNIITYNIILQGLFHTRRTAAAKEL
YVSITKSGTQLELSTYNIILHGLCKNNLTDEALRMFQNLCSDLQLETRTFNIMIGALLKCGRMDEAKDLFAAHSA
NGLVPDVRTYSLMAENLIEQGSLEELDDLFLSMEENGCSADSRMLNSIVRKLLQRGDITRAGTYLFMIDEKHFSL
ASTASFLLESSPIVWEQISRISHLSVNLKLKQPKCTCELGPKWSQNLPKPGTNVGVAQFHLRSRGGYRAYRGGT
TVTALPQGDGNPGPNKDVKNPGRTNLAQNRPVIALPRDASVGP

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**[0049]** Another suitable nucleic acid molecule in accordance with the present invention is isolated from rice and identified herein as *Rhpr2*, which has a nucleotide sequence of SEQ ID NO: 22, as follows:

5 ATGGCGCGCCGCCGCGCTTCCC CGTCCGC CGCGCTGTGGCGCCCTCGCTCGGAGGGCTCGACCCAAGGGC  
GAGGGGGCGCAGGGGGCGAGTGGCGCCGAGGACGCACGCCACGTGTTGACGAATTGCTCCGGCGTGGCAGGGG  
CGCCTCGATCTACGGCTTGAACTGCCCTCGCCACGTCGCCGTACAGCCCCGCCGCCGTGCTCCGCTAC  
AACCGCATGGCCGAGCCCGCGACGAGGTAACCTCCAACCTGTGACCTACGGCATTCTCATCGGTTCTGCT  
GCTGCGCGGGCCGCTTGGACCTCGGTTCGCGCCCTGGCAATGTCATTAAGAAGGGATTAGAGTGGACGCCAT  
10 CGCCTCACTCCTCTGCTCAAGGGCCTCTGTGCTGACAAGAGGACGAGCGACGCAATGGACATAGTGCTCCGAGA  
ATGACCCAGCTGGCTGCATAACAAATGTCCTCTACAATATTCTCTCAAGGGCTGTGTGATGAGAACAGAA  
GCCAAGAAGCTCTCGAGCTGCTCAAATGATGCCGTGATGATGGAGGTGACTGCCACCTGATGTGGTGTGCTATAC  
CACTGTCATCAATGGCTCTCAAGGAGGGATCTGGACAAAGCTTACGGTACATACCATGAAATGCTGGACCCG  
GGGATTTACCAAATGTTTACCTACAGCTCTATTATGCTGCGTTATGCAAGGCTCAAGCTATGGACAAAGCCA  
15 TGGAGGTACTTACCAAGCATGGTAAGAATGGTGTGATGCCATTGCAAGGACGTATAATAGTATCGTCATGGTA  
TTGCTCTCAGGGCAGCGAAAGAGGCATTGGATTTCTCAAAGGATGACAGTGTGATGGTGTGCAACCAGATGTT  
GTTACTTATAACTCGCTCATGGATTATCTTGCAAGAACGGAAGATGACCGGAAGCTAGAAAGATGTTGCTATTCTA  
TGACCAAGAGGGCCTAAAGCCTGAAATTACTACCTATGGTACCCCTGCTCAGGGTATGCTACCAAGGAGCCCT  
TGTGAGATGCATGGCTTGGATTGATGGTACGAAACGTTACCCCTAATCATTATGTTTACGATTCTA  
20 ATATGTCATACGCTAAACAAGGGAAAGTAGATCAGGCAATGCTGTGTTGCAAGGAGCTATGCTATGCTTA  
ATCCGGATACTGACCTATGGAACAGTTAGGCATACTTGCAAGTCAGGAGAGTAGAAGATGCTATGCTTA  
TTTGAGCAGATGATGAAAGACTAAGCCCTGGCAACATTGTTATACTCCCTAATTGATGCTCTGTATC  
TTTGACAAATGGGACAAGGCTAAAGAGTTAATTCTGAAATGTTGGATCGAGGCACTGCTGACACTATTTCT  
TTAATTCAATAATTGACAGTCATTGCAAGAAGGGAGGGTTATAGAATCTGAAAACCTTTGACCTGATGGTACG  
25 TATTGGTGTGAAGCCGATATCATTACGTACAGTACTCTCATCGATGGATATTGCTGGCAGGTAAAGATGGATGAA  
GCAACGAAGTTACTGCCAGCATGGCTCAGTGGAAATGAAACCTGATTGTTACATATAATACCTTGTAAATG  
GCTACTGTAATTAGCAGGATGAAAGATGCGTTAGTTCTTTAGGGAGATGGAGAGCAGTGGTGTAGCCTGA  
TATTATTACGTATAATATAATTCTGCAAGGTTATTCAAACCCAGAAGAACTGCTGTCGAAAGAAACTCTATGTC  
GGGATTACCGAAAGTGGACCGCTGAACTTAGCACATAACATAATCCTCATGGCTTGCAAAACAAATC  
30 TCACTGACGAGGCACCTCGAATGTTCAAGAACCTATGTTGACGGATTACAGCTGGAGACTAGGACTTTAACAT  
TATGATTGGTGCATTGCTTAAAGTTGGCAGAAATGATGAAGCCAAGGATTGTTGCAAGCTCTCGGCTAACGGT  
TTAGTGCCAGATGTTAGGACCTACAGTTAATGGCAGAAAATCTTATAGAGCAGGGTTGCTAGAAGAATTGGATG  
ATCTATTCTTCAATGGAGGAGAATGGCTGACTGCCAATCCCGCATGCTAAATTCCATTGTTAGGAAACTGTT  
ACAGAGGGGTGATATAACCAGGGCTGGCACTTACCTGTTCATGATTGATGAGAAGCACTTCTCCCTGAAAGCATCC  
35 ACTGCTTCCATTGTTAGATCTTGTCTGGGGAAAATCAAGAATATCATAGTGTATTAGAGGAGGGATCT  
TCTCTTATGTTAAATAGCAGGTTCAAGAAAATCTTGTGATTGAGAATCTGGTGTCCATTCTTCTTAA  
ATTATTAAATCCTCCAGTGAATCTGTTGATTCCAAGCACCACATGATAGGTTCAAACCTCTTGAATCAGTAAA  
GTTCAAATGCTTAATGGATCAAATAAGGATTCTGACTGCATTGAGGAAATCCTTCAAAGTTGAAGAGATTC  
TCTTAAGCTGTCAGTGAATGCTGACGCCAGAACAGATGACAAGAAAACAAGGCCAGAACTGTCGAAAGTG  
40 GCTGCTTGTGACAATGGAAAATGCATGCTGTCTGCTGTTGAGTGGAGAGACTTCTGACACAGTGTCCAGA  
GTTGGAGGAAATTAAAGAGACATTAAGGGAGATGGGAGGTCTGATAGTATTTGACGTTATGGTGGATTTC  
ATTCAACATTGGAGAATCTCATAAAGGATACATCCACTTCAGCTTGGACCGAAATGAAGGAACATTTGCAAAG  
TGCTGCTCCCTTGAATGTTGAAAATATTGAAATGCCATATTCTAAGCGATGATAACAAGACCCATTG

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CTTAATATGAGTAGAAAATTGAACCGAAACGCTCCCTGCTTCTTTGGTGTCAATTATCAATACTATTGAGT  
 TATTATCAGCTTTCAATACTTCAGAATTCTCTGTTGTTCCAGCTCACATATCCGAAATCGTCTAAAGTCTC  
 TCAACAGAGTTACTCTGTGGTGATGGGGGGGGCGACCCTGGCGAGGCGTGGAGTGCCTACCGCATCAGGGTGA  
 TCGGCCGCGCTGCTCCGCCCTGGTCCGCAGGCTTGGCGCGAGCTGGCGGGAGGGAGACTGTGGTGAGATCGG  
 5 ATTCGCCGCTGGTGTGCTACCATGGGGATTGCCGCAGGCGCTCAGATCGTTATGCCGGAGCGAAC  
 AAAAGTATGGCGTGGCGCGCGAGTGGACGCCAGGCCTCGCGGAATGGGGCTGGGGACCGAGCCAGTCT  
 CGCTTGCCGGTAACCGGAACCGAGCTCAGCACTACATTGCAAAGATTGGCAACTCTGACAATTCCATGTTCT  
 ACAAGCTTGACGTCGAGGGAAATGGAGAACCTGCCACCGAATAGTAGCCCTGCTATCTATGTTGCGAACCATCAGAG  
 TTTTTGGATATCTATACCCCTCTAACTCTAGGAAGGTGTTCAAGTTATAAGCAAGACAAGTATATTATGTT  
 10 CGAATTATTTGA

*Rhpr2* is a rice homolog of the *Petunia Rf-PPR592* gene.

[0050] The nucleic acid molecule of the present invention which has the nucleotide sequence of SEQ ID NO: 22 encodes a protein or polypeptide having a deduced amino acid sequence corresponding to SEQ ID NO: 23 as follows:

MARRAASRVAGAVGALRSEGSTQGRGGRTGGSGAEDARHVFDELLRRRGASIYGLNCALADVARHSPAAVSRY  
 NRMARAGADEVTPNLCTYGILIGSCCCAGRDLGFAALGNVIKKGFRVDAIAFTPLKGLCADKRTSDAMDIVLRR  
 MTQLGCIPNVFSYNILLKGLCDENRSQEALELLQMMFDGGDCPPDVSVTTVINGFFKEGDLDKAYGYHEMLDR  
 20 GILPNVVITYSSIITAALCKAQAMDKAMEVLTSMVKNGVMPNCRTYNSIVHGYCSSGQPKEAIGFLKKMHSDGVEPDV  
 VTYNSLMDYLCKNGRCTEARKMFDSMTKRLKPEITTYGTLQGYATKGALVEMHGLLDMVRNGIHPNHYVFSIL  
 ICAYAKQGKVDQAMLVFSKMRQQGLNPDTVTYGTIVGILCKSGRVEDAMRYFEQMIIDERLSPGNIVYNSLIHSLCI  
 FDKWDKAELILEMLDRGICLDTIFFNSIIDSHCKEGRVIESEKLFIDLVRIGVKPDIIITYSTLIDGYCLAGKMD  
 ATKLLASMVSVGMPDCVTYNTLINGCKISRMEALVLFRMESSGVSPDIITYNIILQGLFQTRRTAAKELYV  
 25 GITESGTQLELSTYNIILHGLCKNNLTDEALRMFQNLCLTDLQLETRTFNIMIGALLKVRNDEAKDLFAALSANG  
 LVPDVRTYSLMAENLIEQGLLEELDDFLSMEENGCTANSRMLNSTVRKLLQRGDITRAGTYLFMIDEKHFSLEAS  
 TASLFLDLLSGGKYQEYHSCIRGGIFSLCVNSEVQENHLLDSESGVHFLKLLNPVNLDKAPSIGSKLLGISK  
 VQMLNGSNKSDCISEEILSKVEEILSCQVIKSLDKDDKTTTRPELCPKWALLTMENA CLSAVSVETSDTVSR  
 VGGNFKETLREMGGLDSIFDVMVDFHSTLENLIKDTSTSALDRNEGTSLSQAALLKCLKILENAIFLSDDNKTHL  
 30 LNMSRKLNPKRSLLSFVGVIINTIELLSALSILQNSSVSSSTYPKSSKVSQQSY SVMAGGDRGRGVECHPHQGV  
 SAALLRPGPQALAASWRRRETVRSDFAAGGVATMGDPQALSDRLCGSATKVWRGGAETAEAFARNGAAGPSQS  
 RLPVTRNRAQHYIAKIWATLTISMFYKLDVEGMENLPPNSSPAIYVANHQSFLDIYTLLTGRCFKFISKTSIFMF  
 RII

[0051] Another suitable nucleic acid molecule in accordance with the present invention is isolated from rice and identified herein as *Rhpr3*, which has a nucleotide sequence of SEQ ID NO: 24, as follows:

40 ATGGCGCGCCGCGCCGCTTCCCGCGCTGTTGGCGCCCTCGCTCGACGGCTCGATCCAAGGGCGAGGAGGCCGCG  
 CGGGGGCAGTGGCGCCGAGGACGCACGCCACGTGTCGACGAATTGCTCCGGCGTGGCAGGGCGCCTCGATCTA

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CGGCTTGAACCGCGCCCTCGCCGACGTCGCGCGTACAGCCCCGGCCCGTGTCCCCTACAACCGCATGGCC
CGAGCTGGCGCCGACGAGGTAACCTCCGACTTGTGCACCTACGGCATTCTCATCGGTTGCTGCTGCCGCGCGGGCC
GCTTGGACCTCGGTTCGGGCCCTGGGCAATGTCAATAAGAAGGGATTAGAGTGGAAGCCATCACCTTCACCTCC
TCTGCTCAAGGGCCTCTGTGCCGACAAGAGGACGAGCGACGCAATGGACATAGTGCTCCGCAGAACGACCGAGCTC
5 GGTTGCATACCAAATGTCTCTCCTACAATAATCTTCTCAACGGGCTGTGTGATGAGAACAGAACAGCAAGAAGCTC
TCGAGTTGCTGCACATGATGGCTGATGATCGAGGAGGAGTAGGCCACCTGATGTGGTGTGATACCTGCTACATGGCATT
CAATGGCTTCTCAAAGAGGGGATTACAGACAAAGCTTACAGTACATACCATGAAATGCTGGACCGGGGGATTTA
CCTGATGTTGTGACCTACAGCTCTATTATTGCTGGTTATCCAAGGGTCAAGCTATGGGACAAGCCATGGAGTCATT
GCAAAGAAGGGAGGGTTATAGAACATCTGAAAAACTCTTGACCTGATGGTACGTATTGGTGTGAAGCCTGATATCAT
10 TACATACAGTACACTCATCGATGGATTGCTGGCAGGTAAAGATGGATGAAGCAATGAAGTTACTTCTGGCATG
GTCTCAGTTGGGTTGAAACCTAATACTGTTACTTATAGCACTTGATTAATGGCTACTGCCAAAATTAGTAGGATGG
AACACGCGTTAGTTCTTTAAGGAGATGGAGAGCAGTGGTGTAGTCCTGATATTATTACGTATAACATAATTCT
GCAAGGTTATTCAAACACAGAACAGAACGACTGCTGCTGCAAAAGAACACTATGTCAGGATTACCGAACAGTGGAAACGCAG
ATTGAACCTAGCACATACACATAATCCTTCATGGACATTGCAAAACAAACTCACTGATGATGCACTTCAGATGT
15 TTCAGAACCTATGTTGATGGATTGAAAGCTTGAGGCTAGGACTTCAACATTATGATTGATGCATTGCTTAAAGT
TGGCAGAAATGATGAAGCCAAGGATTGTTGCTTCCTCGTCTAACGGTTAGTGGCAATTATTGGACGTAC
AGGGTATGGCTGAAAATTATAGGACAGGGTTGCTAGAAGAATTGGATCAACTCTTCTTCAATGGAGGACA
ATGGCTGTACTGTTGACTCTGGCATGCTAAATTTCATTGTTAGGGAACTGTTGCAGAGAGGAGTAGTGGTGGTGGT
GAGTGGTGAATCTGCCACCCCCACCAACTCTCAAAATTCTGACATGTGGGATCACTGTCAATCCCTCTCC
20 AAGACATGTGGATCACTGTCATCCCTCTCCAAACCAATTGTGCAGACAGGTGCTGGTCAAGGTTAAAGAAG
TTGGCAAAATGCTTCTGAAGAAAGGTTATTGTTGCTTCATCTCAGGAGATTCCAGATGATCCAGTGTCTCCAAC
AATTGAGGCGCTTATTTGCTCCATAGTAAAGCAAGTACACTTGCTGAGAACACCAGGTGACAACACGGTTGTT
GTACCATCAAACAAAGTGGTTGATTCTGGGAAAGGTGAAAGTAATTACTGAAATGAGAACACGGACTGGGG
CTGAAATCCGAGTCACTCAAAAGCAGATAAACCTAAGTACCTGTTGATGAGGAGCTTGTGCACTGATATCAG
25 CCTTATCTTGGTTGATCGGCATGCTGGACGAGCACATCTGTTGTCATCAACTGCTGACTGCTATATATGTGCTG
GTGCTGAATCGATCGATTGTCGTCGCGGAAGTGAAGAACACCAACGGCAGTGTGCTGGCTCTAGCCGCCA
TCAGTTATAACCGTACAAACTCAGTGAATTGCTGGTTACATTGGTTATAATAAAAGGCCTCCGTTAGTT
CAGCCTGGGCTTCAGAATCTCAGGACCGGCCCTGCTCATGATCCTTACACCGTGTATCCTGTAGAGTACTTCTCT
AAAAGAGAGTACCCTAGTGGAACTAGCAAAGTTGCACCATCTGCTTACAGAACATGCAACTACTCGCT
30 TGCCATAATGGAGAACTGCCCTCATCTATTAGTCCTGGTGCCTGATTATGCTCTGCCGTTCTATCTGACCAAGT
ACCTACTGATAGGTACTCTAATAGGTTACACTACAATTAGGCCTCTGAGAGGCCGAAATAGTAATGTGCAACAA
TTAGGAATCACCAGAGCTGAAATTCCAATGCTTATGATTACTGAGGCTGCTGAGCACATGGACGTGAGG
ATTACCGAAGACTGTCAGGTCTACTGGGTATCCAGGTGGCTTCAAGTGGATTCCAATAGTTAACCTGGAG
TCTGTCATTGGTGTGGTGTCTGGTGCAGAGGTGAAGTTGCACGAAGCCCACAGAGCCTCTGCAAGGCTTCATCGCGCAAGCAGCA
35 GTGGAGATCCAGGGCATTCCGGATCAAGTGAAGGCCGACAGAGCCTCTGCAAGGCTTCATCGCGCAAGCAGCA
ACAGCAGGCAGGCAGGCCAGTCCTCTCGCATGGCCCATTATTTTAG

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*Rhpr3* is a rice homolog of the *Petunia Rf-PPR592* gene.

[0052] The nucleic acid molecule of the present invention which has the  
40 nucleotide sequence of SEQ ID NO: 24 encodes a protein or polypeptide having a deduced amino acid sequence corresponding to SEQ ID NO: 25 as follows:

- 26 -

MARRAASRAVGALRSDGSIQGRGGRAGGSGAEDARHVFDELLRRRGASIYGLNRALADVARHSPAAVSRYNRMA  
 RAGADEVTPLCTYGLILIGCCCRAGRLDLGFAALGNVIKKGFRVEAITFTPLLKGLCADKRTSDAMDIVLRRMTEL  
 GCTPNVFSYNLLNLCDENRSQEAELELLHMMADDRGGGSPPDVSYTTVINGFFKEGDSDKAYSTYHEMLDRGIL  
 PDVVTYSSIIAALCKGQAMDKPWSHCKEGRVIESEKLFDLVRIGVKPDIITYSTLIDGYCLAGKMDEAMKLLSGM  
 5 VSVGLKPNTVTYSTLINGCKISRMEDALVLFKEMESSGVSPDIITYNIILQGLFQTRRTAAKELYVRITESGTQ  
 IELSTYNIILHGLCKNKLTDALQMFQNLCIMDLKLEARTFNIMIDALLKVGRNDEAKDLFVAFSSNGLVPNYWTY  
 RLMAENIIQGQLLEELDQLFLSMEDNGCTVDGMLNFIVRELLQRGVVVVSGESATPPPTLKILTCGITVNPFS  
 KTCGITVNPFSKPIVQTGACGQVKVEGVKNASEERLIVVSSQEIPDDPVSPTIEALILLHSKASTLAENHQLTTRLV  
 VPSNKVGICLGEKKVITEMRRRTGAEIRVYSKADPKYLSFDEELVQHISLILVDRHAGRAHLLSHQLLTAIYVL  
 10 VLNRSIVVAEVKNNHGTAAWCWALAAISYNRTNFSDLVSHWIFIKAVSFSTLGLQNLRTGPAHDPTYTVYPVEYFS  
 KREYPSGSSSKVAPSASYERYAATTRLPNGELPSSISPGADYMSCRSYLDQVPTDRYSNRVTLQLGLSRAGNSNVQQ  
 LGITRAGNSNAYDYTEAAEQIHGREDYRRLSGLTGPYGGSSNCGFQIVNWSLSLVLVISGARVHLHEAPGSSEI  
 VEIQGIPDQVKAAQSLLQFIGASSNSRQAPQSSRMAHYF

- 15 [0053] Another suitable nucleic acid molecule in accordance with the present invention is isolated from rice and identified herein as *Rhpr4*, which has a nucleotide sequence of SEQ ID NO: 26, as follows:

ATGCCGCTCGCCACGCTGCTCGGCCACCTCGCCGCCGGCGCTTCGGCTCGTGCAGGCCTCACCGGCCGCCGA  
 20 CCGCGGGCGGCCGCACCGACTCCTCACCTCCTCCGCACAGCGCCGCCCTCCCGACCTCCCGTCCACGGGCTCCTCTCGCCGCCCTC  
 CCTCGCGGGCTGGTCGCGCGCCCACTCCGCGCCGCCCTCCGCTCCAGCTCCACGTCCCTGCCGCCGCGCCACTCCCCCGCAT  
 GCCTCCAAGGGGCTCTACCCCTCCTCCGCTCCAGCTCCACGTCCCTGCCGCCGCGCCACTCGCCGACATGCTCGTCC  
 CCATCCTCCGCGCTCTCCCGTCCCGTCCCGTCCGCTCCACGTCCCTGCCGCCGCGCCACTCGCCGACATGCTCGTCC  
 CGCCCTCGCCAGGGCATCCAGCCCTCAGGGCGTACGACCGGTTCTCTCGCCGGGGAGAGCCACCCGCCAC  
 25 CGCCCTCCACCTCCGTGAACGCCCTCTCGCCGCCCTCGCCGCCAACGGCTGACCTCGCCGAGAAGG  
 CGTCAGGAGCCGCTGCCGCCGCGTGTACCGGACATCTACACCTCAACACCGTCATCTCCGGCTCTGCAG  
 GATCGGCCAGCTCGCAAAGCCGGCATGTCGCCAAGGACATCAAGGCATGGGTCTGGCTCCCTCTGTGGCCACC  
 TACAATAGCCTCATCGATGGTACTGCAAGAAGGGTGGAGCTGGAACATGTACCATGTCGACATGCTTTGAAGG  
 AGATGGTCAAGCCGGATCTCACCGACTGCAATTGTTGATCAATGGGATTGCAAGAACACTCGAA  
 30 TACTGCGGCCAGTGAAGACTTCGAGGAGATGAAGCAGCAGGGATCGCTGCGAGTGTGACGTATAATTG  
 CTAATTTCAGGTCTCTGCAGTGAGGGTAAGGTGGAGGAAGGGGTGAAGCTGATGGAGGAGATGGAGGATTGGGGC  
 TGTCACCCAATGAAATCACCTTGGCTGTGTTCTGAAAGGGTTGTAAGAAGGGATGATGGCAGATGCCAATGA  
 TTGGATTGATGGTATGACAGAGGAATGTGGAACCTGATGTGTTATTACAATATCTTGATCGATGTGATCGC  
 CGTCTGGAAAAATGGAGGATGCAATGGCGTGAAGGAGGCAATGGCAAAGAAGGGATCAGTCCAATGTCACAA  
 35 CATATAATTGCTTGATAACAGGGTTAGCCGAGTGGGGATTGGAGGAGTGCTCTGGCCTCTGGATGAGATGAA  
 GGAGAAAGGTATTGAAGCAGACGTCGTCACTTACAATGTGCTTATTGGCTTGCTGCAAAGGTGAGGTACGG  
 AAAGCTGAAAGCTCTGGATGAAATGTCGGAAGTTGGATTGGAACCAAACCATCTGACCTACAAATACCATAATAC  
 AGGGGTTCTGTGATAAGGTAACATTAAGTCTGCCTATGAAATTAGAACCCAGGATGGAAAAATGTCGGAAACGGGC  
 AAATGTGGTTACGTACAATGTGTTCATCAAGTATTCTGCCAGATAGGGAAAGATGGATGAAAGCTAATGATCTACTC  
 40 AATGAGATGTTGGACAAATGTCTAGTCCAAACGGGATCACTTATGAAACGATAAAAGAGGGGATGATGGAAAAG  
 GCTATACACCAGATATTAGAGGGTGCAGTGTCTACAAGCTCTGAAAACCCAGCAGCATCCTGA

- 27 -

*Rhpr4* is a rice homolog of the *Petunia Rf-PPR592* gene.

[0054] The nucleic acid molecule of the present invention which has the nucleotide sequence of SEQ ID NO: 26 encodes a protein or polypeptide having a deduced amino acid sequence corresponding to SEQ ID NO: 27 as follows:

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MPLATLLGHLAAGRFGLVQALTGAATAAAAHRLLHLLLRTAPPPLPDLVSLARWSRAHFRAPLPLRHGLLLARL
ASKGLYPLLSELHVLAARLHSASIILRALPSPSASASASTPLIADMVLALARASQPLRAYDAFLLAGESHPRH
RPSTSSVNALLAGLVGAKRVDLAEKAFRSALRRVSPDIYTFNTVISGLCRIGQLRKAGDVAKDIKAWGLAPS VAT
YNSLIDGYCKKKGGAGNMYHVDMLLKEMVEAGISPTAVTGFVILINGYCKNSNTAAAVRVFEEMKQQGIAASVVTYNS
10 LISGLCSEGKVVEGVKLMEEMEDLGLSPNEITFGCVLKGFCKKGMADANDWIDGMTERNVEPDVVIYNILIDVYR
RLGKMEDAMAVKEAMAKKGISPNVTTYNCLITGFSRSGDWRSASGLLDEMKEKGIEADVVTYNVLIGALCCKGEVR
KAVKLLDEMSEVGLEPNHLTYNTIIQGFCDKGNIKSAYEIRTRMEKCRKRANVTVNVFIKYFCQIGKMDEANDLL
NEMLDKCLVPNGITYETIKEGMMEKGYPDIRGCTVSQASENPASS
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[0055] Another suitable nucleic acid molecule in accordance with the present invention is isolated from rice and identified herein as *Rhpr5*, which has a nucleotide sequence of SEQ ID NO: 28, as follows:

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ATGGCTGATGATGGTCGCTGCCACCTGATGTGGTGTGCGTATAATACCATTGATGGCTCTCTCAAAGAGGGTG
ATGTGGACAAAGCTTACATCACATACCATGAAATGCTGGACCCGGAGGGTTCTCCAGATGCTGTGACTTACAACTC
TATCATTGCTGCCCTAACAGCAAGGCTCAAGCTATGGACAGGGCCATGGAGGTACTTACAGTGATGGTTATGCCAAT
TGCTTCACATATAATAGTATTATGCATGGATATTGTTCTTCAGGACAGTCGGAAAAGGCTATTGGTATTTCAGAA
AGATGTGCAGTGATGGTATTGAACCAAGATGTTACTTATAACTCGTGATGGACTATCTCTGCAAGAACGGAAA
ATGCACAGAACGCCAGAAAGATTTGATTCTATGGTCAAGAGGGCTCAAGCCTGATATTACTACCTATGGTACC
25 CTGCTTCATGGGTATGCTTCAAAGGAGCTTGTGAGATGCATGATCTCTAGCTTGATGGTACAAAATGGCA
TGCAACTTGATCATCATGCTTCAACATATTAATATGTGCATACACTAAACAAGAAAAGTAGACGAGGTCGTGCT
TGTATTCAAGCAAAATGAGGCAGCAAGGATTGACTCCGAACGCAGTGAACATAGAACAGTGATAGATGGACTTTGC
AAGTTAGGTAGACTAGATGATGCTATGCTTAATTGAGCAGATGATTGATAAAGGACTGACACCTAACGTTGTTG
TTTATACCTCCCTAACATTGCTCTGTACCTATGACAAATGGAGAAGGCCGAGGAGTTAATTGGAAATATT
30 GGATCAAGGTATCAATCCCAACATTGTTTTTAATACAATATTGGACAGTCTTGCAAAAGAAGGGAGGGTTATA
GAATCTAAAAAAACTCTTGACCTGTTGGACATATTGGTGTGAATCCTGATGTCATTACATACAGTACACTCATCG
ATGGATATTGCTTAGCTGGTAAGATGGATGGAGCAATGAAGTTACTCACTGGCATGGCTCAGTTGGTTGAAACC
TGATAGTGTACATATAGCACTTGTAAATGGTTACTGTAATAGAACATGGAGGACGCATTAGCTTTTC
AAGGAGATGGAAAGCAATGGTGTAAATCCTGATATTACATATAACATAATTCTGCATGGTTATTCGCACCA
35 GAAGAACTGCTGCTGCAAAAGAACTATATGCCAGGATTACCGAAAGTGGAACGCAGCTGAACCTAGCACATACAA
CATATAACCTCATGGACTTGCACAAACAAACTCACTGATGATGCACTTCGGATGTTCAAGAACCTATGTTGA
```

*Rhpr5* is a rice homolog of the *Petunia Rf-PPR592* gene.

- 28 -

**[0056]** The nucleic acid molecule of the present invention which has the nucleotide sequence of SEQ ID NO: 28 encodes a protein or polypeptide having a deduced amino acid sequence corresponding to SEQ ID NO: 29 as follows:

5 MADDGRCPDPVSYNTIIDGLFKEGDVDKAYITYHEMLDRRVSPDAVTYNSIIAALSKAQAMDRA  
MEVLTVMVMPNCFTYNSIMHGYCSSGQSEKAIGIFRKMCSDGIEPDVVTYNSLMDYLCKNGKCTEAR  
KIFDMSVKRGLKPDIITTYGTLLHGYASKGALVEMHDLLALMVQNGMQLDHVFNLICAYTKQE  
KVDEVVLVFSKMRQQGLTPNAVNYRTVIDGLCKLGRLDDAMLNFEQMIDKG  
LTPNVVVYTSЛИHALCTYDKWEAEELIFEILDQGINPNIVFFNTILDSLC  
KEGRVI ESKKLFDLLGHIGVNPDVITYSTLIDGYCLAGKMDGAMKLLTGMVS  
VGLKPDSTVYSTLINGYCKINRMEDALALF  
10 KEMESNGVNPDIITYNIIHLGLFRTRTAAKELYARITESGTQLELSTYN  
IILMDFAKTNSLMMHFGCFRTYV

**[0057]** Another suitable nucleic acid molecule in accordance with the present invention is isolated from rice and identified herein as *Rhpr6*, which has a nucleotide sequence of SEQ ID NO: 30, as follows:

15 ATGGCGCGCCGCGCCGCTTCCCGCGCTGTTGGCTCGAGGGCTCGATCCAAGGGCGAGGGGCC  
GCGGGGGCGCGGGCA ATGGCGCCGAGGACGCACGCCACGTGTCACGAATTGCTTC  
GGCGTGGCAAGGGGCCACGATCTACGGCTTGAA  
CCGCGCCCTCGACGACGTCGCGCGTCAACAGCCCCGGCCGGCT  
ACAAACCGCATGGCCCGAGCCGGC  
GCCGACGAGGTAACTCCAACTTGTACACCTACAGCGTTCT  
CATCGGTTGCTGCTGCCGGCGGGCGCTTGGACC  
20 TCGGTTTCGCGCCCTGGCAATGTCATTAAGAAGGGATTAGAGTGG  
AGCCATCACCTTCACTCCTCTGCTCAA  
GGGCCTCTGTGCCGACAAGAGGACGAGCGACGCAATGGAC  
ATAGTGTCTGCAGAATGACCCAGCTGGCTGCATA  
CCAAATGTCTTCTCCTGCACCATTCTCAAGGGTCTGTG  
ATGAGAACAGAACAGCAAGAACGCTCTCGAGCTGC  
TCCAAATGATGCCGTGATGATGGAGGTGACTGCC  
ACCTGATGATGGCTGCTACATACCATGAAATGTTG  
ACCAGGGATTGCGAGATGTTG  
CAAAGAGGGGGATCCGGACAAAGCTTACGCTACATAC  
CATGAAATGTTGAGGCTACTTACACCGTCA  
ATCAATGGCTTCT  
CAAAGAGGGGGATCCGGACAAAGCTTACGCTACATAC  
CATGAAATGTTGAGGCTACTTACACCGTCA  
ATCAATGGCTTCT  
25 ACTTACAGCTCTATTATCGCTGCCTTATGCAAGGCT  
CAAGCTATGGACAAGGCCATGGAGGTACTTAACAC  
CATGGTTAAGAATGGTGT  
CATGCCATTGCA  
AGGACATATAATAGTATTGTC  
ACGGATATTGCTCTTCAGGGCAGTTG  
GAGATGCATGATCTCCT  
AGAGGCTATTGGATTCTCAAAATGATGTC  
AGTGTGCA  
GGCTTGTGAGGCTTGT  
GATGGCTGCA  
ACCGAGATGTTGTT  
ACTTGTA  
ACTTGCTGATG  
GATTATCTTGCAAGAACAGAAC  
GATGCACGG  
AACAGCTAGAAAGATT  
TTCAATTCTATG  
ACCAAGTGTGG  
CTAAAGC  
CTGATATTACTAC  
CTATTGTA  
CCCTGCTTCAGGGT  
ATGCTAC  
CAAAGGCC  
TTGTTGAG  
ATGCATG  
GATCTC  
30 GGATTTGATGGTATGGAACGGTATCCA  
ACCTAATCAT  
CATGATTCA  
ACATTCT  
AAATATG  
TCATAC  
GCTAAACAA  
GAAAAAGTAGATGAGGCGATGCTTG  
ATTCA  
GCAAAATGAGG  
CAGCAAGGATTGAG  
TCCGAATGC  
AGTGA  
ACTACA  
GAACAGTC  
ATAGATG  
TACTCTG  
CAAGCT  
AGGCAGAGT  
ACGATG  
CAGTGC  
ACTACA  
AGGACTAAC  
CCCTGACAT  
ATTG  
TATATAC  
CCCCCT  
AATTCA  
TGGTT  
TTGT  
ACCTGT  
GACAA  
ATGG  
GAGAAGG  
CTAAAGC  
GAGTGA  
ACTACA  
GAGGAG  
GTTA  
ATTTTAA

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*Rhpr6* is a rice homolog of the *Petunia Rf-PPR592* gene.

**[0058]** The nucleic acid molecule of the present invention which has the nucleotide sequence of SEQ ID NO: 30 encodes a protein or polypeptide having a deduced amino acid sequence corresponding to SEQ ID NO: 31 as follows:

- 29 -

MARRAASRAVGSEGSIQGRGGRAGGNGAEDARHVFDELLRRGKGATIYGLNRALDDVARHSPAAAVSRYNRMARAG  
ADEVTPNLYTYSVLIGCCCAGRDLGFAALGNVIKKGFRVEAITFTPLLKGLCADKRTSDAMDIVLCRMTQLGCIPNVFSCTILLKGLCDENRSQEALELLOQMPDDGGDCPPDVLYNTVINGFFKEGDPDKAYATYHEMFDQGILPDVV  
5 TYSSIIAALCKAQAMDKAMEVLNTMVNGVMPNCRTYNSIVHGYCSSGQLTEAIGFLKMMCSDGVEPDVVTCNLLMDYLCNRRCTEARKIFNSMTKCGLPDITTYCTLLQGYATKGALVEMHDLDDLMVWNGIQPNHHVFNILICAYAKQEKVDEAMLVFSKMRQQGLSPNAVNYRTVIDVLCKLGRVYDAVTLKQMINEGLTPDIIVYTPLIHGFCTCDKWEKA  
EELIF

- 10 [0059] Another suitable nucleic acid molecule in accordance with the present invention is isolated from rice and identified herein as *Rhpr7*, which has a nucleotide sequence of SEQ ID NO: 32, as follows:

ATGGCACGCCCGTCGCTGCCCGCGCCCCCGCGCCGGCGTCCCGCGCTCGGAGGGTACGATCCAAGACC  
15 GAGCACCGCGTTGGGAGCGGTGGCGCCGAGGACGCACTCGACGTGTTCGACGAATTGCTCCGGCGAGGCATCGCGCTCGCATCCGCAGCTTGAAACGGCGCTCTCGCCGACGTCGCGCGACAACCCCGGGCGCTGTGTCCCGCTTCAACCGCATGGCACGAGCTGGTCCAGCATTCCACACCTATGGCATCCTCATCGGCTGCTGCTGCA  
CGCATGGCACGAGCTGGTCCAGCATGGTAACTCCCACCGTGCACACCTATGGCATCCTCATCGGCTGCTGCTGCA  
GTGCGGGCCGCTTAGACCTCGGTTCCGGCCCTGGGCATGCGTTAGAAGGGATTCAAGAGTGGAAACCCATCAT  
CTTAATCCTCTGCTCAAGGGCCTGTGCAGACAAGAGGACGGACGACGAATGGACATAGTGTCCGTGGAATG  
20 ACCGAGCTCAGCTCGGTGCCAATGCTTCTCCCACACCATTATTCTCAAGGGACTCTGTCATGAGAACAGAACGCC  
AAGAACGCTCTCGAGCTGCTCCACATGATGGCTGATGGAGGAGGCTGTTACCTAATGTTGTCATACAGCAC  
CGTCATCGATGCCCTTGAAAGGAGGGATCCGGACAAAGCCTACGCTACATACCGTAAATGCTGACCGGAGG  
ATTGGCCAAATGTTGATTTACAGCTCCATTATTGCTGCCCTATGCAAGGGTAAGCAATGGACAAGGCCATGG  
AGGTACACGATAGGATGGTTAGAATGGAGTTACACCCATTGCTCACGTATACTACTCTTGTGATGGATTGG  
25 CTCTTCAGGGCAGTTGACAGAGGCTATTAATCTAGAAAAGATGTCAGCAATGGTGTGAACCAAATGTTGTT  
ACTTATAGCTCGTTATGGACTATCTCTGCAAGAACGGAAGATGCACAGAACGCTAGAAAGATTGGATTCTATGG  
TCAAGAGGGCCTAAAGCCTGATATTACTACCTACAGTAGCTTACTTCATGGGTATGCTATGCAAGGAGCTTGT  
TGAGATGCATGGCTCTTGATGGTACAAAGTGTATGCAACCCGATCATTATGCTTCAACACACTAATA  
TATGCATCCGCCAAGCAAGGAAAGTAGATGAGGCCATGCTGTATTAGCAAAATGAGGCAGCAAGGATTGAAAC  
30 CTAATTGTTACGTATAGCACTTGTATTAATGGTACTGTAAAATTACTAGGATGGAGAATGCTTACTGCACTTT  
CCAAGAGATGGTGAGCAATGGTGTAGCTTAATTTATCACATATAACATAATGCTGCAAGGTTATTCGATACA  
GGAAGAACTGCTACTGCAAAAGAATTCTATGTACAGATTATCAAAAGTGGCAAAAAAGATCTTATAGAACAGGGT  
TGCTAGAAGAATTGGATGATCTATTCTTCAATGGAGGACAATGACTGTAGTACTGTCGACTCCTGCATGCTA  
A

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*Rhpr7* is a rice homolog of the *Petunia Rf-PPR592* gene.

- [0060] The nucleic acid molecule of the present invention which has the nucleotide sequence of SEQ ID NO: 32 encodes a protein or polypeptide having a deduced amino acid sequence corresponding to SEQ ID NO: 33 as follows:

- 30 -

MARRVAARARARAGGVPRSEGTIQDRARVGSGGAEDALDVDELLRRGIGAPIRSNLNGALADVARDNPAAAVSRFN  
 RMARAGASMVTPTVHTYGILIGCCSAGRDLGFAALGHVVKKGFRVEPIIFNPLLKGLCADKRTDDAMDIVLRGM  
 TELSCVPNVFSHTIILKGLCHENRSQEALLEHMMADDGGGCLPNVSYSTVIDGLLKGGDPDKAYATYREMLDRR  
 ILPNVVIYSSIIAALCKGQAMDKAMEVHDRVVKNGVTPNCFTYTSVLHGFCSSGQLTEAIKFLEKMCNSNGVEPNVV  
 5 TYSSFMDYLCKNGRCTEARKIFDSMVKRGLKPDITTYSSLHGYAIEGALVEMHGLFDLMVQSDMQPDHYVFNTLI  
 YASAKQGVDEAMLVFSKMRQQGLKPNCVTYSTLINGYCKITRMENALALFQEMVSNGVSPNFITYNIMLQGLFRT  
 GRTATAKEFYVQI IKSGKKDLIEQGLEELDDLFLSMEDNDNSTVSTPAC

[0061] Another suitable nucleic acid molecule in accordance with the present  
 10 invention is isolated from rice and identified herein as *Rhpr8*, which has a nucleotide  
 sequence of SEQ ID NO: 34, as follows:

ATGGCGCGCCGCCGCTTCCCGCCTGCTGGGCCCTCGCTCGAGGGCTCGATCCAAGGGCGAGGGGGCCGCG  
 CGGGGGCAGTGGCGGTGGCGCGGAGGACGCACGCCACGTGTCGACGAATTGCTCCGTCGTGGCATACCAGATGT  
 15 CTTCTCCTACAAATTCTCTCAACGGGCTGTGTGATGAGAACAGAACAGCCAAGAACGCTCTCGAGTTACTGCACATA  
 ATGGCTGATGATGGAGGTGACTGCCACCTGATGTTGCTGACAGCACCGTCATCAATGGCTTCAAGGAGG  
 GGGATCTGGACAAAATGCTTGACCAGAGGATTTCGCCAAATGTTGACCTACAACCTCTATTATTGCTGCGCTATG  
 CAAGGCTCAAACGTGGACAAGGCCATGGAGGTACTTACCAACATGGTAAGAGTGGTGTATGCCGATTGCGATG  
 ACATATAATAGTATTGTGCATGGTTTGTCTTCAGGGCAGCCAAAGAGGGCTATTGTATTCCTCAAAAAGATGC  
 20 GCAGTGATGGTGTGAAACCAGATGTTTACTTATAACTCGCTCATGGATTATCTTGCAAGAACGGAAGATGCAC  
 GGAAGCAAGAAAGATTTTGATTCTATGACCAAGAGGGCCCTAAAGCCTGATATTACTACCTATGGTACCGTATCCACC  
 CAGGGTATGCTACAAAGGAGCCCTGTTGAGATGCATGGCTCTGGATTTGATGGTACGAAACGGTATCCACC  
 CTAATCATTATGTTTACGCATTCTAGTATGTGCATACGCTAACACAAGAGAAAGTAGAAGAGGCAATGCTTGTATT  
 CAGCAAAATGAGGCAGCAAGGATTGAATCCGAATGCAGTGACCTATGGAACAGTTATAGATGTACTTGCAAGTCA  
 25 GGTAGAGTAGAAGATGCTATGCTTATTTGAGCAGATGATCGATGAAGGACTAAGACCTGACAGCATTGTTATA  
 ACTCCCTAATTCTAGTCTCTGTATCTTGACAAATGGAGAAGGCTGAAGAGTTATTCCTGAAATGTTGGATCG  
 AGGCATCTGTCTTAGCCTATTCTTAATTCAATAATTGACAGTCATTGCAAAAGAAGGGAGGGTTATAGAATCT  
 GGAAACTCTTGACTTGATGGTACGAATTGGTGTGAAGCCGATATCATTACCTTGGCAGGTTTGGGAGCG  
 CAAGGCGCAGACTACTCACTGTTGCTAACACATCTACTTCATCTTCAACATGTCGAACACTGGAGACAAGGAGAA  
 30 GGAGACTCCCGTCAACACCAACGGAGGAATACTGCCCTCAAACCTCCAGCGGAGGACCATTCTGGGCACATACAAC  
 ATAATCCTTCATGGACTTTGCAAAAACAAACTCACTGATGATGCACTTCGAATGTTCAAGAACCTATGTTGATGG  
 ATTTGAAGCTTGAGGCTAGGACTTTCAACATTGATGATGCACTGCTAAAGTGGCAGAAATGATGAAGCCAA  
 GGATTGTTGTTGCTTCTGCTAACGGTTAGTGGCAATTATTGGACGTACAGATTGATGGCTGAAATATT  
 ATAGGACAGGGGTGCTAGAAGAATTGGATCAACTCTTCAATGGAGGACAATGGCTGTACTGTTGACTCTG  
 35 GCATGCTAAATTCTATTGTTAGGAACTGTTGCAAGAGAGGTGAGATAACCAGGGCTGGCACTTACCTTCCATGAT  
 TGATGAGAAGCACTTTCCCTCGAACGATCCACTGCTTCTGTTATAGATCTTGCTGGGGAAAATATCAA  
 GAATATCATATATTCTCCCTGAAAAATACAAGTCCTTATAGAATCTTGAGCTGCTGA

*Rhpr8* is a rice homolog of the *Petunia Rf-PPR592* gene.

- 31 -

**[0062]** The nucleic acid molecule of the present invention which has the nucleotide sequence of SEQ ID NO: 34 encodes a protein or polypeptide having a deduced amino acid sequence corresponding to SEQ ID NO: 35 as follows:

5 MARRAASRAAGALRSEGSIQGRGGRAGGSGGAEDARHVFDELLRRGIPDVFSYNILLNLCDENRSQEALELLHI  
MADDGGDCPPDVVSYSTVINGFFKEGLDKMLDQRISPNVVTYNSIIAALCKAQTVDKAMEVLTMVKGVPDCM  
TYNSIVHGFCSGPKEAIVFLKKMRSDGVEPDVVVTYNSLMDYLCKNGRCTEARKIFDSMTKRGKPDITTYGTLL  
QGYATKGALVEMHGLLDLMVRNGIHPNHYVFSILVCAYAKQEKEVVEAMLVFSKMRQQGLNPNAVTYGTVIDVLCKS  
GRVEDAMLYFEQMIDECLRPDSIVYNSLIHSLCFDKWEKAELFLEMLRGICLSTIFFNSIIDSHCKEGRVIES  
10 GKLFDLMVRIGVKPDIITLGRFLGSARRDYSLFVNIFYFIFTNMSNTGDKEKEPTPVNTNGNTASNSSGGPFLGTYN  
IILHGLCKNKLTDALRMFQNLCMLDKLEARTFNIMIDALLKVRNDEAKDLFVAFSSNGLVPNYWTYRLMAENI  
IGQGLLEELDQLFLSMEDNGCTVDGMLNFIVRELLQRGEITRAGTYLSMIDEKHFSLEASTASLFIDLLSGGKYQ  
EYHIFLPEKYKSFIESLSC

15 **[0063]** Another suitable nucleic acid molecule in accordance with the present invention is isolated from rice and identified herein as *Rhpr9*, which has a nucleotide sequence of SEQ ID NO: 36, as follows:

ATGGCGAGAACAGAGGACGACGATACTGTAGAGCAGAGGAACAGAGGAGCGGGCACTGGTGCTCCGGTGGCTGGGA  
20 GGTGGCGACGTGGCGGCCAATGTGTTCCCGAGCGCCGCGCTGGAGAGCCCCGAGCTGCGACGGCATCACGCCGA  
CTACCGGCCGTGGCGGCCACATGGAGGCAAAGCCGGTCACTTCGCGTCAGGGCGTGCCTCCGGCGGCCAG  
CTGCAGCAGCAGCTCGTCCGGCCCACCCAATCTGGCCGATTGGCCGATCTCAGCCTCCGGAGCGGAGACCGA  
TCTGGGCCGTCCATCCGCGCCGCCAGCCAATCGACGGTGGGTGATTACTGTACTGCCAGGTGGTACCCCTCC  
GCCGCCGGCGGCCGGCGGCCGGCGCAGGCATGGCGCGCCGTGCACCACCTTACCCGCGCCCGACCCCGCC  
25 CGCGCGCGCGCGTCCCCAGCGCGCAGGGTGGTACGACCCAAGACCTAGGGCGCGGGGGCAGTGGCACCGAGG  
GCGCACGCCACGTGCTCGACAATTGGCCTACGGGCTGGGCGCTCGATCTACAGCTCAACCGCACCCCTCAC  
CGACGTCGCGCGTGCAGCCCCAGCCGAGCAGTTCGCTCTTCACCGCATGGCCCGAGCCGGCCGACGAGGTA  
ACTCCGACTTGTGCACCTACAGCATTCTCATCGTTGCTGCCGCGGGCCGCTTGGACCTCGGTTCGCGG  
CCTTGGCAATGTCATTAAGAAGGGATTAGAGTGGAAAGCCATCACCTCGCTCCTCGCTCAAGGGCTCTGTGC  
30 CGACAAGAGGACGAGCGACGCAATGGACATAGTGTCTCCGAGAATGACCGAGCTCAGCTGCATGCCAGATGTTTC  
TCCTGCACCATTCTCAAGGGTCTGTGATGAGAACAGAACAGCAAGCTCTCGAGCTGCTGCACATGATGG  
CTGATGATCGAGGAGGAGGTAGCCACCTGATGGTGTGCTGATACCAACTGTCATCAATGGCTTCTCAAAGAGGG  
GGATTCAAGACAAAGCTTACAGTACATACCATGAAATGCTGATGGAGGATTTCAACAAATGTTGTGACCTACAGC  
TCTATTATTGCTGCGTTATGCAAGGCTCAAGCTATGGACAAAGCCATGGAGGTACTTAACACCATGGTTAAGAATG  
35 GTGTCATGCCGATTGCTGATGACATATAATAGTATTCTGCATGGATATTGCTCTCAGGGCAGCCAAAGAGGCTAT  
TGGAAACTCAAAAGATGCGCAGTGTGGCGTGAACCAAATGTTACTTATAGATCACTGATGAATTATCTT  
TGCAAGAACATGGAAGATGCACCGAAGCTAGAAAGATTTGCTATGACCAAGAGGGCCTGAGATGCATGCTCTTGGATTGAT  
CTACCTATCGTACCCCTGCTTCAGGGGTATGCTACCAAAGGAGCCCTGAGATGCATGCTCTTGGATTGATATTG  
GGATCCTGAGTTCTACAAGTATTGGAGAAGTGA

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*Rhpr9* is a rice homolog of the *Petunia Rf-PPR592* gene.

- 32 -

**[0064]** The nucleic acid molecule of the present invention which has the nucleotide sequence of SEQ ID NO: 36 encodes a protein or polypeptide having a deduced amino acid sequence corresponding to SEQ ID NO: 37 as follows:

5 MARRGRRYCRAEGTEERGTGAPVAGRWRRRPNVFPSALESPELRRHADYRPWAHMEAKPVYFASRRASGRPE  
LQQQLVRPTPIWADWADSLPERRPIWAVHPRRPANRTVGVLILYQCVGDPPPPAAAAAAAGMARRVTTLTRARTRA  
RGGVPSAQGGTTQDLGRAGGSCTEGARHVLDELPLRGWGASIYSFNRTLTDVARDSPAASLNFNRMARAGADEV  
TPDLCTYSILIGCCCRAGRLDLGFAALGNVIKKGFRVEAITFAPLLKGLCADKRTSDAMDIVLRRMTELSCMPDVF  
SCTILLKGLCDENRSQEAELELLHMMADDRGGSPPDVVSYTIVNGFFKEGDSKDAYSTYHEMLDRRISPNVVTYS  
10 SIIAALCKAQAMDKAMEVLNTMVKNVMPDCMTYNISILHGYCSSGQPKEAIGTLKKMRSDGVEPNVVTYRSLMNYL  
CKNGRCTEARKITFDSMTKRGLEPDIATYRTLLQGYATKGALVEMHALLDLMDPEFYKYLEK

**[0065]** Another suitable nucleic acid molecule in accordance with the present invention is isolated from rice and identified herein as *Rhpr10*, which has a nucleotide sequence of SEQ ID NO: 38, as follows:

ATGCCCTTCCGACCGCGCCTCCGCTCCCCCTCCCTCCTGCCTCACCTCCGGGCCGCCCTCC  
CCCCGCCGCCGTGCCCGCTGGAGACCGCTCTCGTATTATCCCTCGCCGGCGGCCGCCGGAGGTGACGGA  
GTCCGAGGAGGACCGGGCTGTTGGCAGGGACACCGAGCCCCCTCCCTCCATCGCGGGATTGCACGGGAGCG  
20 CCTAGGGTTGGCTGCAATGGCGGGGGCTGCCGATGACGAGGAGGTGAGAGGAAGGCCCGCCTGTCGCC  
TCAAGCTCTGCATGAGCTCTCGCGGAGAGGAGGTGGCGCGATGCGGGCAGCCTGGCGCAGCTGGTACTGA  
GCAAGGTGAGCATGCTATGAATTTCcccATTCTGATTATCAACTCTACTCATGTGGTATCTGAATAACTATGGT  
ATTGGTGTGAGGAGGCGTAGGAATGGCATCGGTAGTTGAECTCTGATCGATATGAATGTGTGACACAGGATAT  
ATTGTTTCAGAGGCATTATCAATTGATCATTACCATATAAAAACAGTAAGAAAAGGGTCGAAAGCAATGCAT  
25 ACATAGTTGATTTGGTAGTATTACTGTAATTGTTACTAGAAGGTCTTGCAAGTATGACA  
GTAACATAAAATTGTCGCTTAACTTATTGCGCTTCTGCTGAGGATCTGGTCTGCAGCTGCTCTGTG  
ACATCTTATGAAACAGATTCAAGAGAGTGTGATTCCAACGGTTGTGTATGGGATGCTTAGCGAACAGTTATGCTAG  
AGCTCAGATGGTCATGATGCCCTTACGTTCTAGTAAAATGAGCAGCCTAACATGCAAATCTGGTGT  
TATGACAGTTATTGCAACGGCTTAAGGATGACAGACGTGGCATTGGAGCTTTGAAGAAATGGAGTCTTGTGGT  
30 TCTCTCCAGTGAATATTCGCATAGTATTATTAAATGGCCTCTGTAAGCAAGATAAGGTTGGAGAACGTTATC  
TTTCCTTCAGGAAGCTAGGAAGGAGGGAAAGTTAAACCCCTGGGAATGACCTTAAACATTCTATGCTGCATTG  
TGTAATTGGGGTTGTTAGTCTGCAAAATCATTGCTGATGCTGAAATATGGATTAGTCCCTGACAGGT  
ATACCTTTCTACCCATTACACGGCTATGTAAGTAGGTTCAATGGAGGAAGCATTGGATCTTCTGAGAGAGT  
GACAAAAGAAGGAATGGAACCTTGAGATTGTGACCTACAATAGCCTTATCAATGGTACCGATTGCTTGGTTAAC  
35 AAAGAAATTCTAAAATCATCCAGATGATGAGAGGCCAAGGTGTTGAAACCTGATCTGTTACATATACTACTTA  
TTGCTGGTCACTGCGAAAGTGGTATGTTGAAGAAGGAATGAAGGTAAAGGAAGGATGTCCTAGACCAAGGTTGCA  
GTTGAATATTGTCACATATAGTGTCTTCTCAATGCTCTCTCAAAAGGCATGTTCTGCGAAATTGACAACCTA  
CTCGCGAGATCTACAATATTGGTTGGATATGGATGTTATCGCATATTCCATCCTTATCCATGGTATTGCAACC  
TAGGGAAATTGAAAAGGCCTTCAAGTATGCAATGCAATGTCAGTTCTCAGAGGGTAATGCCAACATCACTGAA  
40 CCATTTCTATTCTTAGGACTTGCAGAAAGGATTGTTAGTGAAGCAAGGTGGTATTGGAAAATGTAGCT  
AGAAAATATCAGCCAACGTGATGATGTTCTATAATGCGTATTGATGGTTATGCAAAACTGGTATATTGAA

- 33 -

ATGCTGTTCGTTGTATGATCAGACTACTGTAGCTGGTATGCACCCAACCATTGTCACATGCAATTCTCTTCTATA  
 TGGTATTGAAAATTGGGGATCTGCAACTTGCCGAGAGCTATTTAGGGCTATTCAAGCTAAGTGGACTTCTACCA  
 ACAGCAGTGACATACACTACCTGATGGATGCACTCTGAAAGCTGGAGAAGTTAATACCATGCTAAGTCTTTG  
 ATGAAATGGTTGCAAAGAGGGATCAAGGCAAATGCAGTAACCTACAGTGTATTGTTAAAGGGCTTGTAAGCAGCT  
 5 CAGATTGAGGCTATCAATGTTCTCAAAGATATGGATAGCAAAGGTATTAATGCTGACCCGATAACCTACAAT  
 ACCCTTATACAAGGTTCTGTGAATCAGAAAACGTTAGATGGCTTCCACATACATGACATCATGTTATGCCGTG  
 GCCTGTCGCCGACACCTGTTACTTATAACTTGCTTATTAAATGTCGTGTTGAAGGGAAAAGTTATTCAAGCAGA  
 AATACTTTGGAGTCCCTCAGAGAAAATGGCATTAAGTTGAGAAAATTGCGTACACAACACTTATCAAAGCTCAG  
 TGCGCAAAGGAATGCCTATCAATGCTGTTGTTAGTTGGTAAGCTTAGATGCAGGATTGAAGCTTCTATTG  
 10 AAGATTCAGTCAGCAATCAATCGACTTGCAAAAGACAATTGCAAAGAACCTTATGTCGTAGGCAGAGCTCTGCAGAAAATAGTGAN  
 GTCTATCTACCCATATTAAATGCACTGCTGTTAAACTGGTATTAA

*Rhpr10* is a rice homolog of the *Petunia Rf-PPR592* gene.

15 [0066] The nucleic acid molecule of the present invention which has the  
 nucleotide sequence of SEQ ID NO: 38 encodes a protein or polypeptide having a  
 deduced amino acid sequence corresponding to SEQ ID NO: 39 as follows:

MPFRPRLPLPLLLLLPHLRRRSSPRPPVPAWRPLSYYPSAAAAAAEVTESEEDAAAVGRDTRAPPSIGGIARGA  
 20 PRVGCGNGGAADDEEVERKARAVARIKLCHELLERRWRAMRAALAQLVTEQSGSAALCDILWNRFRECDNSGC  
 VWDALANSYARAQMVKHDALYVLSKMSSLNMQISVFTYDSLLHGLRMTDALELFEEEMESCGVSPSEYSHSIIINGL  
 CKQDKVGEALSFLQEARKEGKFPLGTMFNILMSALCNWGFVQSAKSFLCLMLKYGLVPDRYTGSTLIHGLCKVGS  
 MEEALDLFERVTKEGMELEIVTYNSLINGYRLLGLTKEIPKIIQMMRGQGVEPDLVTYTILIAGHCESGDVEEGMK  
 VRKDVLQGLQLNIVTYSVLLNALFKKGMFCEIDNLLGETYNIGLMDVIAYSILIHGYCKLGEIEKALQVCNAMC  
 25 SSQRVMPTSLNHSILLGLCKKGLLVEARWYLENVARKYQPTDVVFYNNVIDGYAKLGDIVNAVRLYDQITVAGMH  
 PTIVTCNSLLGYCKIGDLQLAESYFRAIQLSGLLPTAVTYTTLMDALSEAGEVNMLSFLDEMVAKRIKANAVTY  
 SVIVKGLCKLRFDEAINVLKDMDSKGINADPITYNTLIQGFCESENVQMAFHIDIMLCRGLVPTVYNLLINV  
 LCLKGKVIAQELLESRLRENGIKLRKFAYTTLIKAQCAKGMPINAVLLVGKLLDAGEASIEDFSAAINRLCKRQF  
 AKEAFMFVPIMLSVGIYPDTQIYCVLGRALQKNSELVYLPILNALAVKTG  
 30

[0067] Another suitable nucleic acid molecule in accordance with the present  
 invention is isolated from *Arabidopsis thaliana*, which has a nucleotide sequence of  
 SEQ ID NO: 40, as follows:

35 ATGAAGGCTTGAGATTGATTCAAGCCTCATCTCTGAAGACAGGTAGTCTTAGAACTGATTGCTCTGTACCAATT  
 CGAGTTCTTCTAGCTGCGAACGAGACTTTCAAGTATTAGCAATGGAAATGCTGTTCAAGAGAGATTGAG  
 AAGTGGTATTGTTGATATTAAGAAAGATGATGCTATTGCTCTGTTCCAAGAAATGATTAGGTCTCGTCCTCTCCT  
 AGCTTGTGTTAGTCAAGTATTCTTAGTGCCTATTGCCAGAACAAAACAGTTCAATCTCGTGTAGATTCTGCA  
 AGCAACTGGAATTGAATGGGATTGCTCATAACATCTACACTTGAATATCATGATCAACTGCTTTGCCGGTGTG  
 40 TAAAACTTGTTTGCTTATTCTGTTGGAAAAGTAATGAAGCTTGGGTATGAGCCTGACACAACCACGTTAAC

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ACTCTGATCAAAGGACTTTCTTGAGGGTAAAGTGTCTGAAGCTGTGGTTAGTCGATAGGATGGTGGAAAACG  
 GATGTCACCTGATGTGGTTACTTATAATTGATTGAAATGGGATATGTAGATCAGGAGATACTTCTTGGCCTT  
 GGAGTTGCTCAGAAAGATGGAAGAAGAAATGTTAAGGCTGATGTGTTACTTACAGTACAATCATTGATAGTCTT  
 TGTAGAGATGGTGCATAGACGCTGCAATTAGCCTTTCAAGGAAATGGAGACGAAAGGGATTAATCTAGTGTG  
 5 TTACGTATAATTCTCTTGTGAGAGGTCTTGTAAGCCGTAATGGAATGATGGGCACTGTTGTTGAAGGATAT  
 GGTGAGTAGGAAATGTCCTAATGTCATCACTTCAATGTTGATGTTTGTCAAAGAAGGGAGCTT  
 CAGGAGGCTAATGAATTGTACAAAGAGATGATCACAAGAGGTATATCACCTAATATTATTACTTATAATACCTTGA  
 TGGATGGTATTGTATGCGAGAACCGTCTAGTGAGGCCAACAAATATGTTGATCTTATGGTTAGGAATAAGTGCAG  
 TCCTGATATCGTACTTACAAGTCTCATCAAAGGATATTGTATGGTAAAAGAGTTGACGATGGTATGAAGGTC  
 10 TTCCGCAATATTCTAAGAGAGGCTTGGTGCATGCAGTTACTTATAGCATTCTGTCCAAGGGTTTGTCAAT  
 CGGGAAAATAAAGCTCGCAGAGGAACCTTCCAAGAAATGGTTCACACGGTGTCTCCTGATGTTATGACGTA  
 TGGTATTTGCTTGTGGCTGTGACAATGGGAAGCTGAAAAGGCATTGAAATTGGAGGATTTACAAAG  
 AGTAAGATGGATCTTGTATTGTTATGTACAAACCATCATCGAGGGATGTGCAAGGGTGGAAAAGTGGAAAGATG  
 CCTGGAATTATTCTGTAGCCTACCTGTAAGGAGTGAAGCCTAATGTTATGACATACACCGTGTGATTTCAGG  
 15 ATTATGTAAGAAAGGGTCACTGTCTGAAGCAAACATCTGCTTAGAAAATGGAGGAAGATGGGAATGCGCCAAAT  
 GATTGTACATACAACACACTAATCCGGGCACATCTCGAGATGGTACTTAACATGACGCTAAACTTATTGAAG  
 AAATGAAGAGTGTGGGTTCTCAGCAGATGCTTCAGTATTAAGATGGTATCGATATGTTATTGAGTGGTGAATT  
 GGACAAAAGCTTCTAGATATGCTTCGTAA

20 SEQ ID NO: 40 is a *Arabidopsis* homolog of the *Petunia Rf-PPR592* gene.

**[0068]** The nucleic acid molecule of the present invention which has the nucleotide sequence of SEQ ID NO: 40 encodes a protein or polypeptide having a deduced amino acid sequence corresponding to SEQ ID NO: 41 as follows:

25 MKALRLIQPHLLKTGSLRTDLLCTISSFFSSCERDFSSISNGNVCFRERLRSRGIVDIKKDDAIALFOEMIRSRPLP  
 SLVDFSRRFSAIARTKQFNVLDFCKQLELNIAHNIIYTLNIMINCFCRCKTCFAYSVLGKVMKLYEPDTTTFN  
 TLIKGLFLEGKVSEAVVLVDRMVENGQPDVVTYNSIVNGICRSGDTSIALLRKMEERNVKADVFTYSTIIDS  
 CRDGCIDAAISLFKEMETKGKIKSSVVNTYNSILVRGLCKAGKWNDGALLLKDMVSREIVPNVITFNVLLDVFKEGKL  
 QEANELYKEMITRGISPNIITYNTLMDGYCMQNRLSEANNMLDMVRNKCSPDIVFTSLIKGYCMVKRVDDGMKV  
 30 FRNISKRGLVANAVTYSILVQGFCQSGKIKLAEELFQEMVSHGVLPDVMTYGILLDLCNGKLEKALEIFEDLQK  
 SKMDLGIVMYTTIIEGMCKGGKVEDAWNLFCSLPCKGVKPNMVTYTVMISGLCKKGSLSEANILRKMEEDGNAPN  
 DCTYNTLIRAHLRDGLTASAKLIEEMKSCGFSADASSIKMVIDMLLSGELDKSFLDMLS

**[0069]** Another nucleic acid molecule in accordance with the present invention has a nucleotide sequence of SEQ ID NO: 42, identified herein as *Rf-PPR591*, as follows:

1 ATATATATATACAAACTGATTTCTGTCTATTGACAGTGTATTTTACATACCCCTGAAAAAGGGTAGCTCCGCT  
 81 AATAATGTTATCTTACAAAAAATAACAATACTTTTACATAATATACAAACTCATTCTATGTATTGAAATAT  
 40 161 GATAAAAATATTGTTATTTTGTAAATATAGCTATTAGGTAGTCATGTTGTGAAATTTCCTAAAAATATTACCTGAG  
 241 TCGGCCATTGGCTAAAATATTCATTTATAGTCGATATACTCCAAGCTGTATATCCCAGAGCGACAGTATACTTC

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321 AATGGTATACTGATATCTTCTATTATACACCTTTAGTATTGTATACCCCCAATAGTATA  
 401 GTGTACCCCAATGTTGTGGTGTGGCTACAAATTGAGTGATGGTAGTGTAA  
 481 TGGAAACATTCTTTGAAAATTGAGTCAAGTCATAGTACAAAAAAACTGAA  
 561 ATTTATGTTAGTGTGGCTAGTTCTATTAGCGAGGGCAAATAGGCTACATGCC  
 5 641 AAAGATAAGTGTCTGGAAAGTATTGAAAAGATTGAGGGACAAGTGTGC  
 721 TAGCCTAGACAAACACGTCAAATTATGTA  
 801 GAAAATGCACGAAGAAATTCAAAAGCAAATATTGCTTAAGCAAGAGGCAG  
 881 TCAAAGACAAGTGTGCCTTAGGGACTTCAGACTTGTGGGCCA  
 961 ACTTCATACATTGCG  
 10 1041 CCACCTTTGGAACCTCAATCTTGTGCGCTGAACTTCATGCCTAAGT  
 1121 ATTAAAGTTCAACTTCATGCGCTTGAAGTCAACCTGTGGACTGAAGT  
 1201 TGAACCTCGCTAAGCTTGTGCGCTTGAAGTCAACCCACAAGGATT  
 1281 AAAGTTCTCAACCTGTGGCTCTCATGCACCTTACCCATTGCAAAG  
 1361 TGGCTGAAGTCAATTGAGTCAAGTAACTTAACTTAAAGGAAAGTGC  
 15 1441 CCAAGGAACTTGTGGCTTGAAGTCAACCCACAAGGATTAAAGTT  
 1521 CAACTTCATGCGCTTGAAGTCAACCTGTGGACTGAAGTCAACCT  
 1601 TGAAGTCAACCTCGCCCTTATGGTGGCTGAAGTGAAGTCAACCT  
 1681 TCAAGTAACTTCAACATATACCTCCTTATCTCGGTTATGATCTT  
 1761 GTGATGAGTCAAGTCAACCTCAACCTATGGTGTCAAGTGT  
 20 1841 CAATTCTCCAATCACCCCTACTAAGTGAAGTGAAGCGAAGATGAT  
 1921 GAGTGAAGTCAACCTCGAGAGGCCGATTGAAGTTCAGTC  
 2001 CCTTGGGTTCTAATGAATTTCAGAATGTTAGATGATGCTTCA  
 2081 AGCCTCTCCTCTGTTGCCTCTTCTCAAATTGTTGAAAGCTATGG  
 2161 TACATGAGTCAACCTCAACCTATGGTGTCAAGTGT  
 25 2241 GCATCGTACCGATCTGGATTTCTGTATTAGCATTCAAGAAAGGC  
 2321 ATTCAAAAGGGACTTTGCTGAAAGATGCTGTCATTGTC  
 2401 TGTGAGCCTAATGAAGTCATGTATGAAACGGCATGAA  
 2481 GCTCCGGTTAATGAAAGGAAACAGCAACTAACACGCA  
 2561 GGGATGCTAGATGGTGTACCAAGCCTTGAATGAGATGAA  
 30 2641 TTAATTGATGTTGTGAAAGTAACTCAGTGGAAAATGTTAGG  
 2721 ACTTCACCTCAACTCCGTATTGATGGACTATGCAAAGAGGG  
 2801 GATACATGATTGAAAAGGTGAGACCTGATGTC  
 2881 GATGAGTAAAGGCAACGGGAAATTGATTCCATGATCA  
 2961 ATAAGGATATGCCAGGCAAAGAAAATAGACGAGGCA  
 35 3041 GTATTGTTACCTGCAATGTTCTTGCATGGCTTTGAACTT  
 3121 GGGAGACTAACTGGAAAGAAGAGCTGTCATTGACTT  
 3201 GCTGTTGAGGAGAGATACA  
 3281 GATTGTC  
 3361 ATAACATACACTGCAATGATTAGTGGATTGTC  
 40 3441 GAGGCTATGTCACACTTCAAGTGGAAAGAAGAGAG  
 3521 GAGGAGCTTCTGGAGGAAATAGCTGGAAAGAGCTTCT  
 3601 CATTGAGGCTACTGCACTGGATTAAACTGC  
 3681 ACATTCGACATTGCAACACCCAAAGGCTAA  
 3761 AACACTTAAAGAACTATGCAATTGAAACAGGTA  
 45 3841 ATTATGTC  
 3921 CCCACAGTCTGCATTGAAATATGCA  
 4001 AACACCCAAACAAATTAGTC  
 4081 GAGGATGTC  
 4161 ATTGAGGATGTC  
 4241 GAGGAGGATGTC  
 4321 GAGGAGGATGTC  
 4401 GAGGAGGATGTC  
 4481 GAGGAGGATGTC  
 4561 GAGGAGGATGTC  
 4641 GAGGAGGATGTC  
 4721 GAGGAGGATGTC  
 4801 GAGGAGGATGTC  
 4881 GAGGAGGATGTC  
 4961 GAGGAGGATGTC  
 5041 GAGGAGGATGTC  
 5121 GAGGAGGATGTC  
 5201 GAGGAGGATGTC  
 5281 GAGGAGGATGTC  
 5361 GAGGAGGATGTC  
 5441 GAGGAGGATGTC  
 5521 GAGGAGGATGTC  
 5601 GAGGAGGATGTC  
 5681 GAGGAGGATGTC  
 5761 GAGGAGGATGTC  
 5841 GAGGAGGATGTC  
 5921 GAGGAGGATGTC  
 6001 GAGGAGGATGTC  
 6081 GAGGAGGATGTC  
 6161 GAGGAGGATGTC  
 6241 GAGGAGGATGTC  
 6321 GAGGAGGATGTC  
 6401 GAGGAGGATGTC  
 6481 GAGGAGGATGTC  
 6561 GAGGAGGATGTC  
 6641 GAGGAGGATGTC  
 6721 GAGGAGGATGTC  
 6801 GAGGAGGATGTC  
 6881 GAGGAGGATGTC  
 6961 GAGGAGGATGTC  
 7041 GAGGAGGATGTC  
 7121 GAGGAGGATGTC  
 7201 GAGGAGGATGTC  
 7281 GAGGAGGATGTC  
 7361 GAGGAGGATGTC  
 7441 GAGGAGGATGTC  
 7521 GAGGAGGATGTC  
 7601 GAGGAGGATGTC  
 7681 GAGGAGGATGTC  
 7761 GAGGAGGATGTC  
 7841 GAGGAGGATGTC  
 7921 GAGGAGGATGTC  
 8001 GAGGAGGATGTC  
 8081 GAGGAGGATGTC  
 8161 GAGGAGGATGTC  
 8241 GAGGAGGATGTC  
 8321 GAGGAGGATGTC  
 8401 GAGGAGGATGTC  
 8481 GAGGAGGATGTC  
 8561 GAGGAGGATGTC  
 8641 GAGGAGGATGTC  
 8721 GAGGAGGATGTC  
 8801 GAGGAGGATGTC  
 8881 GAGGAGGATGTC  
 8961 GAGGAGGATGTC  
 9041 GAGGAGGATGTC  
 9121 GAGGAGGATGTC  
 9201 GAGGAGGATGTC  
 9281 GAGGAGGATGTC  
 9361 GAGGAGGATGTC  
 9441 GAGGAGGATGTC  
 9521 GAGGAGGATGTC  
 9601 GAGGAGGATGTC  
 9681 GAGGAGGATGTC  
 9761 GAGGAGGATGTC  
 9841 GAGGAGGATGTC  
 9921 GAGGAGGATGTC  
 1000 GAGGAGGATGTC

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4081 TCTACGGGAAATTATAGTTATCTATGTGGTCGTACTTTAAAGAAAAGTATTTGTGGTTAGATTGACTGTTTCCCT
4161 CTGTCATTGATCGACTTCTTTATTACACATCAGAAGTAGGTATATGTGTACAATGCTTAAACAACGGTTGCATGTG
4241 CTCTTATGGTCGCAATTTCATCAACGAAGTCCTTGTCGAGATGCAGCTGACTGTTAAGACAAGAACCTTTCGATT
4321 GGAAATGTGATTATCCCATTCAAAGATACTTGACAGATGCTCATGATGCTTATTGACTCAGAAGAATATTCAGAAAAG
5 4401 GCATGTAGATGTGCAGCAAATGACAGAGTATGTCAATGGGTGAGGAGGACAATTATAACATTGCTCCATTGTTCACTG
4481 GCATAATGGCAATCACCTATGGATCTAAAGGACATGTTCTGCATGTAGCTAGAAGGGATGCAAGTTCACAGGGAACTAG
4561 GGATTTGGTAGACTATACCAGCCTCATTATCAGTTAGTGAATGAAAGAAACCATCAATGTAAGGAAACTCTATGG
4641 TTGTACACCTTTGAAGTTCCAAGTGTAAACTAACCTCTGGTGTATATTAGTATATAACGGTAGAATTCAATTGCA
4721 CAAGTAGATGTATCTTTGCTGGTTAGTCACTAAGGCATAATGTTCACTTAGGTTCATGCATTAAATGAAC
10 4801 ATTCAATTGATCTATGATGATGGAGTCCTGGCTGTGCATATACATGCTCAAAATTATTGTACAATGGGTTGTGACTCC
4881 AATATGTTAACATCATCCACGACATTATCTCTAATAGTTGAGATTTGTGATATTATCGTAAATGCATGTTAAGAT
4961 TATTGTAATTAGACTCTAAAGTTCTTTAGTTCTGGACAACAAAGTAATAAATCTCAAACACATGTTGCTC
5041 TCTTATTCTTGAAATAAAATATTGAGCTTTACAATGTTGACCTTACTACACTGCGTATTTTTTACTATAAGTCTATATTACCT
5121 TAAAAATTACATTACTCATGAAATTCAAAGTATCTATCACACTGCGTATTTTTTACTATAAGTCTATATTACCT
15 5201 TAGGGCAAAATTAGGCAAGTACTTACCCACATGGGTGTATATAACCAATCAACAAAGGAATTTCACACTCTATACCC
5281 ATGAAATTAAAGTATTACAAGTCATACCCATTAAAAAATTACTCTACCCATAAAACTAAAGTATTACCCAAACATACC
5361 CATTTCATTGTATACCTTGTGTTAGGTGTACCTTAAGGCTGCATAAAATTATGGTATAAAAGTCTGGAGGACCA
5441 TTTATTATTGTTACCTTTTACCTTATAATATAAGTCTGGAGGGTAGAGTCAATTATTTATGGTAGC
5521 CAGATAATATTGAAAGTAGTGGATAGTGGCTGTAAATTATTTAGATTCGTGGTATTTGTAAAGTGTCTAATCAACA
20 5601 AATGCACGTCATTGTTACAATACACTACTACACTTAGCCATAATTAAATTAAAGACATTCTCTTCATTACATCAC
5681 ATTACCATAGTTAACATTGCTATGGTTAGGTATATATATCCGGTGTAGTAAATTTCATATAAAATTATGGCAAGACGAG
5761 TAAATATGAAACTACATGCAGAGGCAGATAAAATTGGTATTTGAGTAAATTTCGTAAACATGATTAAATTATC
5841 GCGCAAAACCCCTTCAGTTGTTAACGTGTACTTATTGTTGTTACCTTACCCATAATGAAATTACCTCATTAGTG
5921 CCACATTATCTTCATAATGTGGTATTGTCAAGAACATCAATCGTGCACCTGCTACATTGAAACATGATT
25 6001 CTTTGTGGCTATTAGTCAAAATAGTAACCTGCTTCCATTGTCCTCCGGTCACCTCGGCCAATCCGGCCCTACGTT
6081 CATCAAGTACTTATTTCCATATTGTTAACGTGTACTTATTGTTGTTACCTTACAAATTGTTAATTAAATCATAGAATTAGCTG
6161 ATACACACATATATAGTGAAGGAGACTAAGTCAACTGAAGCAGCTCAAGTCAATTGTTAGGTGAAATTCTCTATCA
6241 GTTATTATGTTGTTGCTTCAAATTAAACATATTGTTAACATAGCCGACCTCAACTAATTACGCATTGATGCAAGTTCATT
6321 GTACTAGGAAAGTAAATTCAATTGTTAGTTATTGAGCAAGTTATATATACACAAATGCAATGTGCTTAT
30 6401 ATCCCTTCAATGCTAACTGACTTCATGAAATTAAATTATAGGTGTTACTTGTGAGGGACGCGAATTAAATTAA
6481 CATCACTGGTAGTGGCGAGCCAGTATTTTACTAAGGAGTATCAAATATAAGTAAATACGAAATATTAAAG
6561 GATAGTGAATCTCCTTTAATGTACTTCATTAAAGTTAGTTATTGAGAAAGTTATATACACAAATGTTGAC
6641 TGATATTCTTGATAATGATGATGCCTATGTGGATAGTGAATCTCCTTTAATGTTGATGAAAAATAAA

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35 *Rf-PPR591*, isolated from *Petunia*, has an open reading frame (“ORF”) of 1776 bp, extending between nucleotides 1882-3657, which is homologous to that of *Rf-PPR592*.

40 [0070] The nucleotide sequence of SEQ ID NO: 42 encodes a protein or polypeptide having a deduced amino acid sequence of SEQ ID NO: 43, as follows:

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MMRISVRYCLNGNPFFSFAYSIAPRHYSTNTCSISVKGNFGVSNEFQNVKCLDDAFSLFRQMVRTKPLPSVASFS
KLLKAMVHMKHYSSVSVLFREIHKLRIPIPHEFILSIVVNNSCLMHRTD LGFSVLAIHFKKGIPYNEVTFTLIRGL
FAENKVKD AVHLFKKLVR ENICEPNEVMYGTVMNGLCKKGHTQKA FDLRLM EQG STKPNTRTY TIVIDAFCKDGM
LDGATSLLNEMKQKSIPPDI FYSTLIDALCKLSQWENVRTL FLEMIHLNIYPNVCTFNSVIDGLCKEGKVEDAEE

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IMRYMIEKGVDVDITYNMIIDGYGLRGQVDRAREIFDSMINKSIEPDIIISYNILINGYARQKKIDEAMQVCREIS  
 QKGLKPSIVTCNVLLHGLFELGRTKSAQNFFDEMLSAGHI PDLYTHCTLGGYFKNGLVEEAMSHFKLERRREDT  
 NIQIYTAVIDGLCKNGKLDKAHATFEKLPLIGLHPDVITYTAMISGYCQEGLLDEAKDMLRKMEDNGCLADNR  
 VIVRGFLRSNKVSEMKAFLLEEIAKSFSEAAVVELMDIIAEDPSITRKMHWIKLHIA

5

**[0071]** Yet another nucleic acid molecule in accordance with the present invention has a nucleotide sequence of SEQ ID NO: 44, identified herein as *rf-PPR592*, as follows:

10 ATGATGAGAATTGCAGTCGCGTTACTGTCTCAATGGTAATCCCTTTCTATTCTTGCTTATTCAATTGCACCCC  
 GACATTATTCTACCAATAACACGTTCCATTCAAGTAAAGGGAAATTGGGTTCTAATGAATTGAGAATGTTAA  
 GTGTTAGATGATGCTTCAGTTGTCGTCAAATGGTAGAACTAACGCCCTCTCTGTTCTCTCT  
 AAATTGTTGAAAGCTTGGTACATATGAAGCATTACTCTTCTGTTCTCTTCGAGAAATCCACAAATTAC  
 GTATTCCCTGTTCATGAATTCACTTGTGAGCATTGTTAACAGTGTGCTTATGCATCGTACCGATCTGGATT  
 15 TTCTGTATTAGCCATTCACTTCAGAAAGGTATTCCATTAACTAAGTTATCTTAAACACCTTACTAACGGGACTC  
 TTTGCTGAAAATAAGGTTAAAGATGCTGTCATTGTTCAAAAAGTTGGTAGGGAGAAATATATGTGAGCCTAATG  
 AAGTCATGTATGGAACGGTCATGAATGGCTTGCAAAAGGGCCATACTCAAAAGCTTTGATTGCTCCGGTT  
 AATGGAACAAGGAAGTACTAACGCCAATACATGTATCTATAGCATTGTTATCGATGCCCTTGCAAAGATGGGATG  
 CTAGATGGTGTACCAAGCCTTGAATGAGATGAAACAAAAAGCATTCCCTCCGACATTAACTTATAGCACTT  
 20 TAATTGATGCTTGTTGTAAGTTAACGTCAGTGGAAAATGTTAGGACTTGTGAGATGATAACATCTTAAATAT  
 TTATCCAAATGTTGTCACCTTCAACTCCGTATTGATGGACTATGCAAAGAGGGGAAAGTAGAGACGCTGAGGAA  
 ATAATGAGATACATGATTGAAAAGGTGTAGACCTGTGATGTCACCTATAATATGATAATTGACGGATATGGCT  
 TGCGTGGTCAAGTGGATAGAGCACGGAAATTGGTATTCCATGATCAATAAGAGCATTGAGCCAATATTAG  
 CTATAATATACTAATAAAATGGATATGCCAGGCAAAGAAAATAGACGAGGCAATGCAAGTCTGCCGTGAAATTCT  
 25 CAAAAGGGATTGAAACCTAGTATTGTTACCTGCAATGTTCTTGATGGCTTTGAACCTGGAAAGAAACTAAAT  
 CTGCACAAAATTCTTGATGAGATGCTATCTGGGGGACACATACCTGATTATACACTCATTGTTACTTGCTTGG  
 TGGTTATTAAAGAATGGACTTGTGAAAGAGGCTATGTCACACTTCCATAAGTTGGAAAAGAAGGGAGAGAAGATA  
 AATATTCAAATTACACGGCTGTCATTGATGGATTGTGCAAAAGGGTAAGCTCGACAAAGCTCATGCTACGTTG  
 AGAAGCTCCCTGATAGGCTTACATCCTGATGTGATAACACACTGCAATGATTAGTGGATATTGTCAAGAAGG  
 30 GTTGTGTTAGATGAAAGCTAAAGATATGCTAAGGAAAATGGAGGACAATGGTTGTTGGCAGACAACCGAACATACA  
 GTTATTGTCGGGGATTCTCAGAAGCAATAAGTTAGTGAATGAAGGCTTCTGGAGGAAATAGCTGGAAAGA  
 GCTTCTCATTGAGGCAAGCTACTGTAGAGTTATTGATGGATATTAGCAGAGGATCCTTCTTGCTTAACATGAT  
 TCCAGAATTTCACCGGGATAATAAGAAGTGA

35 *rf-PPR592* is a gene homologous to *Rf-PPR592* and is isolated from a non-restoring *Petunia* line.

**[0072]** The nucleotide sequence of SEQ ID NO: 44 encodes a protein or polypeptide having a deduced amino acid sequence of SEQ ID NO: 45, as follows:

MMRIAVRYCLNGNPFFSFAYSIAPRHYSTNTRSISVKGNFGVSNEFENVKCLDDAFSLFRQMVRTKPLPSVVSFS  
KLLKALVHMKYSSVSVLFREIHKLRLIPVHEFILSIVVNNSCLMHTDLGFSVLAIHFKKGIPFNQVIFNTLLRGL  
FAENKVKDADVHLFKKLVRENNICEPNEVMYGTVMNGLCKKGHTQKAFLRLMEQGSTKPNTCIYSIVIDAFCKDGM  
LDGATSLLNEMKQKSIPPDIIFTYSTLIDALCKLSQENVRTLFLLEMILHNLTYPNVCTFNSVIDGLCSEGKVEDAEE  
5 IMRYMIEKGVDPDVITYNMIDIYGYGLRGQVDRAREIFDSMINKSIEPNIISYNILINGYARQKKIDEAMQVCREIS  
QKGLKPSIVTCNVLLHGLFELGRTKSAQNFFDEMLSAGHIPDLYTHCTLLGGYFKNGLVEEAMSHFKLERREDT  
NIQIYTAVIDGLCKNGKLDKAHATFEKLPLIGLHPDVITYTAMISGYCQEGLDEAKDMLRKMEDNGCLADNRTYN  
VIVRGFLRSNKVSEMAKFLEEIAAGKSFSFEAATVELLMDIIAEDPSLLNMIPEFHRSNKK

10 [0073] Also suitable in the present invention are other forms of the nucleic acid molecules shown above. An example of a nucleic acid suitable in the present invention is a nucleic acid molecule which hybridizes to a nucleotide sequence of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, 15 SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, or SEQ ID NO: 40 under stringent conditions of a hybridization buffer comprising 20% formamide in 0.9M saline/0.09M SSC buffer at a temperature of 42°C.

[0074] For the purposes of defining the level of stringency, reference can conveniently be made to Sambrook et al., Molecular Cloning: A Laboratory Manual, 20 2nd Edition, Cold Spring Harbor, NY, Cold Spring Harbor Laboratory Press, at 11.45 (1989) which is hereby incorporated by reference in its entirety. An example of low stringency conditions is 4-6X SSC/0.1-0.5% w/v SDS at 37°-45° C for 2-3 hours. Depending on the source and concentration of the nucleic acid involved in the hybridization, alternative conditions of stringency may be employed such as medium 25 stringent conditions. Examples of medium stringent conditions include 1-4X SSC/0.25% w/v SDS at > 45° C for 2-3 hours. An example of high stringency conditions includes 0.1-1X SSC/0.1% w/v SDS at 60 C for 1-3 hours. The skilled artisan is aware of various parameters which may be altered during hybridization and washing and which will either maintain or change the stringency conditions. For 30 example, another stringent hybridization condition is hybridization at 4X SSC at 65° C, followed by a washing in 0.1X SSC at 65° C for about one hour. Alternatively, an exemplary stringent hybridization condition is in 50% formamide, 4XSSC, at 42° C. Still another example of stringent conditions include hybridization at 62° C in 6X SSC, .05X BLOTO, and washing at 2X SSC, 0.1% SDS at 62° C.

[0075] The isolated nucleic acid molecule of the present invention can be from petunia, *Arabidopsis thaliana*, or rice.

[0076] The present invention also relates to an isolated protein encoded by the isolated nucleic acid molecule of the present invention which restores fertility to 5 cytoplasmic male sterile plants and modifies expression of toxic mitochondria proteins by the plant.

[0077] The present invention also relates to an isolated expression system that contains the nucleic acid molecule of the present invention which restores fertility to cytoplasmic male sterile plants and modifies expression of toxic mitochondria 10 proteins by the plant. This involves incorporating the nucleic acid molecules of the present invention into host cells using conventional recombinant DNA technology. Generally, this involves inserting the nucleic acid molecule into an expression system to which the nucleic acid molecule is heterologous (i.e., not normally present). The 15 heterologous nucleic acid molecule is inserted into the expression system which includes the necessary elements for the transcription and translation of the inserted protein coding sequences. In one embodiment, the isolated expression system of the present invention contains the nucleic acid molecule of the present invention in proper sense orientation.

[0078] The nucleic acid molecules of the present invention may be inserted 20 into any of the many available expression vectors and cell systems using reagents that are well known in the art. Suitable vectors include, but are not limited to, the following viral vectors such as lambda vector system gt11, gt WES.tB, Charon 4, and plasmid vectors such as pBR322, pBR325, pACYC177, pACYC1084, pUC8, pUC9, pUC18, pUC19, pLG339, pR290, pKC37, pKC101, SV 40, pBluescript II SK +/- or 25 KS +/- (see "Stratagene Cloning Systems" Catalog (1993) from Stratagene, La Jolla, CA, which is hereby incorporated by reference in its entirety), pQE, pIH821, pGEX, pET series (see Studier et al., "Use of T7 RNA Polymerase to Direct Expression of Cloned Genes," Gene Expression Technology, vol. 185 (1990), which is hereby incorporated by reference in its entirety), and any derivatives thereof. Recombinant 30 molecules can be introduced into cells via transformation, particularly transduction, conjugation, mobilization, or electroporation. The DNA sequences are cloned into the vector using standard cloning procedures in the art, as described by Sambrook et al., Molecular Cloning: A Laboratory Manual, 2nd Edition, Cold Spring Harbor, NY,

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Cold Spring Harbor Laboratory Press (1989); and Ausubel et al., Current Protocols in Molecular Biology, John Wiley & Sons, New York, N.Y., (1989) which are hereby incorporated by reference in their entirety.

[0079] In preparing a DNA vector for expression, the various DNA sequences 5 may normally be inserted or substituted into a bacterial plasmid. Any convenient plasmid may be employed, which will be characterized by having a bacterial replication system, a marker which allows for selection in a bacterium and generally one or more unique, conveniently located restriction sites. Numerous plasmids, referred to as transformation vectors, are available for plant transformation. The 10 selection of a vector will depend on the preferred transformation technique and target species for transformation. A variety of vectors are available for stable transformation using *Agrobacterium tumefaciens*, a soilborne bacterium that causes crown gall. Crown gall are characterized by tumors or galls that develop on the lower stem and main roots of the infected plant. These tumors are due to the transfer and 15 incorporation of part of the bacterium plasmid DNA into the plant chromosomal DNA. This transfer DNA (T-DNA) is expressed along with the normal genes of the plant cell. The plasmid DNA, pTi or Ti-DNA, for "tumor inducing plasmid," contains the vir genes necessary for movement of the T-DNA into the plant. The T-DNA carries genes that encode proteins involved in the biosynthesis of plant 20 regulatory factors, and bacterial nutrients (opines). The T-DNA is delimited by two 25 bp imperfect direct repeat sequences called the "border sequences." By removing the oncogene and opine genes, and replacing them with a gene of interest, it is possible to transfer foreign DNA into the plant without the formation of tumors or the multiplication of *Agrobacterium tumefaciens* (Fraley et al., "Expression of Bacterial 25 Genes in Plant Cells," Proc. Nat'l Acad. Sci., 80:4803-4807 (1983), which is hereby incorporated by reference in its entirety).

[0080] Further improvement of this technique led to the development of the binary vector system (Bevan, "Binary *Agrobacterium* Vectors for Plant Transformation," Nucleic Acids Res., 12:8711-8721 (1984), which is hereby 30 incorporated by reference in its entirety). In this system, all the T-DNA sequences (including the borders) are removed from the pTi, and a second vector containing T-DNA is introduced into *Agrobacterium tumefaciens*. This second vector has the advantage of being replicable in *E. coli* as well as *A. tumefaciens*, and contains a

multiclonal site that facilitates the cloning of a transgene. An example of a commonly used vector is pBin19 (Frisch et al., "Complete Sequence of the Binary Vector Bin19," Plant Molec. Biol., 27:405-409 (1995), which is hereby incorporated by reference in its entirety). Any appropriate vectors now known or later described for 5 genetic transformation are suitable for use with the present invention.

[0081] U.S. Patent No. 4,237,224 issued to Cohen and Boyer, which is hereby incorporated by reference in its entirety, describes the production of expression systems in the form of recombinant plasmids using restriction enzyme cleavage and ligation with DNA ligase. These recombinant plasmids are then introduced by means 10 of transformation and replicated in unicellular cultures including prokaryotic organisms and eukaryotic cells grown in tissue culture.

[0082] Certain "control elements" or "regulatory sequences" are also incorporated into the vector-construct. These include non-translated regions of the vector, promoters, and 5' and 3' untranslated regions which interact with host cellular 15 proteins to carry out transcription and translation. Such elements may vary in their strength and specificity. Depending on the vector system and host utilized, any number of suitable transcription and translation elements, including constitutive and inducible promoters, may be used.

[0083] A constitutive promoter is a promoter that directs expression of a gene 20 throughout the development and life of an organism. Examples of some constitutive promoters that are widely used for inducing expression of transgenes include the nopaline synthase ("NOS") gene promoter, from *Agrobacterium tumefaciens* (U.S. Patent No. 5,034,322 issued to Rogers et al., which is hereby incorporated by reference in its entirety), the cauliflower mosaic virus ("CaMV") 35S and 19S 25 promoters (U.S. Patent No. 5,352,605 issued to Fraley et al., which is hereby incorporated by reference in its entirety), those derived from any of the several actin genes, which are known to be expressed in most cell types (U.S. Patent No. 6,002,068 issued to Privalle et al., which is hereby incorporated by reference in its entirety), and the ubiquitin promoter ("ubi"), which is the promoter of a gene product 30 known to accumulate in many cell types.

[0084] An inducible promoter is a promoter that is capable of directly or indirectly activating transcription of one or more DNA sequences or genes in response to an inducer. In the absence of an inducer, the DNA sequences or genes will not be

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transcribed. The inducer can be a chemical agent, such as a metabolite, growth regulator, herbicide or phenolic compound, or a physiological stress directly imposed upon the plant such as cold, heat, salt, toxins, or through the action of a pathogen or disease agent such as a virus or fungus. A plant cell containing an inducible promoter  
5 may be exposed to an inducer by externally applying the inducer to the cell or plant such as by spraying, watering, heating, or by exposure to the operative pathogen. An example of an appropriate inducible promoter for use in the present invention is a glucocorticoid-inducible promoter (Schena et al., "A Steroid-Inducible Gene Expression System for Plant Cells," Proc. Natl. Acad. Sci., 88:10421-5 (1991), which  
10 is hereby incorporated by reference in its entirety). Expression of the transgene-encoded protein is induced in the transformed plants when the transgenic plants are brought into contact with nanomolar concentrations of a glucocorticoid, or by contact with dexamethasone, a glucocorticoid analog (Schena et al., "A Steroid-Inducible Gene Expression System for Plant Cells," Proc. Natl. Acad. Sci. USA, 88:10421-5  
15 (1991); Aoyama et al., "A Glucocorticoid-Mediated Transcriptional Induction System in Transgenic Plants," Plant J., 11: 605-612 (1997), and McNellis et al., "Glucocorticoid-Inducible Expression of a Bacterial Avirulence Gene in Transgenic Arabidopsis Induces Hypersensitive Cell Death, Plant J., 14(2):247-57 (1998), which  
20 are hereby incorporated by reference in their entirety). In addition, inducible promoters include promoters that function in a tissue specific manner to regulate the gene of interest within selected tissues of the plant. Examples of such tissue specific promoters include seed, flower, or root specific promoters as are well known in the field (U.S. Patent No. 5,750,385 issued to Shewmaker et al., which is hereby incorporated by reference in its entirety). In the preferred embodiment of the present  
25 invention, a heterologous promoter is linked to the nucleic acid of the construct, where "heterologous promoter" is defined as a promoter to which the nucleic acid of the construct is not linked in nature.

[0085] The nucleic acid construct also includes an operable 3' regulatory region, selected from among those which are capable of providing correct transcription termination and polyadenylation of mRNA for expression in the host cell of choice, operably linked to a DNA molecule which encodes for a protein of choice. A number of 3' regulatory regions are known to be operable in plants. Exemplary 3' regulatory regions include, without limitation, the nopaline synthase ("nos") 3'

regulatory region (Fraley et al., "Expression of Bacterial Genes in Plant Cells," Proc. Nat'l Acad. Sci. USA, 80:4803-4807 (1983), which is hereby incorporated by reference in its entirety) and the cauliflower mosaic virus ("CaMV") 3' regulatory region (Odell et al., "Identification of DNA Sequences Required for Activity of the 5 Cauliflower Mosaic Virus 35S Promoter," Nature, 313(6005):810-812 (1985), which is hereby incorporated by reference in its entirety). Virtually any 3' regulatory region known to be operable in plants would suffice for proper expression of the coding sequence of the nucleic acid of the present invention.

[0086] The vector of choice, suitable promoter, and an appropriate 3' 10 regulatory region can be ligated together to produce the nucleic acid construct which contains the nucleic acid molecule of the present invention, or suitable fragments thereof, using well known molecular cloning techniques as described in Sambrook et al., Molecular Cloning: A Laboratory Manual, 2nd Edition, Cold Spring Harbor, NY, Cold Spring Harbor Laboratory Press (1989); Ausubel et al., Current Protocols in 15 Molecular Biology, John Wiley & Sons, New York, N.Y. (1989), which are hereby incorporated by reference in their entirety.

[0087] Once the nucleic acid construct has been prepared, it is ready to be incorporated into a host cell. Accordingly, in another embodiment, the present invention is an isolated host cell containing the nucleic acid molecule of the present 20 invention which restores fertility to cytoplasmic male sterile plants and modifies expression of toxic mitochondria proteins by the plant. Basically, this method is carried out by transforming a host cell with the expression system of the present invention under conditions effective to yield transcription of the nucleic acid molecule in the host cell, using standard cloning procedures known in the art, such as described 25 by Sambrook et al., Molecular Cloning: A Laboratory Manual, 2nd Edition, Cold Spring Harbor, NY, Cold Spring Harbor Laboratory Press, (1989), which is hereby incorporated by reference in its entirety. Suitable host cells include, but are not limited to, bacteria, virus, yeast, mammalian cells, insect, plant, and the like. Preferably the host cells are either a bacterial cell or a plant cell. Methods of 30 transformation may result in transient or stable expression of the DNA under control of the promoter. Preferably, the nucleic acid construct of the present invention is stably inserted into the genome of the recombinant plant cell as a result of the transformation, although transient expression can serve an important purpose,

particularly when the plant under investigation is slow-growing. Plant tissue suitable for transformation include leaf tissue, root tissue, meristems, zygotic and somatic embryos, callus, protoplasts, tassels, pollen, embryos, anthers, and the like. The means of transformation chosen is that most suited to the tissue to be transformed.

5 [0088] An appropriate method of stably introducing the nucleic acid construct into plant cells is to infect a plant cell with *Agrobacterium tumefaciens* or *Agrobacterium rhizogenes* previously transformed with the nucleic acid construct. As described above, the Ti (or R1) plasmid of *Agrobacterium* enables the highly successful transfer of a foreign DNA into plant cells. Another approach to  
10 transforming plant cells with a gene which imparts resistance to pathogens is particle bombardment (also known as biolistic transformation) of the host cell, as disclosed in U.S. Patent Nos. 4,945,050, 5,036,006, and 5,100,792, all to Sanford et al., and in Emerschad et al., "Somatic Embryogenesis and Plant Development from Immature Zygotic Embryos of Seedless Grapes (*Vitis vinifera*)," Plant Cell Reports, 14:6-12  
15 (1995), which are hereby incorporated by reference in their entirety. Yet another method of introduction is fusion of protoplasts with other entities, either minicells, cells, lysosomes or other fusible lipid-surfaced bodies (Fraley et al., Proc. Natl. Acad. Sci. USA, 79:1859-63 (1982), which is hereby incorporated by reference in its entirety). The DNA molecule may also be introduced into the plant cells by  
20 electroporation (Fromm et al., Proc. Natl. Acad. Sci. USA, 82:5824 (1985), which is hereby incorporated by reference in its entirety). In this technique, plant protoplasts are electroporated in the presence of plasmids containing the expression cassette. Electrical impulses of high field strength reversibly permeabilize biomembranes allowing the introduction of the plasmids. Electroporated plant protoplasts reform the  
25 cell wall, divide, and regenerate. The precise method of transformation is not critical to the practice of the present invention. Any method that results in efficient transformation of the host cell of choice is appropriate for practicing the present invention.

[0089] After transformation, the transformed plant cells must be regenerated.  
30 Plant regeneration from cultured protoplasts is described in Evans et al., Handbook of Plant Cell Cultures, Vol. 1, MacMillan Publishing Co., NY (1983); Vasil (ed.), Cell Culture and Somatic Cell Genetics of Plants, Acad. Press, Orlando, Vol. I, 1984, and Vol. III (1986); and Fitch et al., "Somatic Embryogenesis and Plant Regeneration

from Immature Zygotic Embryos of Papaya (*Carica papaya L.*)," Plant Cell Rep., 9:320 (1990), which are hereby incorporated by reference in their entirety.

[0090] Means for regeneration vary from species to species of plants, but generally a suspension of transformed protoplasts or a petri plate containing explants 5 is first provided. Callus tissue is formed and shoots may be induced from callus and subsequently rooted. Alternatively, embryo formation can be induced in the callus tissue. These embryos germinate as natural embryos to form plants. The culture media will generally contain various amino acids and hormones, such as auxin and cytokinins. Efficient regeneration will depend on the medium, on the genotype, and 10 on the history of the culture. If these three variables are controlled, then regeneration is usually reproducible and repeatable.

[0091] Preferably, transformed cells are first identified using a selection marker simultaneously introduced into the host cells along with the nucleic acid construct of the present invention. Suitable selection markers include, without 15 limitation, markers encoding for antibiotic resistance, such as the nptII gene which confers kanamycin resistance (Fraley et al., "Expression of Bacterial Genes in Plant Cells," Proc. Natl. Acad. Sci. USA, 80:4803-4807 (1983), which is hereby incorporated by reference in its entirety), and the genes which confer resistance to gentamycin, G418, hygromycin, streptomycin, spectinomycin, tetracycline, 20 chloramphenicol, and the like. Cells or tissues are grown on a selection medium containing the appropriate antibiotic, whereby generally only those transformants expressing the antibiotic resistance marker continue to grow. Other types of markers are also suitable for inclusion in the expression cassette of the present invention. For example, a gene encoding for herbicide tolerance, such as tolerance to sulfonylurea is 25 useful, or the dhfr gene, which confers resistance to methotrexate (Bourouis et al., EMBO J., 2:1099-1104 (1983), which is hereby incorporated by reference in its entirety). Similarly, "reporter genes," which encode for enzymes providing for production of an identifiable compound are suitable. The most widely used reporter gene for gene fusion experiments has been uidA, a gene from *Escherichia coli* that 30 encodes the β-glucuronidase protein, also known as GUS (Jefferson et al., "GUS Fusions: β Glucuronidase as a Sensitive and Versatile Gene Fusion Marker in Higher Plants," EMBO J., 6:3901-3907 (1987), which is hereby incorporated by reference in its entirety). Similarly, enzymes providing for production of a compound identifiable

by luminescence, such as luciferase, are useful. The selection marker employed will depend on the target species; for certain target species, different antibiotics, herbicide, or biosynthesis selection markers are preferred.

- [0092] Plant cells and tissues selected by means of an inhibitory agent or other selection marker are then tested for the acquisition of the transgene by Southern blot hybridization analysis, using a probe specific to the transgenes contained in the given cassette used for transformation (Sambrook et al., "Molecular Cloning: A Laboratory Manual," Cold Spring Harbor, NY, Cold Spring Harbor Laboratory Press (1989), which is hereby incorporated by reference in its entirety).
- [0093] After the expression cassette is stably incorporated in transgenic plants, it can be transferred to other plants by sexual crossing. Any of a number of standard breeding techniques can be used, depending upon the species to be crossed. Once transgenic plants of this type are produced, the plants themselves can be cultivated in accordance with conventional procedure so that the nucleic acid construct is present in the resulting plants. Alternatively, transgenic seeds or propagules (e.g., cuttings) are recovered from the transgenic plants. These seeds can then be planted in the soil and cultivated using conventional procedures to produce transgenic plants.
- [0094] Thus, in other embodiments, the present invention includes transgenic plants and seeds produced by transformation with the nucleic acid molecule of the present invention which restores fertility to cytoplasmic male sterile plants and modifies expression of toxic mitochondria proteins by the plant. Examples of transgenic plants include crop plants such as alfalfa, rice, wheat, barley, rye, cotton, sunflower, peanut, corn, potato, sweet potato, bean, pea, chicory, lettuce, endive, cabbage, brussel sprout, beet, parsnip, turnip, cauliflower, broccoli, radish, spinach, onion, garlic, eggplant, pepper, celery, carrot, squash, pumpkin, zucchini, cucumber, apple, pear, melon, citrus, strawberry, grape, raspberry, pineapple, soybean, tobacco, tomato, sorghum, and sugarcane. Other examples of transgenic plants include ornamental plants such as *Arabidopsis thaliana*, *Saintpaulia*, petunia, pelargonium, poinsettia, chrysanthemum, carnation, and zinnia.
- [0095] The nucleic acid molecule of the present invention can be utilized to restore fertility to cytoplasmic male sterile plants for a wide variety of crop plants and ornamental plants. Thus, the present invention also relates to a method of restoring fertility to cytoplasmic male sterile plants involving transforming a cytoplasmic male

sterile plant with a nucleic acid molecule of the present invention under conditions effective to restore fertility to the cytoplasmic male sterile plant. The plant can have 2 or more copies of the nucleic acid molecule.

[0096] The nucleic acid molecule of the present invention can be utilized in a 5 method for identifying genes affecting male fertility or mitochondrial gene expression in other species. The first PPR gene family member whose function was characterized is *crp1*, a gene involved in RNA processing in chloroplasts (Fisk et al., “Molecular Cloning of the Maize Gene *crp1* Reveals Similarity Between Regulators of Mitochondrial and Chloroplast Gene Expression,” EMBO J., 18: 2621-2630 10 (1999), which is hereby incorporated by reference in its entirety). *Rf-PPR592* and *crp1* exhibit some sequence similarity, though there are other PPR motif proteins in the databases with greater similarity to *Rf-PPR592*. Both of these genes affect accumulation of particular RNA transcripts.

[0097] Thus, another aspect of the present invention relates to a method of 15 identifying a candidate gene restoring fertility in plants. The method involves analyzing the candidate gene for the presence of the above nucleic acid molecule in accordance with the present invention.

[0098] Identification of the nucleic acid molecules of the present invention suggests new strategies for identification of restorers or nuclear male sterility (ms) 20 alleles in crop species that are more important agriculturally than petunia. Thus, another aspect of the present invention relates to a method of identifying a candidate plant suitable for breeding with a cytoplasmic male sterile plant. The method involves analyzing the candidate plant for the presence, in its genome, of the above nucleic acid molecule of the present invention.

[0099] For example, it is possible that a nucleic acid molecule of the present 25 invention corresponds to a restorer allele in rice. Since the complete genome sequence of rice is publicly available, using the above-described method for identifying PPR motif-containing genes, candidates for rice restorer genes can be identified in the rice chromosomal region which is genetically linked to the rice 30 restoration phenotype. Using standard methods of cloning and rice transformation, the candidate rice restorer gene can be introduced as a transgene into a rice CMS line and the fertility of the transformants can be evaluated to determine whether the PPR gene is actually a restorer gene.

[0100] The fact that the nucleic acid molecules of the present invention can be a PPR motif gene can also be used to identify putative genes in other species that might encode male sterility when disrupted. The homolog of a restorer gene in one species could, when mutated, be a male sterility-encoding gene in another species.

5 Creating a male sterile line can be valuable for certain applications. For example, flowers of petunia and some other horticultural species undergo a phenomenon called pollination-induced senescence (Xu et al., "Programmed Cell Death During Pollination-Induced Petal Senescence in Petunia," *Plant Phys.*, 122:1323-1333 (2000), which is hereby incorporated by reference in its entirety). Flowers are triggered to 10 senesce when pollinated. A male sterile flower will last longer when the plant is male sterile, because no self pollen will be available to pollinate it.

[0101] Since it is possible to introduce genes into yeast mitochondria, it is likely that methods for introducing genes into plant mitochondria can be developed. When it becomes possible to introduce the *pcf* gene or a toxic homolog containing the 15 sequences on which the nucleic acid molecule of the present invention operates, then a new CMS/restorer system can be created in a different species. In such a system, a male sterile line is created by introducing *pcf* or a *pcf* homolog into the mitochondrial genome. This CMS line can then be crossed with a line containing the nucleic acid molecule of the present invention in the nuclear genome, introduced by standard 20 transformation methods, to create hybrid seed that will give rise to fertile progeny plants.

[0102] Thus, the present invention also relates to a method of producing hybrid plant seed. The method first involves providing a cytoplasmic male sterile plant. Next, a second plant containing the above nucleic acid molecule in accordance 25 with the present invention is provided. Finally, the cytoplasmic male sterile plant and the second plant are bred under conditions effective to produce hybrid progeny seed which yield fertile plants.

[0103] Another aspect of the present invention relates to a method of producing plant seeds for an inbred line of plants. The method first involves 30 providing a cytoplasmic male sterile plant. Next, a second plant containing the above nucleic acid molecule in accordance with the present invention is provided. Then, the cytoplasmic male sterile plant and the second plant are bred under conditions effective to produce hybrid progeny seed which yield fertile plants. Next, hybrid fertile plants

are produced from the hybrid progeny seeds. Finally, the hybrid fertile plants and the second plant are backcrossed to produce seed which yield inbred progeny plants.

[0104] Backcrossing methods can be used with the present invention to improve or introduce a characteristic into the present cultivar. The term backcrossing 5 refers to the repeated crossing of a hybrid progeny back to one of the parental plants for that hybrid. The parental plant which contributes the locus for the desired characteristic is termed the nonrecurrent or donor parent. This terminology refers to the fact that the nonrecurrent parent is used one time in the backcross protocol and, therefore, does not recur. The parental plant to which the locus or loci from the 10 nonrecurrent parent are transferred is known as the recurrent parent as it is used for several rounds in the backcrossing protocol (Poehlman et al., Breeding Field Crops, 4th Ed., Ames, Iowa, Iowa State University Press, (1995); Fehr, ed., Principles of Cultivar Development, Vol. 1: Theory and Technique, NY, NY, Macmillan Publishing Company (1987); and Fehr, ed., Principles of Cultivar Development, Vol. 15 2: Crop Species, NY, NY, Macmillan Publishing Company (1987), which are hereby incorporated by reference in their entirety).

[0105] In a typical backcross protocol, the phenotypically and/or commercially appealing cultivar or accession (recurrent parent) is crossed with a second cultivar (nonrecurrent parent) that carries the single locus of interest (e.g., the 20 GSB resistance gene locus) to be transferred. The resulting progeny from this cross are then crossed again to the recurrent parent and the process is repeated until a plant is obtained where essentially all of the desired morphological and physiological characteristics of the recurrent parent are recovered in the converted plant, in addition to the single transferred locus from the nonrecurrent parent.

[0106] The selection of a suitable recurrent parent is an important step for a 25 successful backcrossing procedure. The goal of a backcross protocol is to alter or substitute a single trait or characteristic in the original cultivar or accession. To accomplish this, a single locus of the recurrent cultivar is modified or substituted with the desired locus from the nonrecurrent parent, while retaining essentially all of the 30 rest of the desired genetic, and therefore the desired physiological and morphological constitution of the original cultivar. The choice of the particular nonrecurrent parent will depend on the purpose of the backcross; one of the major purposes is to add some commercially desirable, agronomically important trait to the plant. The exact

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backcrossing protocol will depend on the characteristic or trait being altered to determine an appropriate testing protocol. Although backcrossing methods are simplified when the characteristic being transferred is a dominant allele, a recessive allele may also be transferred. In this instance it may be necessary to introduce a test 5 of the progeny to determine if the desired characteristic has been successfully transferred.

[0107] A technique, known as modified backcrossing, uses different recurrent parents during the backcrossing. Modified backcrossing may be used to replace the original recurrent parent with a cultivar having certain more desirable characteristics 10 or multiple parents may be used to obtain different desirable characteristics from each.

[0108] Presently, it is possible to introduce genes into chloroplast genomes, which are maternally inherited in many species. Therefore, a CMS/restorer system can also be created by introducing the *pcf* locus, modified for chloroplast expression, 15 into the chloroplast genome. The nucleic acid molecule of the present invention can be modified to replace the mitochondrial transit sequence with a chloroplast transit sequence, using standard methods (Kohler et al., "Exchange of Protein Molecules Through Connections Between Higher Plant Plastids," Science, 276:2039-2042 (1997), which is hereby incorporated by reference in its entirety), so that the protein 20 encoded by the nucleic acid molecule of the present invention will turn off toxic gene expression in the chloroplast. Without the restorer, PCF in the chloroplast is likely to be toxic; PCF is toxic to *E. coli* bacteria. Thus, the present invention also relates to a method of producing plants with a cytoplasmic male sterile plant restoration system. The method first involves transforming a first plant in its chloroplast genome with a 25 nucleic acid which causes the plant to become male sterile. Next, a second plant is transformed with the above nucleic acid molecule in accordance with the present invention whose protein product is targeted to the chloroplast. Finally, the first and second plants are crossed to produce progeny plants possessing a cytoplasmic male sterile plant restoration system.

[0109] Since the nucleic acid molecule of the present invention prevents the expression of an organelle gene, it can be used to control the expression of a chimeric gene introduced into chloroplasts. If there is a useful protein that is desired to be produced from plant chloroplasts by introduction of a gene encoding the valuable 30

protein into the chloroplast genome, production of the valuable protein could deliberately be turned off by expressing the nucleic acid molecule of the present invention from a conditional promoter. When desired, the expression of the nucleic acid molecule of the present invention could be turned off, so that the valuable protein

5 is produced. In further detail, the method of the present invention involves: (1) engineering a chimeric gene including the coding region of a desirable protein and the *pcf* gene sequences that are regulated by the nucleic acid molecule of the present invention; (2) introducing this chimeric gene into chloroplasts of a plant and obtaining a chloroplast transgenic line; (3) engineering the nucleic acid molecule of the present

10 invention or its homologs so that the protein is targeted into chloroplasts; (4) introducing the engineered nucleic acid molecule of the present invention into the nuclear genome of a plant and obtaining a nuclear transgenic line; and (5) crossing plants to set up a regulated system. Alternatively, the plant in (2) can be made first and the gene in (3) introduced by transformation, or the plant in (4) can be made first

15 and the gene in (2) introduced into the plant in (4). When it becomes possible to transform plant mitochondrial genomes, the chimeric gene containing the *pcf* sequence can be introduced into mitochondria and the product of the nucleic acid molecule of the present invention can be targeted to mitochondria to create the analogous system.

20 [0110] The nucleic acid molecule of the present invention must be expressed in most of the plant, because in every tissue examined, the PCF protein is reduced in the presence of the nucleic acid molecule of the present invention (Nivison et al., "Identification of a Mitochondrial Protein Associated With Cytoplasmic Male Sterility in *Petunia*," Plant Cell, 1:1121-30 (1989), Nivision et al., "Sequencing,

25 Processing, and Localization of the *Petunia* CMS-Associated Mitochondrial Protein," Plant J., 5:613-623 (1994), which are hereby incorporated by reference in their entirety). The PCF protein does nevertheless vary in abundance (Conley et al., "Tissue-Specific Protein Expression in Plant Mitochondria," Plant Cell, 6:85-91 (1994), which is hereby incorporated by reference in its entirety) and is more highly

30 expressed in anthers. Very few promoters have been identified that confer expression in many tissues and in developing microspores at an early stage of pollen development. Thus, the promoter sequence of the nucleic acid molecule of the

present invention could be useful to express any of many different coding regions in a variety of tissues.

[0111] It is also possible to dissect the regulatory sequences of the nucleic acid molecule of the present invention by standard methods to identify those regions 5 that confer expression in particular tissue types. Typically, such regulatory sequences are 5' to the coding region, though the 3' flanking region can also be important. The most novel aspect of the regulatory sequences of the nucleic acid molecule of the present invention is that they confer expression in early microsporogenesis. Most of the published anther-specific promoters are not effective at the early stage of pollen 10 development, when it is critical to restore proper mitochondrial function in plants carrying the CMS cytoplasm. For example, the promoter of a nucleic acid molecule of the present invention could be used with a different coding region to restore fertility to a species with a different CMS-encoding gene. Alternatively, the promoter could be used to control a gene toxic to pollen to confer male sterility, or regulatory 15 elements from this promoter could be combined with those of another promoter to confer expression in early microsporogenesis.

[0112] Thus, another aspect of the present invention relates to a method of directing gene expression to plant mitochondria. The method involves transforming a plant with a chimeric nucleic acid molecule containing a transgene operatively linked 20 to a promoter or a terminator from a plant gene which restores fertility to cytoplasmic male sterile plants and modifies expression of toxic mitochondria proteins by the plant under conditions effective to direct expression of the transgene in the mitochondria of the transformed plant. The promoter has a nucleotide sequence of from nucleotide 1 to 1981 of SEQ ID NO: 1. The terminator has a nucleotide sequence of from 25 nucleotide 3761 to 4593 of SEQ ID NO: 1.

[0113] The present invention also relates to a promoter from a plant gene which restores fertility to cytoplasmic male sterile plants and modifies expression of toxic mitochondria proteins by the plant under conditions effective to direct expression of the transgene in the mitochondria of the transformed plant. The 30 promoter has a nucleotide sequence of from nucleotide 1 to 1981 of SEQ ID NO: 1.

[0114] Another aspect of the present invention relates to a terminator from a plant gene which restores fertility to cytoplasmic male sterile plants and modifies expression of toxic mitochondria proteins by the plant under conditions effective to

direct expression of the transgene in the mitochondria of the transformed plant. The terminator has a nucleotide sequence of from nucleotide 3761 to 4593 of SEQ ID NO: 1.

[0115] Another aspect of the present invention relates to a nucleic acid construct. The nucleic acid construct includes: (i) a promoter or a terminator from a plant gene which restores fertility to cytoplasmic male sterile plants and modifies expression of toxic mitochondria proteins by the plant under conditions effective to direct expression of the transgene in the mitochondria of the transformed plant and (ii) a nucleic acid heterologous to and operatively coupled to the promoter or the terminator. The promoter has a nucleotide sequence of from nucleotide 1 to 1981 of SEQ ID NO: 1. The terminator has a nucleotide sequence of from nucleotide 3761 to 4593 of SEQ ID NO: 1.

[0116] Other embodiments of the present invention include isolated expression systems, host cells, transgenic plants, and transgenic plant seeds containing the nucleic acid construct of the present invention.

[0117] Another aspect of the present invention is a method of expressing a gene preferentially in roots of a plant. The method involves transforming a plant with a nucleic acid construct containing a promoter suitable for driving expression preferentially in roots having a nucleotide sequence of from 1 to 1388 of SEQ ID NO: 44; a nucleic acid heterologous to the promoter, where the promoter is operatively coupled 5' to the nucleic acid to induce transcription of the nucleic acid; and a terminator having a nucleotide sequence of from nucleotide 3168 to 4016 of SEQ ID NO: 44, where the terminator is operably coupled 3' to the nucleic acid. This method can be used to express genes in roots of a plant, but not in stems, leaves, or buds.

[0118] The nucleic acid molecule of the present invention or its homologues could also be used to deliberately alter floral morphology to produce novel flowers. Thus, in another embodiment, the present invention is a method of altering plant floral morphology in ornamental plants by transforming an ornamental plant with a nucleic acid molecule of the present invention which restores fertility to cytoplasmic male sterile plants and modifies expression of toxic mitochondria proteins by the plant. Combinations of certain nuclear genes with particular mitochondrial backgrounds have been found to result in altered floral morphology in some genera. For example, in tobacco, combinations of the nuclear genome of one species with the cytoplasm of

another sometimes results in very abnormal flowers, such as flowers in which anthers have been converted to petals. While these particular plants may not be desirable horticulturally, it is possible that the coding region or expression of the nucleic acid molecule of the present invention could be manipulated so that interesting, valuable 5 floral alterations could be obtained. For example, flowers with a second set of petals in place of anthers could be attractive. A similar strategy could be pursued with other species in which novel floral morphology is desirable. Manipulation of the nucleic acid molecule of the present invention could occur, for example, by overexpressing it on a different promoter, changing the coding region, or using standard antisense or 10 gene silencing methods to underexpress homologous genes.

[0119] Another aspect of the present invention relates to a method of producing plants with a cytoplasmic male sterile plant restoration system. The method first involves mutagenizing a first plant having a nucleic acid which encodes a protein. The protein has a motif having an amino acid sequence corresponding to any 15 of SEQ ID NOs: 3 to 18 or an amino acid sequence identified with a METAMEME software using the amino acid sequence of SEQ ID NO: 2 as input or an amino acid sequence identified as significantly similar to SEQ ID NO: 2 using a NCBI BLAST software (threshold= E less than or equal to 15) with SEQ ID NO: 2 as input. Next, the mutagenized first plant is crossed with a wild-type plant having mitochondrial 20 DNA polymorphisms compared to mitochondrial DNA in the mutagenized first plant to produce progeny plants. Finally, it is determined if the progeny plants are fertile, whereby fertile progeny plants can be used as a fertile maintainer line, where the mutagenized first plant, the fertile maintainer line, and a wild-type allele present in the first plant before mutagenesis comprises a new cytoplasmic male sterile plant 25 restoration system. Figures 7A-B show this aspect of the present invention in detail.

[0120] The *Arabidopsis thaliana* genome sequence contains a PPR gene highly similar to the nucleic acid molecule of the present invention. There are existing, publicly available insertional element collections that can be screened by 30 standard methods to find a mutant in which the homolog of the nucleic acid molecule of the present invention is disrupted. The mutant can be examined to determine whether it encodes male sterility. Because the nucleic acid molecule of the present invention is important for nuclear/organelle interaction to produce male fertility, its

homologs in other species are likely to be essential for proper pollen development in those species.

- [0121] By mutating a PPR gene in a plant that does not have a CMS/restorer system, such a system can be created. In this strategy, a PPR gene is mutated and the 5 plant becomes male sterile. The mutated PPR alleles are then crossed with plants carrying other cytoplasms, present in other varieties of the plant or in intercrossable species. If a cytoplasm can be found in which the plant is fertile in the presence of the two mutated PPR alleles, then a new CMS/restorer system will have been created. A line carrying the new cytoplasm plus the mutated alleles becomes the maintainer line. 10 The line carrying the first cytoplasm plus the mutated alleles becomes *rfrf* CMS. A line carrying an unmutated allele plus a mutated allele in the presence of the CMS cytoplasm becomes *Rfrf* CMS. These lines can then be exploited just as standard maintainer, sterile, and restored lines are currently used in hybrid seed production (Figure 7).
- [0122] For example, tomato does not have a CMS/restorer system. It is known that markers near petunia *Rf* map to a region of the tomato genome where two nuclear male sterility alleles exist. Possibly, the tomato ortholog of the nucleic acid molecule of the present invention, when mutated, results in male sterility. If so, then the cytoplasms of the intercrossable wild tomato species can be tested to determine 20 whether they can confer male fertility to a tomato line homozygous for the mutated PPR gene.

- [0123] The nucleic acid molecule of the present invention may not be usable directly to restore fertility to CMS lines of most other species. Current information indicates that different mitochondrial genes are present in different CMS lines. In 25 most cases, restorer genes will have a specific mechanism of action—suppression of expression of the abnormal mitochondrial gene. However, by chance, there may be a few species that carry a CMS cytoplasm whose abnormality can be ameliorated by the nucleic acid molecule of the present invention. This can be determined by introducing the nucleic acid molecule of the present invention into the other species 30 and determining whether the transgenic plants become male fertile. If so, the nucleic acid molecule of the present invention can be used as a fertility restorer for this species.

## EXAMPLES

[0124] The following examples are provided to illustrate embodiments of the present invention but are by no means intended to limit its scope.

5    **Example 1 – Identification of Two PPR-Containing ORFs as Potential Candidates for the Rf Gene**

[0125] Previously, the isolation of a 37.5-kb BIBAC clone, SB5, that cosegregates with the Rf gene has been reported (Bentolila et al., “Identification of a 10 BIBAC Clone That Co-Segregates With the Petunia Restorer of Fertility (Rf) Gene,” *Mol. Genet. Genomics*, 266:223-230 (2001), which is hereby incorporated by reference in its entirety). SB5 is part of a contig that was constructed by screening a Petunia BIBAC library with a marker, EACA/MCTC, tightly linked to Rf. No recombination was identified between EACA/MCTC and Rf after examining 15 1,078 meiotic events. The genetic delimitation of the Rf locus was achieved only partially on the BIBAC contig. One extremity of the contig was separated from Rf by the occurrence of four recombination events, whereas no crossing-over was found between Rf and the other extremity (Bentolila et al., “Identification of a BIBAC Clone That Co-Segregates With the Petunia Restorer of Fertility (Rf) Gene,” *Mol. 20 Genet. Genomics*, 266:223-230 (2001), which is hereby incorporated by reference in its entirety). Because of the possibility that Rf might lie further away in the area not covered by the contig, a walk was initiated by screening the BIBAC library with a probe lying on the extremity that cosegregates with Rf. Unfortunately, the only hits were clones already isolated in the contig, demonstrating the presence of a gap in the 25 Petunia BIBAC library.

[0126] Before increasing the redundancy of the library to find new clones covering the gap, it was determined whether the Rf gene might lie in the SB5 clone. Because the BIBAC vector is a binary vector allowing Agrobacterium-mediated plant transformation (Hamilton, “A Binary-BAC System for Plant Transformation With 30 High-Molecular-Weight DNA,” *Gene*, 200:107-116 (1997), which is hereby incorporated by reference in its entirety), SB5 was used to restore fertility to CMS plants. Unfortunately, although SB5 is stable in *E. coli*, it underwent multiple rearrangements when introduced into *A. tumefaciens*, thus precluding its use in

transgenic experiments. Randomly chosen clones of various sizes did not show this instability in *A. tumefaciens*, pointing to special features in the sequence of the SB5 insert.

[0127] To address whether Rf might lie in the SB5 clone, shotgun sequencing 5 of the entire clone was carried out and the predicted ORFs for candidate Rf genes were examined.

[0128] Because of difficulties in contig assembly caused by the presence of 10 repeated sequences, BamHI subclones rather than the entire BIBAC clone were used as the starting material for shotgun sequencing. DNA was sonicated into 1- to 3-kb fragments, which were gel purified (Geneclean Spin Kit, Bio 101, Vista, CA), end-repaired with T4 DNA polymerase (GIBCO/BRL, Rockfield, MD) in the presence of all four dNTPs, and ligated at a mass ratio of 3 inserts to 1 vector into the SmaI site of the pTrueBlue vector (Genomics One, Buffalo, NY). The ligation product was introduced into Electromax DH10B Escherichia coli cells (GIBCO/BRL), and DNA 15 obtained from the white colonies by minipreparation was sequenced with the T7 primer in the Cornell BioResource Center. The sequences were assembled into contigs with SEQUENCHER (Gene Codes, Ann Arbor, MI).

[0129] ORFs from BIBAC SB5, their promoter region, and poly(A) signals 20 were predicted by using GENSCAN (Burge et al., "Prediction of Complete Gene Structures in Human Genomic DNA," *J. Mol. Biol.*, 268:78-94 (1997), which is hereby incorporated by reference in its entirety) with the Arabidopsis parameter matrix. Duplicated blocks in the Rf locus were determined by aligning the genomic sequence against itself by using the dot-plot feature from the MEGALIGN program (DNAstar, Madison, WI) with a 90% match. The presence of a transit peptide in the 25 ORFs was determined by using PREDOTAR version 0.5 , TARGETP (Emanuelsson et al., "Predicting Subcellular Localization of Proteins Based on Their N-Terminal Amino Acid Sequence," *J. Mol. Biol.*, 300:1005-1016 (2000); Nielsen et al., "Identification of Prokaryotic and Eukaryotic Signal Peptides and Prediction of Their Cleavage Sites," *Prot. Eng.*, 10:1-6 (1997), which are hereby incorporated by 30 reference in their entirety), and MITOPROT (Scharfe et al., *Nucleic Acids Res.*, 28:155-158 (2000), which is hereby incorporated by reference in its entirety). The length of the transit peptide was predicted by TARGETP and MITOPROT.

[0130] PPR motifs were identified in Rf-PPR592 and Rf-PPR591 by the MEME software (Bailey et al., in "Proceedings of the Second International Conference on Intelligent Systems for Molecular Biology," in Altman, eds. (Am. Assoc. Artificial Intelligence Press, Menlo Park, CA), pp. 28-36 (1994), which is hereby incorporated by reference in its entirety). The parameters for motif searching were set as minimum width = 35, maximum width = 35. The PPR consensus motif computed from the comparison of 1,303 motifs has been described previously (Small et al., "The PPR Motif - A TPR-Related Motif Prevalent in Plant Organellar Proteins," *Trends Biochem. Sci.*, 25:46-47 (2000), which is hereby incorporated by reference in its entirety).

[0131] Because the Rf gene is expected to be targeted to mitochondria where it can act upon the pcf gene to prevent its expression, an ORF predicted to carry mitochondrial transit sequences was searched. Two ORFs with putative mitochondrial targeting signals were identified. The two ORFs are adjacent to each other and appear to have originated from duplications in the promoter and coding region, but carry divergent 3' flanking regions (Figure 1A). The ORFs were 92% identical at the nucleotide level, and the predicted proteins were 93% similar, with C termini that differ completely in their final 12 aa. Both ORFs carry PPR motifs; one encodes 591 aa and the other encodes 592 aa, and were therefore named Rf-PPR591 and Rf-PPR592. A third PPR-containing ORF might lie in the vicinity of the two PPR-containing ORFs shown in Figure 1A. On the left extremity lies a genomic block that shares high similarity with the end of the coding sequence of Rf-PPR592 and its terminator region.

[0132] According to cleavage prediction programs, both putative proteins exhibited 28-residue mitochondrial transit peptides. Predicted transit peptides of Rf-PPR592 and Rf-PPR591 differed by only one substitution. To determine whether the predicted transit peptide could target a passenger protein to mitochondria, 44 codons from the 5' end of the Rf-PPR592 coding region were inserted 5' to the coding region of an enhanced GFP. DNAs of this construct and of one known to target GFP to mitochondria were bombarded into onion epidermal cells. Both GFPs appeared to be localized to the same type of organelle in the single cells shown in Figures 1B and C. Because the predicted transit peptides of Rf-PPR592 and Rf-PPR591 differed by only

one amino acid, it was expected that not only Rf-PPR592 but also Rf-PPR591 would be mitochondrially localized.

[0133] Most of the predicted mature protein (87%) of Rf-PPR592 consisted of 14 PPRs (Figure 1D). These repeats extended from the amino acid in position 54 to 5 the amino acid in position 544 and are organized in two sets of tandem repeats, one set containing 3 PPRs from amino acid 54 to amino acid 158, the other set containing 11 PPRs from amino acid 160 to amino acid 544. Because the Rf-PPR591 and Rf-PPR592 proteins are 93% similar and differ mainly in the last 12 C-terminal amino acids, their organization with respect to PPRs is identical. There was a very good 10 agreement between the consensus motif derived from the 14 PPRs found in Rf-PPR592 (hereafter designated 14 PPR consensus) and the consensus motif derived from 1,303 PPRs (hereafter designated 1303 PPR consensus) reported previously (Small et al., "The PPR Motif - A TPR-Related Motif Prevalent in Plant Organellar Proteins," *Trends Biochem. Sci.*, 25:46-47 (2000), which is hereby incorporated by 15 reference in its entirety) (Figure 1D). Whenever a discrepancy occurred between the consensus motif of the 14 PPRs in Rf-PPR592 and the 1303 PPR consensus, the difference usually was a conservative substitution. For instance, the aspartic acid in the first position of the 14 PPR consensus is replaced by a glutamic acid in the 1303 PPR consensus. Moreover, when the most frequent amino acid in the 14 PPR 20 consensus at a given position differed from the corresponding amino acid found in the 1303 PPR consensus, the amino acid in the 1303 consensus was generally the second most frequent in the 14 PPR consensus (glutamic acid at position 1, asparagine at position 18, alanine at position 28, tyrosine at position 29; Figure 1D).

[0134] It has been demonstrated that *Rf-PPR592*, a gene encoding a 592-aa 25 protein containing 14 PPRs, was able to restore fertility to CMS plants. The PPR motif, a degenerate 35-aa repeat, has been found in a very large gene family in the *Arabidopsis* genome (Small et al., "The PPR Motif - A TPR-Related Motif Prevalent in Plant Organellar Proteins," *Trends Biochem. Sci.*, 25:46-47 (2000), which is hereby incorporated by reference in its entirety). The repeats are organized in tandem 30 arrays with the number of motifs per peptide ranging from 2 to 26. About two-thirds of these *Arabidopsis* PPR proteins are predicted to be targeted to either mitochondria or chloroplasts (Small et al., "The PPR Motif - A TPR-Related Motif Prevalent in Plant Organellar Proteins," *Trends Biochem. Sci.*, 25:46-47 (2000), which is hereby

incorporated by reference in its entirety). Although distinct from the tetratricopeptide repeat (TPR), a motif that is likely to be involved in protein binding, the PPR motif shares with the former a predicted spatial structure consisting of two  $\alpha$ -helices (Small et al., "The PPR Motif - A TPR-Related Motif Prevalent in Plant Organellar Proteins," Trends Biochem. Sci., 25:46-47 (2000); Das et al., "The Structure of the Tetratricopeptide Repeats of Protein Phosphatase 5: Implications for TPR-Mediated Protein-Protein Interactions," EMBO J., 17:1192-1199 (1998), which are hereby incorporated by reference in their entirety). Tandem PPRs are thought to form a superhelix with a central spiral groove that presumably serves as the ligand-binding surface in a similar way as the one predicted for the tandem TPRs (Small et al., "The PPR Motif - A TPR-Related Motif Prevalent in Plant Organellar Proteins," Trends Biochem. Sci., 25:46-47 (2000), which is hereby incorporated by reference in its entirety). However, unlike in the TPR motif, the side chains lining the central groove of the PPR are almost exclusively hydrophilic, suggesting that some or all of the PPR motifs are RNA-binding rather than protein-binding motifs. This hypothesis is supported by the involvement in RNA metabolism and/or translation of the very few PPR motif-containing proteins characterized so far: maize chloroplast CRP1, involved in chloroplast *petD* RNA processing and *petD* and *petA* translation (Fisk et al., "Molecular Cloning of the Maize Gene *Crp1* Reveals Similarity Between Regulators of Mitochondrial and Chloroplast Gene Expression," EMBO J., 18:2621-2630 (1999), which is hereby incorporated by reference in its entirety), *Chlamydomonas* MCA1, required for the accumulation of the chloroplast *petA* transcript (Lown et al., "Chlamydomonas Nuclear Mutants That Fail to Assemble Respiratory or Photosynthetic Electron Transfer Complexes," Biochem. Soc. Trans., 29:452-455(2001), which is hereby incorporated by reference in its entirety), yeast PET309, required for the stability and translation of the *coxI* mitochondrial mRNA (Manthey et al., "The Product of the Nuclear Gene PET309 is Required for Translation of Mature mRNA and Stability or Production of Intron-Containing RNAs Derived from the Mitochondrial COX1 Locus of *Saccharomyces cerevisiae*," EMBO J., 14:4031-4043 (1995), which is hereby incorporated by reference in its entirety), and *Drosophila* BSF, which binds to and stabilizes the bicoid mRNA (Mancebo et al., "BSF Binds Specifically to the *bicoid* mRNA 3' Untranslated Region and Contributes to Stabilization of *bicoid* mRNA," Mol. Cell. Biol., 21:3462-3471 (2001), which is

hereby incorporated by reference in its entirety). That *Petunia Rf* belongs to this family is consistent with its similarity of action to *crp1*, *mca1*, and *pet309*. Mutations in these three genes result in lack of accumulation of a particular transcript and reduced abundance of an organelle protein. Likewise, in *Petunia* restored plants,

5 among the population of *pcf* transcripts with different 5' termini, the ones with termini at -121 exhibit reduced abundance and the amount of the PCF protein is greatly reduced (Pruitt et al., "Transcription of the Petunia mitochondrial CMS-Associated Pcf Locus in Male Serile and Fertility-Restored Lines," Mol. Gen. Genet., 227:348-355 (1991); Nivison et al., "Identification of a Mitochondrial Protein Associated with

10 Cytoplasmic Male Sterility in Petunia," Plant Cell, 1:1121-1130 (1989), which are hereby incorporated by reference in their entirety). However, the alleles of the other PPR genes that are known to reduce RNA and/or protein accumulation are recessive, whereas the *Petunia Rf* allele is dominant. *Rf* genes from other species have been shown to alter the RNA transcript profile of the CMS-associated genes (Wise et al.,

15 "Mutator-Induced Mutations of the *rfl* Nuclear Fertility Restorer of T-Cytoplasm Maize Alter the Accumulation of T-*urf13* Mitochondrial Transcripts," Genetics, 143:1383-1394 (1996); Singh et al., "Suppression of Cytoplasmic Male Sterility by Nuclear Genes Alters Expression of a Novel Mitochondrial Gene Region," Plant Cell, 3:1349-1362 (1991); Tang et al., "Transcript Processing Internal to a Mitochondrial

20 Open Reading Frame is Correlated with Fertility Restoration in Male-Sterile Sorghum," Plant J., 10:123-133 (1996); Moneger et al., "Nuclear Restoration of Cytoplasmic Male Sterility in Sunflower is Associated with the Tissue-Specific Regulation of a Novel Mitochondrial Gene," EMBO J., 13:8-17 (1994), which are hereby incorporated by reference in their entirety). In some cases, restoration has

25 been shown to result from enhanced processing of the CMS-associated transcripts (Tang et al., "Transcript Processing Internal to a Mitochondrial Open Reading Frame is Correlated with Fertility Restoration in Male-Sterile Sorghum," Plant J., 10:123-133 (1996); Menassa et al., "Post-Transcriptional and Developmental Regulation of a CMS-Associated Mitochondrial Gene Region by a Nuclear Restorer Gene," Plant J.,

30 17:491-499 (1999), which are hereby incorporated by reference in their entirety). Taken together, these observations suggest that *Rfs* in other species could also be PPR-containing genes like the *Petunia Rf*.

- [0135] The data presented here show that a pair of duplicated PPR-containing genes, denoted *Rf-PPR591* and *Rf-PPR592*, lie in the *Petunia Rf* locus. A third related PPR gene might lie in the area not covered by the SB5 BIBAC clone as suggested by the high similarity between the sequence available at the end of the clone and the  
5 sequence present at the end of the coding sequence of *Rf-PPR592* and in its terminator region.
- [0136] In *Brassica napus*, the restorer locus has been shown to affect the transcripts of several mitochondrial genes, two of them being associated with the *nap* and *pol* CMS (Singh et al., "Nuclear Genes Associated With a Single Brassica CMS  
10 Restorer Locus Influence Transcripts of Three Different Mitochondrial Gene Regions," *Genetics*, 143:505-516 (1996); Li et al., "Restorer Genes for Different Forms of *Brassica* Cytoplasmic Male Sterility Map to a Single Nuclear Locus That Modifies Transcripts of Several Mitochondrial Genes," *Proc. Natl. Acad. Sci. USA*, 95:10032-10037 (1998), which are hereby incorporated by reference in their entirety).  
15 At the same locus have been mapped *Rfp*, the restorer gene to the *pol* CMS, that modifies the transcripts of the *pol* CMS-associated *orf224/atp6* mitochondrial DNA region, *Rfn*, the restorer gene to the *nap* CMS that modifies the transcripts of the *nap* CMS-associated *orf222/nad5c/orf139* mitochondrial DNA region, and *Mmt* (modifier of mitochondrial transcripts), a gene that modifies the transcripts of the *nad4* gene and  
20 another gene possibly involved in cytochrome *c* biogenesis (Li et al., "Restorer Genes for Different Forms of *Brassica* Cytoplasmic Male Sterility Map to a Single Nuclear Locus That Modifies Transcripts of Several Mitochondrial Genes," *Proc. Natl. Acad. Sci. USA*, 95:10032-10037 (1998), which is hereby incorporated by reference in its entirety). The resolution of the genetic mapping in these studies did not allow the  
25 authors to address whether the three genes represent different alleles of a single gene or whether the restorer locus might contain multiple, related, tightly linked genes. A similar situation occurs in *Sorghum*, where at the *Rf3* locus, one of the two restorers to A3 CMS, has been mapped a gene that regulates the transcript-processing activity of A3 CMS-associated *orf107* and the *Mmt1* gene that enhances the transcript processing  
30 of *urf209* (Tang et al., "Cosegregation of Single Genes Associated with Fertility Restoration and Transcript Processing of Sorghum Mitochondrial *orf107* and *urf209*," *Genetics*, 150:383-391 (1998), which is hereby incorporated by reference in its

entirety). As in *Brassica napus*, either a multiallelic model or tightly linked genes could account for this result.

- [0137] It will be worthwhile to determine whether *Rf-PPR591* affects the profile of mitochondrial transcripts other than *pcf* in transgenic plants. If so, it would  
5 strengthen the hypothesis that *Rf* alleles arise as modifications, perhaps through duplication, of existing alleles that control mitochondrial gene expression. According to this theory, once CMS occurs in a plant species, there may be strong selective pressure for the plant to overcome it by recruiting preexisting activities and redirecting them to down-regulate the expression of CMS-encoding genes.  
10 Conceivably, recombination among closely related PPR-containing genes could have led to the appearance of the *Rf-PPR592* gene.

**Example 2 - A Deletion in the Promoter of *rf-PPR592* Prevents Its Expression in CMS Floral Buds**

- [0138] If one of the candidate ORFs, *Rf-PPR591* or *Rf-PPR592*, is the *Rf* gene, some sequence polymorphism between the allele of these ORFs found in a restorer line (*Rf/Rf*) and the allele found in a CMS plant (*rf/rf*) might be expected. Presumably some difference in the sequences of the dominant *Rf* allele vs. the  
15 recessive nonrestoring allele *rf* must reflect their opposite restoring ability. The sequence of *rf-PPR592* was obtained by amplifying genomic DNA of a Petunia hybrida *rf/rf* plant, where *rf* was inherited from a *P. hybrida* line called 2423, with the PfuTurbo Hotstart DNA polymerase (Stratagene, La Jolla, CA) and PCR primers flanking *Rf-PPR591* (5'-TGCACAGTGTATATTACATACCC-3'; SEQ ID NO:  
20 46) and *Rf-PPR592* (5'-TTTATGATACATGGATTCAACGAC-3'; SEQ ID NO:  
47). A PCR product was obtained only with a primer specific to the 3' flanking region of *Rf-PPR592*, not with a primer specific to the 3' flanking region of *Rf-PPR591*. The *rf-PPR592* PCR product showed a reduction in size of about 500nt compared with the  
25 *Rf-PPR592* PCR product amplified from the genomic DNA of an *Rf/Rf* line (Figure 2A). Using the same primers, a PCR product similar in size to *rf-PPR592* was amplified from another nonrestoring *P. hybrida* line as well as from a nonrestoring Petunia parodii line. The *rf-PPR592* PCR product amplified from the *P. hybrida* 2423 sequence was cloned into the pCR-Blunt II-TOPO vector (Invitrogen, Carlsbad, CA) and sequenced, revealing a gene 97% identical to *Rf-PPR591* and 94% identical

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to Rf-PPR592 in the coding region, with the predicted proteins 98% and 94% similar, respectively. Because of the primer design, the rf-PPR592 sequence lacks 35 nt available for Rf-PPR592. Similarity blocks between rf-PPR592, Rf-PPR592, and Rf-PPR591 were determined by comparing the aligned sequences with SEQUENCHER.

5 Percent similarity was computed by using the MEGALIGN program. Comparison of the similarities of regions of the three different PPR genes revealed that the 5' promoter region of rf-PPR592 is most similar to Rf-PPR591, whereas the 3' flanking region of rf-PPR592 is most similar to Rf-PPR592. The genomic structure of rf-PPR592 was consistent with the past occurrence of recombination between two genes  
10 similar to Rf-PPR591 and Rf-PPR592 (Figure 2B). Because PCR amplification could have resulted in an artificial recombination between Rf-PPR591 and Rf-PPR592 due to their high similarity, the Rf-PPR592 PCR product was resequenced as a control experiment. The sequences of three rf-PPR592 and Rf-PPR592 clones were determined. No evidence of recombination was found in any of the sequenced Rf-  
15 PPR592 clones, thus precluding PCR amplification as the source of the genetic mosaic found in the rf-PPR592 ORF.

[0139] rf-PPR592 carries a 530-nt deletion from -556 to -27 relative to the start codon of Rf-PPR592. This deletion is responsible for the observed difference in the sizes of the respective amplicons. Rf-PPR591 has a 49-nt gap within the same  
20 region, from -273 to -224 relative to the start codon of Rf-PPR592 (Figure 2B).

[0140] RT-PCR experiments were performed to determine whether both Rf-PPR592 and rf-PPR592 are expressed in Petunia floral buds. The RT reaction was performed with Superscript II RNase H- reverse transcriptase (GIBCO), and the PCR was performed with the PfuTurbo Hotstart DNA polymerase. The reverse primer R3 used for reverse transcription (RT)-PCR lies in the 3' untranslated region of the Rf-PPR592 gene at position +430 to +454. The forward primer used for the PCR lies in the coding sequence and is specific to the rf or Rf allele, F2S or F2, respectively, because of DNA polymorphisms between rf and Rf in this area. Primer pairs F2SR3 amplified a 1333-bp product and F2R3 amplified a 1507-bp product. R3, 5'-  
25 TGAAAATGACAATCGAACAGAAAA-3' (SEQ ID NO: 48); F2, 5'-AACATTCCCTCCAGACATTATTACA-3' (SEQ ID NO: 49); F2S, 5'-GACGCTGAGGAAATAATGAGATAC-3' (SEQ ID NO: 50).

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[0141] An Rf-PPR592 transcript was detected in floral buds in lines carrying the Rf allele, but no transcripts of rf-PPR592 were detected in a homozygous nonrestoring rf/rf line (Figure 3A). The absence of the upstream 530-nt region in rf-PPR592 is likely to prevent the expression of PPR592 in the floral buds of  
5 nonrestoring lines.

[0142] Since rf-PPR592 encodes a protein that is very similar to the one encoded by Rf-PPR592, a survey of its expression was conducted in tissues other than the floral buds. From all of the tissues analyzed, an rf-PPR592 transcript was detected only in roots of a nonrestoring rf/rf line (Figure 3B).

[0143] A deletion of 530 nt in the promoter area of the *rf-PPR592* gene is the likely cause of its nonexpression in the floral buds of CMS plants. That the *rf-PPR592* gene, which encodes a protein 98% similar to Rf-PPR591 and 94% similar to Rf-PPR592, has not yet accumulated missense mutations suggests either a recent deletion in the promoter or a functional expression in plant organs other than the floral buds. This latter possibility was supported by the finding of an *rf-PPR592* transcript in the roots of homozygous nonrestoring *rf/rf* line.  
15

[0144] Sequence inspection demonstrated that a recombination event between two genes similar to *Rf-PPR591* and *Rf-PPR592* can explain the formation of *rf-PPR592*. Perhaps once *Rf-PPR592* was generated and happened to prevent the expression of *pcf*, its maintenance required the presence of the CMS-associated gene. The absence of the CMS-associated gene in new nucleocytoplasmic combinations might have resulted in recombination between *Rf-PPR591* and *Rf-PPR592* because of their high similarity. In *Brassica* and related genera, *Rfn* is found only in association with the *nap* cytoplasm, suggesting that the evolutionary appearance of the *nap* cytoplasm and the attending male sterility may have provided the selective pressure for the origin, and possibly the continued presence, of *Rfn* in *B. napus* (Li et al.,  
25 “Restorer Genes for Different Forms of *Brassica* Cytoplasmic Male Sterility Map to a Single Nuclear Locus That Modifies Transcripts of Several Mitochondrial Genes,” *Proc. Natl. Acad. Sci. USA*, 95:10032-10037 (1998), which is hereby incorporated by reference in its entirety). Sampling of more *rf-PPR592* genes from different *Petunia* species should help in understanding the evolution of CMS and fertility restoration in this genus.  
30

**Example 3 – Rf-PPR592 Is Able to Restore Fertility to CMS Plants**

[0145] A sequence encoding the N-terminal 44 aa of Rf-PPR592 was inserted 5' to the green fluorescent protein (GFP) sequence in the pOL vector (Peeters et al.,  
5 “Duplication and Quadruplication of Arabidopsis Thaliana Cysteinyl- and Asparaginyl-tRNA Synthetase Genes of Organellar Origin,” J. Mol. Evol., 50:413-423 (2000), which is hereby incorporated by reference in its entirety) to use in transient assay of protein localization. As a control, a vector carrying GFP fused with a known mitochondrial coxIV transit peptide (Akashi et al., “Potential Dual Targeting  
10 of an Arabidopsis Archaeabacterial-Like Histidyl-Trna Synthetase to Mitochondria and Chloroplasts,” FEBS Lett., 431:39-44 (1998), which is hereby incorporated by reference in its entirety) was also used in the transient assays. DNAs of GFP constructs were bombarded into onion epidermal cells as described in Scott et al., “Model System For Plant Cell Biology: GFP Imaging In Living Onion Epidermal  
15 Cells,” BioTechniques, 26:1125, 1128-1132 (1999), which is hereby incorporated by reference in its entirety.

[0146] For the stable transformation experiments, genomic DNA from the Rf-PPR592 gene was amplified from the SB5 BIBAC clone with the PfuTurbo Hotstart DNA polymerase and the primers F11-XbaI (5'-  
20 TCTAGAAAAAAATGAAGGGGGATCAAT-3'; SEQ ID NO: 51) and R11-EcoRI (5'-GAATTCACTTGCTCTCACGATAAACTAAGA-3';SEQ ID NO: 52)  
(underlined are the restriction sites added to the 5' end of the primers for further use in the cloning of the PCR product). The PCR product was first cloned into the pCR-Blunt II-TOPO vector, and its sequence was checked to be free of possible mutations  
25 generated by the polymerase. The PCR product was then released from the pCR-Blunt II-TOPO vector by digestion with XbaI and EcoRI, gel purified, and cloned into XbaI/EcoRI-digested binary vector pGPTVKan (Becker et al., “New Plant Binary Vectors With Selectable Markers Located Proximal to the Left T-DNA Border,” Plant Mol. Biol., 20:1195-1197 29 (1992), which is hereby incorporated by reference in its entirety). Petunia transformation and regeneration were performed as described in Horsch et al., “A Simple and General-Method for Transferring Genes Into Plants,” Science, 227:1229-1231 30 (1985), which is hereby incorporated by reference in its entirety. Transformants were selected on 300 mg/liter kanamycin, 100 mg/liter

ticarcillin/clavulanic acid (15:1, Duchefa Biochemie, Harlem, The Netherlands).

Shoots were rooted on N13 medium (O'Connell et al., "Somatic Hybridization Between *Lycopersicon-Esculentum* and *Lycopersicon-Pennellii*," Theor. Appl. Genet., 70:1-12 (1985), which is hereby incorporated by reference in its entirety)

5 before transfer to soil.

[0147] To determine whether Rf-PPR592 could restore fertility to rf/rf CMS lines, a 4.6-kb fragment carrying the entire coding region was introduced into the binary vector pGPTVKan. This fragment carries 2007 nt upstream of the start codon and 861 nt downstream of the stop codon. The pGPTVKan-4.6 kb Rf-PPR592 vector 10 was transferred into *A. tumefaciens* strain LBA4404, which was used to transform a *P. parodii* rf/rf CMS line (Figure 4A) and a *P. hybrida* rf/rf CMS line (Figure 4C). More than two dozen independent transformants were obtained and grown to 15 flowering. Fertile transformants were observed after transformation of both lines (Figures 4B and D). Among these were several fertile transformants carrying a single copy of the introduced Rf-PPR592 genomic DNA. Flowers of one of the *P. parodii* primary transformant plants were selfed, and a population of 40 T1 progeny was grown to flowering.

[0148] DNA extractions and Southern blotting were performed as described in Bentolila et al., "Locating the Petunia Rf Gene on a 650 kb DNA Fragment," Theor. Appl. Genet., 96:980-988 (1998), which is hereby incorporated by reference in its entirety. Floral bud protein was prepared for cell culture protein as described in Kohler et al., "The Green Fluorescent Protein as a Marker to Visualize Plant - Mitochondria *in vivo*," Plant Journal, 11:613-621 (1997), which is hereby incorporated by reference in its entirety. After separation by SDS/PAGE (15%), 25 immunoblots on Hybond-P poly(vinylidene difluoride) membranes (PVDF; Amersham Pharmacia, Picataway, NJ) were prepared as previously described (Reed et al., "High-Level Expression of a Synthetic Red-Shifted GFP Coding Region Incorporated into Transgenic Chloroplasts," Plant J., 27:257-2653 (2001), which is hereby incorporated by reference in its entirety) and probed with a 1:5000 dilution of 30 the anti-PCF antibody (Nivison et al., "Sequencing, Processing, and Localization of the Petunia CMS-Associated Mitochondrial Protein," Plant J., 5:613-623 (1994), which is hereby incorporated by reference in its entirety).

[0149] DNA blot hybridization revealed that the fertile phenotype cosegregated with the Rf-PPR592 transgene (Figure 5A). The T1 progeny were also surveyed for the presence of the CMS-associated 19.5-kDa PCF protein. The 19.5-kDa protein was found to be decreased about 10-fold in fertile progeny restored by 5 Rf-PPR592 relative to sterile progeny and the parental CMS line (Figure 5B). Thus, Rf-PPR592 was capable of restoring fertility by decreasing the amount of the PCF protein.

[0150] The cloning of a gene that can restore fertility to male-sterile *Petunia* lines will facilitate elucidation of the mechanism by which expression of the CMS-10 associated mitochondrial gene is suppressed. The reduced amount of the PCF protein could be due to a reduction in the abundance of one of the *Petunia* CMS-associated transcripts, which was reported previously (Pruitt et al., "Transcription of the *Petunia* Mitochondrial CMS-Associated *pcf* Locus in Male Sterile and Fertility-Restored Lines," Mol. Gen. Genet., 227:348-355 (1991), which is hereby incorporated by 15 reference in its entirety), or to a translation defect that destabilizes the transcript. In yeast, mutation in a transcript-specific translation factor destabilizes the particular transcript with which the factor normally interacts (Poutre et al., "PET111, a *Saccharomyces cerevisiae* Nuclear Gene Required for Translation of the Mitochondrial mRNA Encoding Cytochrome C Oxidase Subunit II," Genetics, 20: 115:637-647 (1987), which is hereby incorporated by reference in its entirety).

[0151] A number of fertility restorer genes in other species are known to alter transcript profiles and mitochondrial gene product accumulation (Moneger et al., "Nuclear Restoration of Cytoplasmic Male Sterility in Sunflower is Associated with the Tissue-Specific Regulation of a Novel Mitochondrial Gene," EMBO J., 13:8-17 25 (1994); Singh et al., "Nuclear Genes Associated With a Single Brassica CMS Restorer Locus Influence Transcripts of Three Different Mitochondrial Gene Regions," Genetics, 143:505-516 (1996); Dewey et al., "Novel Recombinations in the Maize Mitochondrial Genome Produce a Unique Transcriptional Unit in the Texas Male-Sterile Cytoplasm," Cell, 44:439-49 (1986); Wise et al., "Mitochondrial Transcript Processing and Restoration of Male Fertility in T-Cytoplasm Maize," J Hered, 30 90:380-385 (1999), which are hereby incorporated by reference in their entirety). In addition to the molecular phenotype of restoration, the *Petunia* Rf locus and Rf loci from other species may be similar in genomic organization (Li et al., "Restorer Genes

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for Different Forms of *Brassica* Cytoplasmic Male Sterility Map to a Single Nuclear Locus That Modifies Transcripts of Several Mitochondrial Genes," *Proc. Natl. Acad. Sci. USA*, 95:10032-10037 (1998); Tang et al., "Cosegregation of Single Genes

Associated with Fertility Restoration and Transcript Processing of Sorghum

- 5 Mitochondrial *orf107* and *urf209*," *Genetics*, 150:383-391 (1998), which are hereby incorporated by reference in their entirety). The identification of *Petunia Rf* as a PPR family member suggests that searching for PPR motif genes near known restorer loci should be a useful strategy to identify candidate restorer genes in other species.
- Further studies of *Rf-PPR592* and other PPR motif-containing genes in plants, fungi,
- 10 and animals will be required to determine whether the motif has a direct role in RNA-protein and/or protein-protein interactions.

**Example 4 – Use of *Rf-PPR592* or its Homologs/Derivatives to Create Novel Floral Structures**

- 15 [0152] In Petunia, recombination events near the *Rf* locus in standard sexual crosses resulted in plants with abnormal floral appearance. Moreover, a few of the initial transgenic plants transformed by *Rf-PPR592* produced flowers with abnormal appearance. Furthermore, a number of transgenic plants transformed by *Rf-PPR592* and *Rf-PPR591* exhibit abnormalities in floral and vegetative structures. An example of abnormal flowers seen in some transgenic plants are shown in Figure 6.

**Example 5 - Identification of a Rice Fertility Restorer Gene**

- 25 [0153] The complete rice genome sequence, which has been deposited in EMBL/GenBank/DDBJ, was examined for genes similar to the petunia *Rf* gene, using BLASTP. The gene most similar to the petunia *Rf* locus was termed as *Rice homolog of Petunia restorer 1 (Rhpr1)*. This gene is located very close to the rice *Rf4* marker C1261. There were a total of 10 PPR genes in the vicinity of this marker on rice
- 30 chromosome 10, which were termed as *Rhpr1* to *Rhpr10* (SEQ ID NOS: 20, 22, 24, 26, 28, 30, 32, 34, 36, and 38).

[0154] The most immediate usefulness of identification of the rice restorer gene is in marker-assisted selection. This facilitates introduction of the natural wild abortive-restorer *Rf4* gene, which can restore fertility to the wild abortive cytoplasm

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by traditional crosses into elite breeding lines for use as a parent in a three-line breeding scheme. This use of the information does not involve genetically modified organisms and, therefore, can proceed without any of the attendant issues. Random screening has identified some molecular markers that are already being used to 5 transfer *Rf* genes in certain nuclear backgrounds (Ichikawa et al., "A Rapid PCR-Aided Selection of a Rice Line Containing the *Rf-I* Gene Which is Involved in Restoration of the Cytoplasmic Male Sterility," Molecular Breeding, 3:195-202 (1997); Jing et al., "Mapping Fertility-Restoring Genes of Rice WA Cytoplasmic 10 Male Sterility Using SSLP Markers," Bot. Bull. Acad. Sin., 42:167-171 (2001), which are hereby incorporated by reference in their entirety). Knowing the actual *Rf* gene sequence makes laborious screening for markers suitable between different breeding 15 lines unnecessary.

[0155] The next possibility is to more rapidly transfer the *Rf4* gene into existing elite breeding lines by transformation rather than by sexual crosses. In such a 15 strategy, the entire natural *Rf4* gene would be used to transform a rice line for the three-line hybrid rice production method.

[0156] Because the three-line method for hybrid rice production requires time-consuming breeding and labor, presently there are attempts to exploit temperature-sensitive male sterility mutants for a two-line method of hybrid seed production. The 20 three-line method for hybrid rice production involves construction of three lines. Two lines are backcrossed repeatedly so that they contain the same nuclear genome. One contains the CMS cytoplasm ("CMS parent") and is male sterile while the other ("Fertile Maintainer") contains the normal cytoplasm but no restorer of fertility (*Rf*) alleles. By crossing the maintainer as male and the CMS line as female, seeds of the 25 CMS line with a known nuclear background can be produced in large quantity. The third line is homozygous for one or more fertility restoration loci. Hybrid seed is produced by crossing the third line with the CMS parent. The nuclear genomes of the third line and the CMS line are selected by breeders to optimize heterosis and desirable characteristics for the region in which the hybrid rice will be grown.

[0157] Rice plants have been found that contain a mutant allele that encodes 30 male sterility at high temperatures but fertility at low temperatures (Dong et al., "Molecular Mapping of a Rice Gene Conditioning Thermosensitive Genic Male Sterility Using AFLP, RFLP and SSR Techniques," Theor Appl Genet., 100:727-734

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(2000), which is hereby incorporated by reference in its entirety). By growing the rice at high temperatures, it can be used as the sterile parent in a cross with an elite breeding line. The mutant rice can be propagated by selfing when grown at low temperatures. The use of this method in the field on large-scale has not been reported 5 in the literature, so the feasibility of using a natural temperature-sensitive mutant is not known.

[0158] A cloned *Rf4* gene could also be used in a two-line method. In this scheme, the *Rf4* gene regulatory sequences would be engineered so that it could be turned on when desired. Then, both a CMS line and a maintainer line, which requires 10 multiple crosses over a number of years to produce, are not needed. A single line, the CMS line containing the engineered *Rf4* gene, would serve both as CMS parent and as its own maintainer line (Figure 8). The CMS line would be propagated by selfing by turning on the *Rf4* gene. Without induction of the engineered *Rf4* gene, however, the line would be sterile and therefore could be used as a CMS parent.

[0159] Although the invention has been described in detail for the purpose of illustration, it is understood that such detail is solely for that purpose, and variations 15 can be made therein by those skilled in the art without departing from the spirit and scope of the invention which is defined by the following claims.

**WHAT IS CLAIMED:**

1. An isolated nucleic acid molecule which restores fertility to cytoplasmic male sterile plants and modifies expression of toxic mitochondria proteins by the plant, wherein the nucleic acid molecule: (1) encodes a protein having an amino acid sequence selected from the group consisting of SEQ ID NOs: 2, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, and 41; (2) encodes a protein comprising a motif having an amino acid sequence corresponding to any of SEQ ID NOs: 3 to 18 or an amino acid sequence identified by a METAMEME software using the amino acid sequence of SEQ ID NO: 2 as input or an amino acid sequence identified as significantly similar to SEQ ID NO: 2 using a NCBI BLAST software (threshold= E less than or equal to 15) with SEQ ID NO: 2 as input; (3) hybridizes to a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO: 40 under stringent conditions of a hybridization buffer comprising 20% formamide in 0.9M saline/0.09M SSC buffer at a temperature of 42°C; or (4) has a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO: 40.

2. The isolated nucleic acid molecule according to claim 1, wherein the nucleic acid molecule encodes a protein having an amino acid sequence selected from the group consisting of SEQ ID NOs: 2, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, and 41.

3. The isolated nucleic acid molecule according to claim 1, wherein the nucleic acid encodes a protein comprising a motif having an amino acid sequence corresponding to any of SEQ ID NOs: 3 to 18 or an amino acid sequence identified with a METAMEME software using the amino acid sequence of SEQ ID NO: 2 as input or an amino acid sequence identified as significantly similar to SEQ

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ID NO: 2 using a NCBI BLAST software (threshold= E less than or equal to 15) with SEQ ID NO: 2 as input.

4. The isolated nucleic acid molecule according to claim 1, wherein the nucleic acid molecule hybridizes to a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO: 40 under stringent conditions of a hybridization buffer comprising 20% formamide in 0.9M saline/0.09M SSC buffer at a temperature of 42°C.

5. The isolated nucleic acid molecule according to claim 1, wherein the nucleic acid molecule has a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO: 40.

6. The isolated nucleic acid molecule according to claim 1, wherein the nucleic acid molecule is from petunia, *Arabidopsis thaliana*, or rice.

7. A isolated protein encoded by the nucleic acid molecule according to claim 1.

8. The isolated protein according to claim 7, wherein the protein has an amino acid sequence selected from the group consisting of SEQ ID NOs: 2, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, and 41.

9. The isolated protein according to claim 7, wherein the protein encodes a protein comprising a motif having an amino acid sequence corresponding to any of SEQ ID NOs: 3 to 18 or an amino acid sequence identified with a METAMEME software using the amino acid sequence of SEQ ID NO: 2 as input or

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an amino acid sequence identified as significantly similar to SEQ ID NO: 2 using a NCBI BLAST software (threshold= E less than or equal to 15) with SEQ ID NO: 2 as input.

10. The isolated protein according to claim 7, wherein the nucleic acid molecule hybridizes to a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO: 40 under stringent conditions of a hybridization buffer comprising 20% formamide in 0.9M saline/0.09M SSC buffer at a temperature of 42°C.

11. The isolated protein according to claim 7, wherein the nucleic acid molecule has a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO: 40.

12. The isolated protein according to claim 7, wherein the nucleic acid molecule is from petunia, *Arabidopsis thaliana*, or rice.

13. An isolated expression system comprising the nucleic acid molecule of claim 1.

14. The isolated expression system according to claim 13, wherein the nucleic acid molecule is in proper sense orientation.

15. An isolated host cell comprising the nucleic acid molecule of claim 1.

16. The isolated host cell according to claim 15, wherein the nucleic acid molecule is in an expression system.

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17. The isolated host cell according to claim 15, wherein the host cell is a plant cell.

18. The isolated host cell according to claim 15, wherein the host cell is a bacterial cell.

19. A transgenic plant transformed with the nucleic acid molecule according to claim 1.

20. The transgenic plant according to claim 19, wherein the nucleic acid molecule encodes a protein having an amino acid sequence selected from the group consisting of SEQ ID NOs: 2, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, and 41.

21. The transgenic plant according to claim 19, wherein the nucleic acid encodes a protein comprising a motif having an amino acid sequence corresponding to any of SEQ ID NOs: 3 to 18 or an amino acid sequence identified with a METAMEME software using the amino acid sequence of SEQ ID NO: 2 as input or an amino acid sequence identified as significantly similar to SEQ ID NO: 2 using a NCBI BLAST software (threshold= E less than or equal to 15) with SEQ ID NO: 2 as input.

22. The transgenic plant according to claim 19, wherein the nucleic acid molecule hybridizes to a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO: 40 under stringent conditions of a hybridization buffer comprising 20% formamide in 0.9M saline/0.09M SSC buffer at a temperature of 42°C.

23. The transgenic plant according to claim 19, wherein the nucleic acid molecule has a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID

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NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO: 40.

24. The transgenic plant according to claim 19, wherein the nucleic acid molecule is from petunia, *Arabidopsis thaliana*, or rice.

25. The transgenic plant according to claim 19, wherein the transgenic plant is a crop plant.

26. The transgenic plant according to claim 25, wherein the crop plant is selected from the group consisting of alfalfa, rice, wheat, barley, rye, cotton, sunflower, peanut, corn, potato, sweet potato, bean, pea, chicory, lettuce, endive, cabbage, brussel sprout, beet, parsnip, turnip, cauliflower, broccoli, radish, spinach, onion, garlic, eggplant, pepper, celery, carrot, squash, pumpkin, zucchini, cucumber, apple, pear, melon, citrus, strawberry, grape, raspberry, pineapple, soybean, tobacco, tomato, sorghum, and sugarcane.

27. The transgenic plant according to claim 19, wherein the transgenic plant is an ornamental plant.

28. The transgenic plant according to claim 27, wherein the ornamental plant is selected from the group consisting of *Arabidopsis thaliana*, *Saintpaulia*, petunia, pelargonium, poinsettia, chrysanthemum, carnation, and zinnia.

29. A transgenic plant seed transformed with the nucleic acid molecule according to claim 1.

30. The transgenic plant seed according to claim 29, wherein the nucleic acid molecule encodes a protein having an amino acid sequence selected from the group consisting of SEQ ID NOS: 2, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, and 41.

31. The transgenic plant seed according to claim 29, wherein the nucleic acid encodes a protein comprising a motif having an amino acid sequence

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corresponding to any of SEQ ID NOs: 3 to 18 or an amino acid sequence identified with a METAMEME software using the amino acid sequence of SEQ ID NO: 2 as input or an amino acid sequence identified as significantly similar to SEQ ID NO: 2 using a NCBI BLAST software (threshold= E less than or equal to 15) with SEQ ID NO: 2 as input.

32. The transgenic plant seed according to claim 29, wherein the nucleic acid molecule hybridizes to a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO: 40 under stringent conditions of a hybridization buffer comprising 20% formamide in 0.9M saline/0.09M SSC buffer at a temperature of 42°C.

33. The transgenic plant seed according to claim 29, wherein the nucleic acid molecule has a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO: 40.

34. The transgenic plant seed according to claim 29, wherein the nucleic acid molecule is from petunia, *Arabidopsis thaliana*, or rice.

35. The transgenic plant seed according to claim 29, wherein the transgenic plant is a crop plant.

36. The transgenic plant seed according to claim 35, wherein the crop plant is selected from the group consisting of alfalfa, rice, wheat, barley, rye, cotton, sunflower, peanut, corn, potato, sweet potato, bean, pea, chicory, lettuce, endive, cabbage, brussel sprout, beet, parsnip, turnip, cauliflower, broccoli, radish, spinach, onion, garlic, eggplant, pepper, celery, carrot, squash, pumpkin, zucchini,

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cucumber, apple, pear, melon, citrus, strawberry, grape, raspberry, pineapple, soybean, tobacco, tomato, sorghum, and sugarcane.

37. The transgenic plant seed according to claim 29, wherein the transgenic plant is an ornamental plant.

38. The transgenic plant seed according to claim 37, wherein the ornamental plant is selected from the group consisting of *Arabidopsis thaliana*, *Saintpaulia*, petunia, pelargonium, poinsettia, chrysanthemum, carnation, and zinnia.

39. A method of restoring fertility to cytoplasmic male sterile plants comprising:

transforming a cytoplasmic male sterile plant with a nucleic acid molecule according to claim 1 under conditions effective to restore fertility to the cytoplasmic male sterile plant.

40. The method according to claim 39, wherein the nucleic acid molecule encodes a protein having an amino acid sequence selected from the group consisting of SEQ ID NOs: 2, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, and 41.

41. The method according to claim 39, wherein the nucleic acid encodes a protein comprising a motif having an amino acid sequence corresponding to any of SEQ ID NOs: 3 to 18 or an amino acid sequence identified with a METAMEME software using the amino acid sequence of SEQ ID NO: 2 as input or an amino acid sequence identified as significantly similar to SEQ ID NO: 2 using a NCBI BLAST software (threshold= E less than or equal to 15) with SEQ ID NO: 2 as input.

42. The method according to claim 39, wherein the nucleic acid molecule hybridizes to a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO:

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40 under stringent conditions of a hybridization buffer comprising 20% formamide in 0.9M saline/0.09M SSC buffer at a temperature of 42°C.

43. The method according to claim 39, wherein the nucleic acid molecule has a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO: 40.

44. The method according to claim 39, wherein the nucleic acid molecule is from petunia, *Arabidopsis thaliana*, or rice.

45. The method according to claim 39, wherein the plant is a crop plant.

46. The method according to claim 45, wherein the crop plant is selected from the group consisting of alfalfa, rice, wheat, barley, rye, cotton, sunflower, peanut, corn, potato, sweet potato, bean, pea, chicory, lettuce, endive, cabbage, brussel sprout, beet, parsnip, turnip, cauliflower, broccoli, radish, spinach, onion, garlic, eggplant, pepper, celery, carrot, squash, pumpkin, zucchini, cucumber, apple, pear, melon, citrus, strawberry, grape, raspberry, pineapple, soybean, tobacco, tomato, sorghum, and sugarcane.

47. The method according to claim 39, wherein the plant is an ornamental plant.

48. The method according to claim 47, wherein the ornamental plant is selected from the group consisting of *Arabidopsis thaliana*, *Saintpaulia*, petunia, pelargonium, poinsettia, chrysanthemum, carnation, and zinnia.

49. The method according to claim 39, wherein the plant has 2 or more copies of the nucleic acid molecule.

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50. A method of identifying a candidate plant suitable for breeding with a cytoplasmic male sterile plant comprising:

analyzing the candidate plant for the presence, in its genome, of a nucleic acid molecule according to claim 1.

51. The method according to claim 50, wherein the nucleic acid molecule encodes a protein having an amino acid sequence selected from the group consisting of SEQ ID NOs: 2, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, and 41.

52. The method according to claim 50, wherein the nucleic acid encodes a protein comprising a motif having an amino acid sequence corresponding to any of SEQ ID NOs: 3 to 18 or an amino acid sequence identified with a METAMEME software using the amino acid sequence of SEQ ID NO: 2 as input or an amino acid sequence identified as significantly similar to SEQ ID NO: 2 using a NCBI BLAST software (threshold= E less than or equal to 15) with SEQ ID NO: 2 as input.

53. The method according to claim 50, wherein the nucleic acid molecule hybridizes to a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO: 40 under stringent conditions of a hybridization buffer comprising 20% formamide in 0.9M saline/0.09M SSC buffer at a temperature of 42°C.

54. The method according to claim 50, wherein the nucleic acid molecule has a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO: 40.

55. The method according to claim 50, wherein the nucleic acid molecule is from petunia, *Arabidopsis thaliana*, or rice.

56. The method according to claim 50, wherein the plant is a crop plant.

57. The method according to claim 56, wherein the crop plant is selected from the group consisting of alfalfa, rice, wheat, barley, rye, cotton, sunflower, peanut, corn, potato, sweet potato, bean, pea, chicory, lettuce, endive, cabbage, brussel sprout, beet, parsnip, turnip, cauliflower, broccoli, radish, spinach, onion, garlic, eggplant, pepper, celery, carrot, squash, pumpkin, zucchini, cucumber, apple, pear, melon, citrus, strawberry, grape, raspberry, pineapple, soybean, tobacco, tomato, sorghum, and sugarcane.

58. The method according to claim 50, wherein the plant is an ornamental plant.

59. The method according to claim 58, wherein the ornamental plant is selected from the group consisting of *Arabidopsis thaliana*, *Saintpaulia*, petunia, pelargonium, poinsettia, chrysanthemum, carnation, and zinnia.

60. A method of identifying a candidate gene restoring fertility in plants, said method comprising:

analyzing the candidate gene for the presence of a nucleic acid molecule according to claim 1.

61. The method according to claim 60, wherein the nucleic acid molecule encodes a protein having an amino acid sequence selected from the group consisting of SEQ ID NOs: 2, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, and 41.

62. The method according to claim 60, wherein the nucleic acid encodes a protein comprising a motif having an amino acid sequence corresponding to any of SEQ ID NOs: 3 to 18 or an amino acid sequence identified with a METAMEME software using the amino acid sequence of SEQ ID NO: 2 as input or an amino acid sequence identified as significantly similar to SEQ ID NO: 2 using a

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NCBI BLAST software (threshold= E less than or equal to 15) with SEQ ID NO: 2 as input.

63. The method according to claim 60, wherein the nucleic acid molecule hybridizes to a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO: 40 under stringent conditions of a hybridization buffer comprising 20% formamide in 0.9M saline/0.09M SSC buffer at a temperature of 42°C.

64. The method according to claim 60, wherein the nucleic acid molecule has a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO: 40.

65. The method according to claim 60, wherein the nucleic acid molecule is from petunia, *Arabidopsis thaliana*, or rice.

66. The method according to claim 60, wherein the plant is a crop plant.

67. The method according to claim 66, wherein the crop plant is selected from the group consisting of alfalfa, rice, wheat, barley, rye, cotton, sunflower, peanut, corn, potato, sweet potato, bean, pea, chicory, lettuce, endive, cabbage, brussel sprout, beet, parsnip, turnip, cauliflower, broccoli, radish, spinach, onion, garlic, eggplant, pepper, celery, carrot, squash, pumpkin, zucchini, cucumber, apple, pear, melon, citrus, strawberry, grape, raspberry, pineapple, soybean, tobacco, tomato, sorghum, and sugarcane.

68. The method according to claim 60, wherein the plant is an ornamental plant.

69. The method according to claim 68, wherein the ornamental plant is selected from the group consisting of *Arabidopsis thaliana*, *Saintpaulia*, petunia, pelargonium, poinsettia, chrysanthemum, carnation, and zinnia.

70. A method of producing hybrid plant seed comprising:  
providing a cytoplasmic male sterile plant;  
providing a second plant comprising a nucleic acid molecule according to claim 1; and  
breeding the cytoplasmic male sterile plant and the second plant under conditions effective to produce hybrid progeny seed which yield fertile plants.

71. The method according to claim 70, wherein the nucleic acid molecule encodes a protein having an amino acid sequence selected from the group consisting of SEQ ID NOs: 2, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, and 41.

72. The method according to claim 70, wherein the nucleic acid encodes a protein comprising a motif having an amino acid sequence corresponding to any of SEQ ID NOs: 3 to 18 or an amino acid sequence identified with a METAMEME software using the amino acid sequence of SEQ ID NO: 2 as input or an amino acid sequence identified as significantly similar to SEQ ID NO: 2 using a NCBI BLAST software (threshold= E less than or equal to 15) with SEQ ID NO: 2 as input.

73. The method according to claim 70, wherein the nucleic acid molecule hybridizes to a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO: 40 under stringent conditions of a hybridization buffer comprising 20% formamide in 0.9M saline/0.09M SSC buffer at a temperature of 42°C.

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74. The method according to claim 70, wherein the nucleic acid molecule has a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO: 40.

75. The method according to claim 70, wherein the nucleic acid molecule is from petunia, *Arabidopsis thaliana*, or rice.

76. The method according to claim 70, wherein the plants are crop plants.

77. The method according to claim 76, wherein the crop plants are selected from the group consisting of alfalfa, rice, wheat, barley, rye, cotton, sunflower, peanut, corn, potato, sweet potato, bean, pea, chicory, lettuce, endive, cabbage, brussel sprout, beet, parsnip, turnip, cauliflower, broccoli, radish, spinach, onion, garlic, eggplant, pepper, celery, carrot, squash, pumpkin, zucchini, cucumber, apple, pear, melon, citrus, strawberry, grape, raspberry, pineapple, soybean, tobacco, tomato, sorghum, and sugarcane.

78. The method according to claim 70, wherein the plants are ornamental plants.

79. The method according to claim 78, wherein the ornamental plants are selected from the group consisting of *Arabidopsis thaliana*, *Saintpaulia*, petunia, pelargonium, poinsettia, chrysanthemum, carnation, and zinnia.

80. The hybrid progeny seed produced by the method according to claim 70.

81. The hybrid progeny plants grown from the hybrid progeny seed produced by the method according to claim 70.

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82. A method of producing plant seeds for an inbred line of plants comprising:

providing a cytoplasmic male sterile plant;

providing a second plant comprising a nucleic acid molecule according to claim 1;

breeding the cytoplasmic male sterile plant and the second plant under conditions effective to produce hybrid progeny seed which yield fertile plants;

producing hybrid fertile plants from the hybrid progeny seeds;

and

backcrossing the hybrid fertile plants and the second plant to produce seed which yield inbred progeny plants.

83. The method according to claim 82, wherein the nucleic acid molecule encodes a protein having an amino acid sequence selected from the group consisting of SEQ ID NOs: 2, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, and 41.

84. The method according to claim 82, wherein the nucleic acid encodes a protein comprising a motif having an amino acid sequence corresponding to any of SEQ ID NOs: 3 to 18 or an amino acid sequence identified with a METAMEME software using the amino acid sequence of SEQ ID NO: 2 as input or an amino acid sequence identified as significantly similar to SEQ ID NO: 2 using a NCBI BLAST software (threshold= E less than or equal to 15) with SEQ ID NO: 2 as input.

85. The method according to claim 82, wherein the nucleic acid molecule hybridizes to a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO: 40 under stringent conditions of a hybridization buffer comprising 20% formamide in 0.9M saline/0.09M SSC buffer at a temperature of 42°C.

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86. The method according to claim 82, wherein the nucleic acid molecule has a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO: 40.

87. The method according to claim 82, wherein the nucleic acid molecule is from petunia, *Arabidopsis thaliana*, or rice.

88. The method according to claim 82, wherein the plants are crop plants.

89. The method according to claim 88, wherein the crop plants are selected from the group consisting of alfalfa, rice, wheat, barley, rye, cotton, sunflower, peanut, corn, potato, sweet potato, bean, pea, chicory, lettuce, endive, cabbage, brussel sprout, beet, parsnip, turnip, cauliflower, broccoli, radish, spinach, onion, garlic, eggplant, pepper, celery, carrot, squash, pumpkin, zucchini, cucumber, apple, pear, melon, citrus, strawberry, grape, raspberry, pineapple, soybean, tobacco, tomato, sorghum, and sugarcane.

90. The method according to claim 82, wherein the plants are ornamental plants.

91. The method according to claim 90, wherein the ornamental plants are selected from the group consisting of *Arabidopsis thaliana*, *Saintpaulia*, petunia, pelargonium, poinsettia, chrysanthemum, carnation, and zinnia.

92. The seed which yield inbred progeny plants produced by the method according to claim 82.

93. The inbred progeny plants grown from the seed which yield inbred progeny plants produced by the method of claim 82.

94. A method of directing gene expression to plant mitochondria comprising:

transforming a plant with a chimeric nucleic acid molecule comprising a transgene operatively linked to a promoter or a terminator from a plant gene which restores fertility to cytoplasmic male sterile plants and modifies expression of toxic mitochondria proteins by the plant under conditions effective to direct expression of the transgene in the mitochondria of the transformed plant, wherein the promoter has a nucleotide sequence of from nucleotide 1 to 1981 of SEQ ID NO: 1 and the terminator has a nucleotide sequence of from nucleotide 3761 to 4593 of SEQ ID NO: 1.

95. The method according to claim 94, wherein the method is carried out with a promoter having a nucleotide sequence of from nucleotide 1 to 1981 of SEQ ID NO: 1.

96. The method according to claim 94, wherein the method is carried out with a terminator having a nucleotide sequence of from nucleotide 3761 to 4593 of SEQ ID NO: 1.

97. The method according to claim 94, wherein the plant is a crop plant.

98. The method according to claim 97, wherein the crop plant is selected from the group consisting of alfalfa, rice, wheat, barley, rye, cotton, sunflower, peanut, corn, potato, sweet potato, bean, pea, chicory, lettuce, endive, cabbage, brussel sprout, beet, parsnip, turnip, cauliflower, broccoli, radish, spinach, onion, garlic, eggplant, pepper, celery, carrot, squash, pumpkin, zucchini, cucumber, apple, pear, melon, citrus, strawberry, grape, raspberry, pineapple, soybean, tobacco, tomato, sorghum, and sugarcane.

99. The method according to claim 94, wherein the plant is an ornamental plant.

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100. The method according to claim 99, wherein the ornamental plant is selected from the group consisting of *Arabidopsis thaliana*, *Saintpaulia*, petunia, pelargonium, poinsettia, chrysanthemum, carnation, and zinnia.

101. A promoter from a plant gene which restores fertility to cytoplasmic male sterile plants and modifies expression of toxic mitochondria proteins by the plant under conditions effective to direct expression of the transgene in the mitochondria of the transformed plant, wherein the promoter has a nucleotide sequence of from nucleotide 1 to 1981 of SEQ ID NO: 1.

102. A terminator from a plant gene which restores fertility to cytoplasmic male sterile plants and modifies expression of toxic mitochondria proteins by the plant under conditions effective to direct expression of the transgene in the mitochondria of the transformed plant, wherein the terminator has a nucleotide sequence of from nucleotide 3761 to 4593 of SEQ ID NO: 1.

103. A nucleic acid construct comprising:  
a promoter or a terminator from a plant gene which restores fertility to cytoplasmic male sterile plants and modifies expression of toxic mitochondria proteins by the plant under conditions effective to direct expression of the transgene in the mitochondria of the transformed plant and  
a nucleic acid heterologous to and operatively coupled to the promoter or the terminator, wherein the promoter has a nucleotide sequence of from nucleotide 1 to 1981 of SEQ ID NO: 1 and the terminator has a nucleotide sequence of from nucleotide 3761 to 4593 of SEQ ID NO: 1.

104. A nucleic acid construct according to claim 103, wherein the construct is provided with a promoter from a plant gene which restores fertility to cytoplasmic male sterile plants and modifies expression of toxic mitochondria proteins by the plant under conditions effective to direct expression of the transgene in the mitochondria of the transformed plant, said promoter having a nucleotide sequence of from nucleotide 1 to 1981 of SEQ ID NO: 1.

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105. A nucleic acid construct according to claim 103, wherein the construct is provided with a terminator from a plant gene which restores fertility to cytoplasmic male sterile plants and modifies expression of toxic mitochondria proteins by the plant under conditions effective to direct expression of the transgene in the mitochondria of the transformed plant, said terminator having a nucleotide sequence of from nucleotide 3761 to 4593 of SEQ ID NO: 1.

106. An isolated expression system comprising the nucleic acid construct according to claim 103.

107. An isolated host cell comprising the nucleic acid construct according to claim 103.

108. A transgenic plant comprising the nucleic acid construct according to claim 103.

109. A transgenic plant seed comprising the nucleic acid construct according to claim 103.

110. A method of expressing a gene preferentially in roots of a plant comprising:

transforming a plant with a nucleic acid construct comprising: a promoter suitable for driving expression preferentially in roots having a nucleotide sequence of from 1 to 1388 of SEQ ID NO: 44; a nucleic acid heterologous to the promoter, wherein the promoter is operatively coupled 5' to the nucleic acid to induce transcription of the nucleic acid; and a terminator having a nucleotide sequence of from nucleotide 3168 to 4016 of SEQ ID NO: 44, wherein the terminator is operably coupled 3' to the nucleic acid.

111. The method according to claim 110, wherein the plant is a crop plant.

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112. The method according to claim 111, wherein the crop plant is selected from the group consisting of alfalfa, rice, wheat, barley, rye, cotton, sunflower, peanut, corn, potato, sweet potato, bean, pea, chicory, lettuce, endive, cabbage, brussel sprout, beet, parsnip, turnip, cauliflower, broccoli, radish, spinach, onion, garlic, eggplant, pepper, celery, carrot, squash, pumpkin, zucchini, cucumber, apple, pear, melon, citrus, strawberry, grape, raspberry, pineapple, soybean, tobacco, tomato, sorghum, and sugarcane.

113. The method according to claim 111, wherein the plant is an ornamental plant.

114. The method according to claim 113, wherein the ornamental plant is selected from the group consisting of *Arabidopsis thaliana*, *Saintpaulia*, petunia, pelargonium, poinsettia, chrysanthemum, carnation, and zinnia.

115. A method of altering plant floral morphology in ornamental plants comprising:

transforming an ornamental plant with a nucleic acid molecule according to claim 1.

116. The method according to claim 115, wherein the nucleic acid molecule encodes a protein having an amino acid sequence selected from the group consisting of SEQ ID NOs: 2, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, and 41.

117. The method according to claim 115, wherein the nucleic acid encodes a protein comprising a motif having an amino acid sequence corresponding to any of SEQ ID NOs: 3 to 18 or an amino acid sequence identified with a METAMEME software using the amino acid sequence of SEQ ID NO: 2 as input or an amino acid sequence identified as significantly similar to SEQ ID NO: 2 using a NCBI BLAST software (threshold= E less than or equal to 15) with SEQ ID NO: 2 as input.

118. The method according to claim 115, wherein the nucleic acid molecule hybridizes to a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO: 40 under stringent conditions of a hybridization buffer comprising 20% formamide in 0.9M saline/0.09M SSC buffer at a temperature of 42°C.

119. The method according to claim 115, wherein the nucleic acid molecule has a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO: 40.

120. The method according to claim 115, wherein the nucleic acid molecule is from petunia, *Arabidopsis thaliana*, or rice.

121. The method according to claim 115, wherein the plant is an ornamental plant.

122. The method according to claim 121, wherein the ornamental plant is selected from the group consisting of *Arabidopsis thaliana*, *Saintpaulia*, petunia, pelargonium, poinsettia, chrysanthemum, carnation, and zinnia.

123. A method of producing plants with a cytoplasmic male sterile plant restoration system comprising:

transforming a first plant in its chloroplast genome with a nucleic acid which causes the plant to become male sterile;

transforming a second plant with a nucleic acid molecule according to claim 1 whose protein product is targeted to the chloroplast; and  
crossing the first and second plants to produce progeny plants possessing a cytoplasmic male sterile plant restoration system.

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124. The method according to claim 123, wherein the nucleic acid molecule encodes a protein having an amino acid sequence selected from the group consisting of SEQ ID NOs: 2, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, and 41.

125. The method according to claim 123, wherein the nucleic acid molecule encodes a protein comprising a motif having an amino acid sequence corresponding to any of SEQ ID NOs: 3 to 18 or an amino acid sequence identified with a METAMEME software using the amino acid sequence of SEQ ID NO: 2 as input or an amino acid sequence identified as significantly similar to SEQ ID NO: 2 using a NCBI BLAST software (threshold= E less than or equal to 15) with SEQ ID NO: 2 as input.

126. The method according to claim 123, wherein the nucleic acid molecule hybridizes to a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO: 40 under stringent conditions of a hybridization buffer comprising 20% formamide in 0.9M saline/0.09M SSC buffer at a temperature of 42°C.

127. The method according to claim 123, wherein the nucleic acid molecule has a nucleotide sequence selected from the group consisting of from nucleotide 1982 to 3760 of SEQ ID NO: 1, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, and SEQ ID NO: 40.

128. The method according to claim 123, wherein the nucleic acid molecule is from petunia, *Arabidopsis thaliana*, or rice.

129. A method of producing plants with a cytoplasmic male sterile plant restoration system comprising:

mutagenizing a first plant having a nucleic acid which encodes a protein comprising a motif having an amino acid sequence corresponding to any of

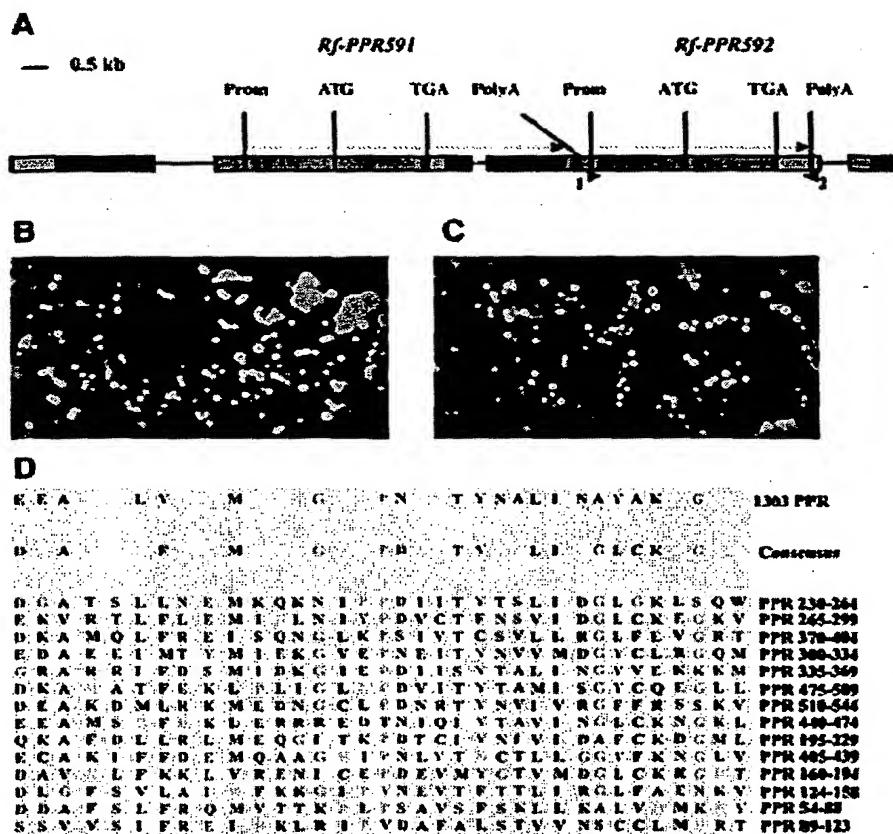
- 93 -

SEQ ID NOs: 3 to 18 or an amino acid sequence identified with a METAMEME software using the amino acid sequence of SEQ ID NO: 2 as input or an amino acid sequence identified as significantly similar to SEQ ID NO: 2 using a NCBI BLAST software (threshold= E less than or equal to 15) with SEQ ID NO: 2 as input;

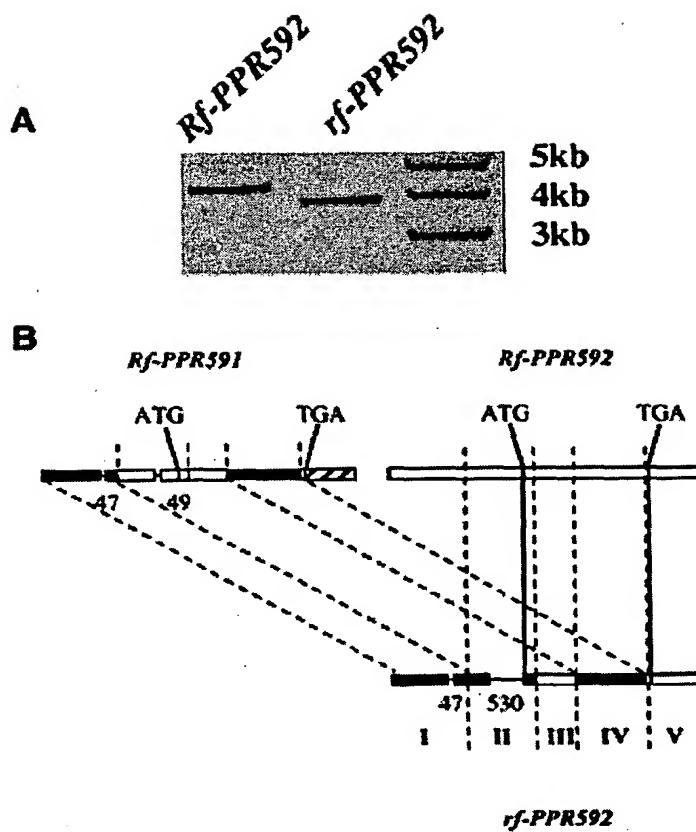
crossing the mutagenized first plant with a wild-type plant having mitochondrial DNA polymorphisms compared to mitochondrial DNA in the mutagenized first plant to produce progeny plants; and

determining if the progeny plants are fertile, whereby fertile progeny plants can be used as a fertile maintainer line, wherein the mutagenized first plant, the fertile maintainer line, and a wild-type allele present in the first plant before mutagenesis comprises a new cytoplasmic male sterile plant restoration system.

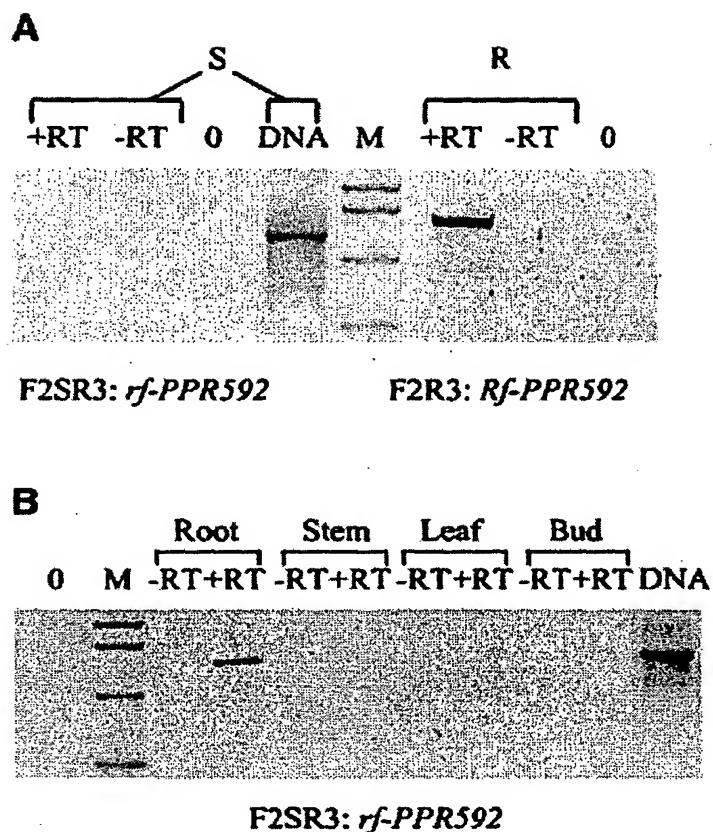
130. An isolated nucleic acid sequence corresponding to SEQ ID NO: 42 or SEQ ID NO: 44.



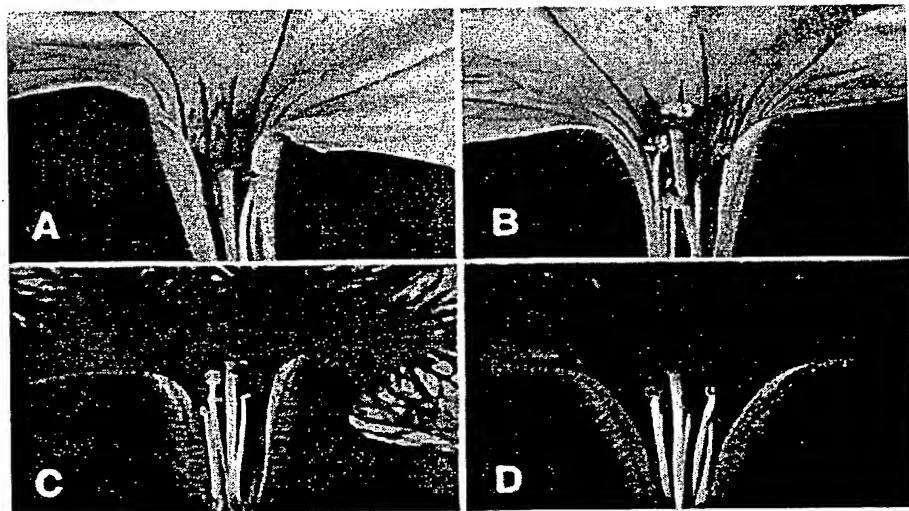
**Figure 1**



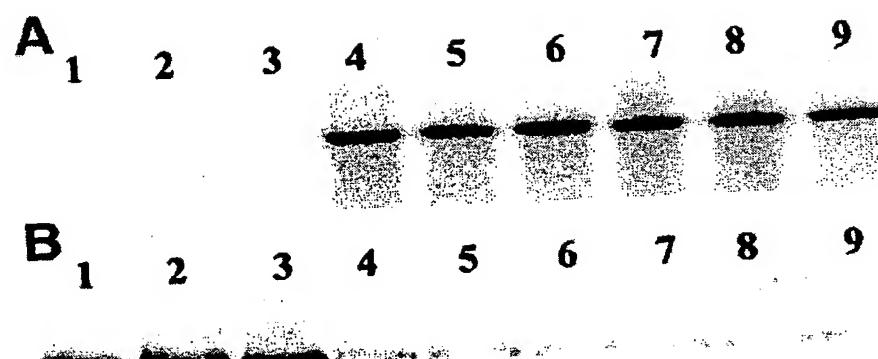
**Figure 2**



**Figure 3**

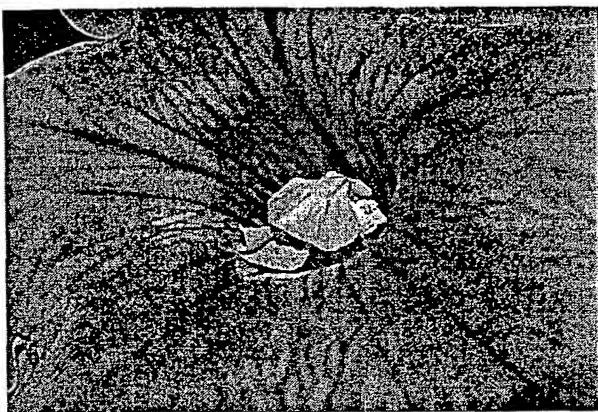


**Figure 4**



**Figure 5**

A.

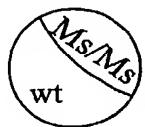
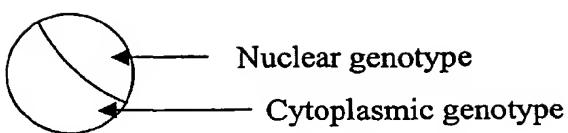


B.



**Figure 6**

## Diagram of a plant genotype



Wild-type plant with wild-type nuclear alleles  
of gene Ms, a PPR motif gene needed for male  
fertility



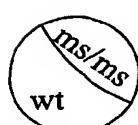
Mutagenesis of the Ms gene (e.g by an insertional element, radiation,  
chemicals, etc.)



Heterozygous mutant at Ms locus



Self-cross



Mutant male sterile plant



By sexual crosses or cybridization with genotypes carrying different  
organelle genomes than the initial wt genome, or by mutagenesis of  
genome(s) in the wt cytoplasm, create a genotype with mutant  
ms/ms nuclear alleles that is male fertile



Male fertile plant

These genotypes are then utilized as a CMS/restorer system for hybrid seed production and breeding as follows:



CMS line



Fertile maintainer line



Fertility restorer lines

**Figure 7A**

Examples illustrating how new lines can be used as a CMS/restorer system

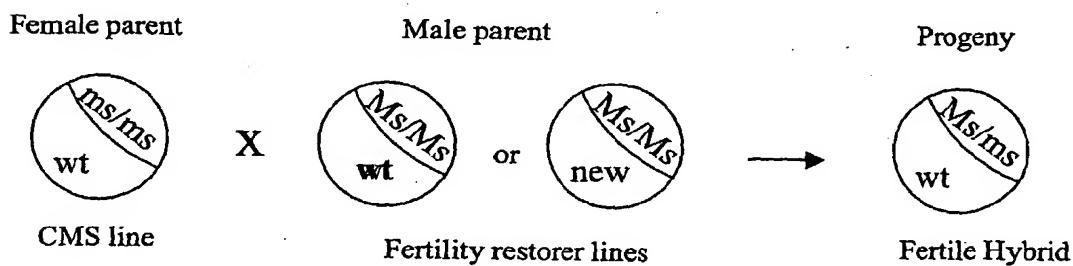
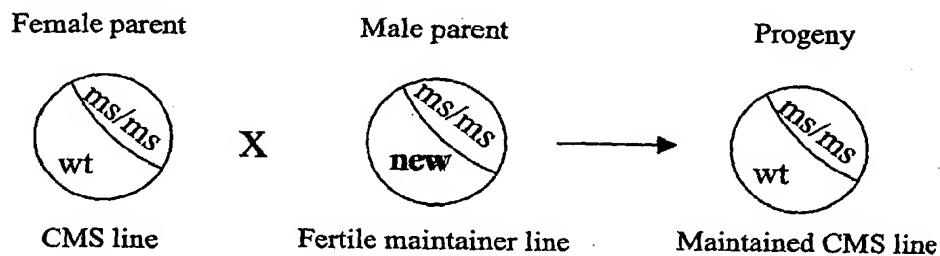
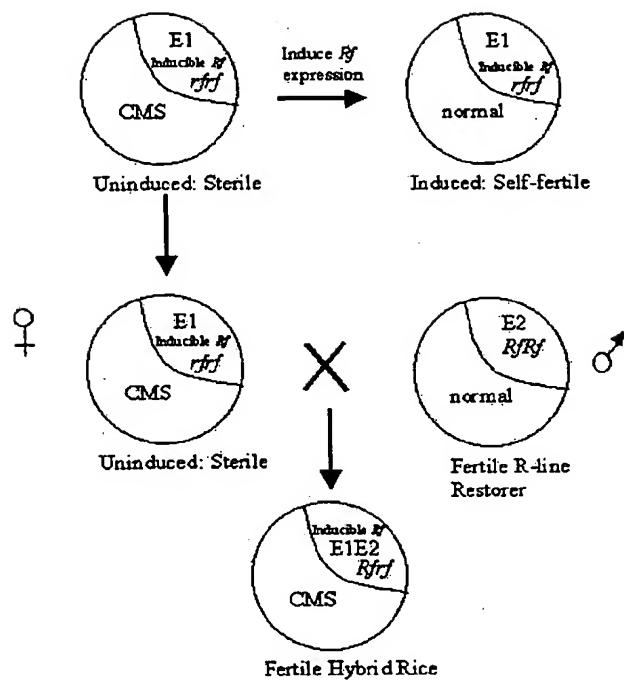


Figure 7B



**Figure 8**

## SEQUENCE LISTING

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<120> GENES FOR ALTERING MITOCHONDRIAL FUNCTION AND FOR  
HYBRID SEED PRODUCTION

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<151> 2002-01-10

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<170> PatentIn Ver. 2.1

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<211> 4593

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 <212> PRT  
 <213> Petunia sp.

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 Cys Ser Ile Ser Val Lys Gly Asn Phe Gly Val Ser Asn Glu Phe Glu  
 35 40 45  
 Asn Val Lys Cys Leu Asp Asp Ala Phe Ser Leu Phe Arg Gln Met Val  
 50 55 60  
 Thr Thr Lys Pro Leu Pro Ser Ala Val Ser Phe Ser Lys Leu Leu Lys  
 65 70 75 80  
 Ala Leu Val His Met Lys His Tyr Ser Ser Val Val Ser Ile Phe Arg  
 85 90 95  
 Glu Ile His Lys Leu Arg Ile Pro Val Asp Ala Phe Ala Leu Ser Thr  
 100 105 110  
 Val Val Asn Ser Cys Cys Leu Met His Arg Thr Asp Leu Gly Phe Ser  
 115 120 125  
 Val Leu Ala Ile His Phe Lys Lys Gly Ile Pro Tyr Asn Glu Val Thr  
 130 135 140  
 Phe Thr Thr Leu Ile Arg Gly Leu Phe Ala Glu Asn Lys Val Lys Asp  
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 Ala Val His Leu Phe Lys Lys Leu Val Arg Glu Asn Ile Cys Glu Pro  
 165 170 175  
 Asp Glu Val Met Tyr Gly Thr Val Met Asp Gly Leu Cys Lys Lys Gly

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His Thr Gln Lys Ala Phe Asp Leu Leu Arg Leu Met Glu Gln Gly Ile		
195	200	205
Thr Lys Pro Asp Thr Cys Ile Tyr Asn Ile Val Ile Asp Ala Phe Cys		
210	215	220
Lys Asp Gly Met Leu Asp Gly Ala Thr Ser Leu Leu Asn Glu Met Lys		
225	230	235
Gln Lys Asn Ile Pro Pro Asp Ile Ile Thr Tyr Thr Ser Leu Ile Asp		
245	250	255
Gly Leu Gly Lys Leu Ser Gln Trp Glu Lys Val Arg Thr Leu Phe Leu		
260	265	270
Glu Met Ile His Leu Asn Ile Tyr Pro Asp Val Cys Thr Phe Asn Ser		
275	280	285
Val Ile Asp Gly Leu Cys Lys Glu Gly Lys Val Glu Asp Ala Glu Glu		
290	295	300
Ile Met Thr Tyr Met Ile Glu Lys Gly Val Glu Pro Asn Glu Ile Thr		
305	310	315
Tyr Asn Val Val Met Asp Gly Tyr Cys Leu Arg Gly Gln Met Gly Arg		
325	330	335
Ala Arg Arg Ile Phe Asp Ser Met Ile Asp Lys Gly Ile Glu Pro Asp		
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Ile Ile Ser Tyr Thr Ala Leu Ile Asn Gly Tyr Val Glu Lys Lys Lys		
355	360	365
Met Asp Lys Ala Met Gln Leu Phe Arg Glu Ile Ser Gln Asn Gly Leu		
370	375	380
Lys Pro Ser Ile Val Thr Cys Ser Val Leu Leu Arg Gly Leu Phe Glu		
385	390	395
Val Gly Arg Thr Glu Cys Ala Lys Ile Phe Phe Asp Glu Met Gln Ala		
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Ala Gly His Ile Pro Asn Leu Tyr Thr His Cys Thr Leu Leu Gly Gly		
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Tyr Phe Lys Asn Gly Leu Val Glu Glu Ala Met Ser His Phe His Lys		

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Leu Glu Arg Arg Arg Glu Asp Thr Asn Ile Gln Ile Tyr Thr Ala Val  
450                    455                    460  
  
Ile Asn Gly Leu Cys Lys Asn Gly Lys Leu Asp Lys Ala His Ala Thr  
465                    470                    475                    480  
  
Phe Glu Lys Leu Pro Leu Ile Gly Leu His Pro Asp Val Ile Thr Tyr  
485                    490                    495  
  
Thr Ala Met Ile Ser Gly Tyr Cys Gln Glu Gly Leu Leu Asp Glu Ala  
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Lys Asp Met Leu Arg Lys Met Glu Asp Asn Gly Cys Leu Pro Asp Asn  
515                    520                    525  
  
Arg Thr Tyr Asn Val Ile Val Arg Gly Phe Phe Arg Ser Ser Lys Val  
530                    535                    540  
  
Ser Glu Met Lys Ala Phe Leu Lys Glu Ile Ala Gly Lys Ser Phe Ser  
545                    550                    555                    560  
  
Phe Glu Ala Ala Thr Val Glu Leu Leu Met Asp Ile Ile Ala Glu Asp  
565                    570                    575  
  
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Peeters

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<220>  
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Asn Lys Val  
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Pro Asp Glu Val Met Tyr Gly Thr Val Met Asp Gly Leu Cys Lys Lys  
20 25 30

Gly His Thr  
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Gly Met Leu  
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Pro Asn Glu Ile Thr Tyr Asn Val Val Met Asp Gly Tyr Cys Leu Arg  
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Gly Gln Met  
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<400> 13  
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Lys Lys Met  
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<210> 14  
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<400> 14  
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Gly Lys Leu  
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<212> PRT  
<213> Petunia sp.

<400> 17  
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<210> 18  
<211> 35  
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<213> Oryza sativa

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<211> 878

<212> PRT

<213> Oryza sativa

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Gly	Ser	Gly	Ala	Glu	Asp	Ala	Arg	His	Val	Phe	Asp	Glu	Leu	Leu	Arg
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Arg	Gly	Arg	Gly	Ala	Ser	Ile	Tyr	Gly	Leu	Asn	Arg	Ala	Leu	Ala	Asp
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Val	Ala	Arg	His	Ser	Pro	Ala	Ala	Ala	Val	Ser	Arg	Tyr	Asn	Arg	Met
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Ala	Arg	Ala	Gly	Ala	Gly	Lys	Val	Thr	Pro	Thr	Val	His	Thr	Tyr	Ala
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Ile	Leu	Ile	Gly	Cys	Cys	Cys	Arg	Ala	Gly	Arg	Leu	Asp	Leu	Gly	Phe
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Ala	Ala	Leu	Gly	Asn	Val	Val	Lys	Lys	Gly	Phe	Arg	Val	Asp	Ala	Ile
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Thr	Phe	Thr	Pro	Leu	Leu	Lys	Gly	Leu	Cys	Ala	Asp	Lys	Arg	Thr	Ser
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Asp	Ala	Met	Asp	Ile	Val	Leu	Arg	Arg	Met	Thr	Glu	Leu	Gly	Cys	Ile
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Pro	Asp	Val	Phe	Ser	Tyr	Asn	Asn	Leu	Leu	Lys	Gly	Leu	Cys	Asp	Glu
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195 200 205

Asn Gly Phe Phe Lys Glu Gly Asp Ser Asp Lys Ala Tyr Ser Thr Tyr  
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Ser Ile Ile Ala Ala Leu Cys Lys Ala Gln Ala Met Asp Lys Ala Met  
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Glu Val Leu Asn Thr Met Val Lys Asn Gly Val Met Pro Asp Cys Met  
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Thr Tyr Asn Ser Ile Leu His Gly Tyr Cys Ser Ser Gly Gln Pro Lys  
275 280 285

Glu Ala Ile Gly Thr Leu Lys Lys Met Arg Ser Asp Gly Val Glu Pro  
290 295 300

Asn Val Val Thr Tyr Ser Ser Leu Met Asn Tyr Leu Cys Lys Asn Gly  
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Arg Ser Thr Glu Ala Arg Lys Ile Phe Asp Ser Met Thr Lys Arg Gly  
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Thr Lys Gly Ala Leu Val Glu Met His Ala Leu Leu Asp Leu Met Val  
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Arg Asn Gly Ile Gln Pro Asp His His Val Phe Asn Ile Leu Ile Cys  
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Ala Tyr Ala Lys Gln Glu Lys Val Asp Gln Ala Met Leu Val Phe Ser  
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Val Ile Glu Ser Glu Lys Leu Phe Asp Leu Met Val Arg Ile Gly Val  
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Asp Glu Lys His Phe Ser Leu Glu Ala Ser Thr Ala Ser Phe Leu Leu  
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Glu Ser Ser Pro Ile Val Trp Glu Gln Ile Ser Arg Ile Ser His Leu  
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Ser Val Asn Leu Lys Leu Ile Lys Gln Pro Lys Cys Thr Cys Glu Leu  
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Gly Pro Lys Trp Ser Gln Asn Leu Pro Lys Pro Gly Thr Asn Ser Val  
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Gly Ser Val Ala Gln Phe His Leu Ser Arg Gly Gly Tyr Arg Ala Tyr  
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Arg Gly Gly Thr Thr Val Thr Ala Leu Pro Gln Gly Asp Gly Asn Pro  
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<211> 3660

<212> DNA

<213> Oryza sativa

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<211> 1219

<212> PRT

<213> Oryza sativa

<400> 23

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Ala Arg His Ser Pro Ala Ala Val Ser Arg Tyr Asn Arg Met Ala  
 65 70 75 80

Arg Ala Gly Ala Asp Glu Val Thr Pro Asn Leu Cys Thr Tyr Gly Ile  
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Ala Leu Gly Asn Val Ile Lys Lys Gly Phe Arg Val Asp Ala Ile Ala  
 115 120 125

Phe Thr Pro Leu Leu Lys Gly Leu Cys Ala Asp Lys Arg Thr Ser Asp  
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Arg Gln Gln Gly Leu Asn Pro Asp Thr Val Thr Tyr Gly Thr Val Ile		
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Glu Ser Glu Lys Leu Phe Asp Leu Met Val Arg Ile Gly Val Lys Pro		
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Asp Ile Ile Thr Tyr Ser Thr Leu Ile Asp Gly Tyr Cys Leu Ala Gly		
515	520	525
Lys Met Asp Glu Ala Thr Lys Leu Leu Ala Ser Met Val Ser Val Gly		
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Met Lys Pro Asp Cys Val Thr Tyr Asn Thr Leu Ile Asn Gly Tyr Cys		
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Lys Ile Ser Arg Met Glu Asp Ala Leu Val Leu Phe Arg Glu Met Glu		
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Ser Ser Gly Val Ser Pro Asp Ile Ile Thr Tyr Asn Ile Ile Leu Gln		
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Gly Leu Phe Gln Thr Arg Arg Thr Ala Ala Ala Lys Glu Leu Tyr Val		
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Gly Ile Thr Glu Ser Gly Thr Gln Leu Glu Leu Ser Thr Tyr Asn Ile		
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Ile Leu His Gly Leu Cys Lys Asn Asn Leu Thr Asp Glu Ala Leu Arg		
625	630	635
Met Phe Gln Asn Leu Cys Leu Thr Asp Leu Gln Leu Glu Thr Arg Thr		
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Phe Asn Ile Met Ile Gly Ala Leu Leu Lys Val Gly Arg Asn Asp Glu		
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Ala Lys Asp Leu Phe Ala Ala Leu Ser Ala Asn Gly Leu Val Pro Asp		

675	680	685
Val Arg Thr Tyr Ser Leu Met Ala Glu Asn Leu Ile Glu Gln Gly Leu		
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Leu Glu Glu Leu Asp Asp Leu Phe Leu Ser Met Glu Glu Asn Gly Cys		
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Thr Ala Asn Ser Arg Met Leu Asn Ser Ile Val Arg Lys Leu Leu Gln		
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Arg Gly Asp Ile Thr Arg Ala Gly Thr Tyr Leu Phe Met Ile Asp Glu		
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Lys His Phe Ser Leu Glu Ala Ser Thr Ala Scr Leu Phe Leu Asp Leu		
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Leu Ser Gly Gly Lys Tyr Gln Glu Tyr His Ser Cys Ile Arg Gly Gly		
770	775	780
Ile Phe Ser Leu Cys Val Asn Ser Glu Val Gln Glu Asn His Leu Leu		
785	790	795
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Val Asn Leu Val Asp Ser Lys Ala Pro Ser Ile Gly Ser Lys Leu Leu		
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Gly Ile Ser Lys Val Gln Met Leu Asn Gly Ser Asn Lys Asp Ser Asp		
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Cys Ile Ser Glu Glu Ile Leu Ser Lys Val Glu Glu Ile Leu Leu Ser		
850	855	860
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885	890	895
Cys Leu Ser Ala Val Ser Val Glu Glu Thr Ser Asp Thr Val Ser Arg		
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Val Gly Gly Asn Phe Lys Glu Thr Leu Arg Glu Met Gly Gly Leu Asp		
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Ser Ile Phe Asp Val Met Val Asp Phe His Ser Thr Leu Glu Asn Leu		

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Leu Gln Ser Ala Ala Leu Leu Leu Lys Cys Leu Lys Ile Leu Glu Asn		
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Ala Ile Phe Leu Ser Asp Asp Asn Lys Thr His Leu Leu Asn Met Ser		
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Arg Lys Leu Asn Pro Lys Arg Ser Leu Leu Ser Phe Val Gly Val Ile		
995	1000	1005
Ile Asn Thr Ile Glu Leu Leu Ser Ala Leu Ser Ile Leu Gln Asn Ser		
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Ser Val Val Ser Ser Ser Thr Tyr Pro Lys Ser Ser Lys Val Ser Gln		
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Gln Ser Tyr Ser Val Val Met Ala Gly Gly Asp Arg Gly Arg Gly Val		
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Glu Cys His Pro His Gln Gly Val Ser Ala Ala Leu Leu Arg Pro Gly		
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Pro Gln Ala Leu Ala Ala Ser Trp Arg Arg Arg Glu Thr Val Val Arg		
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Ser Asp Phe Ala Ala Gly Gly Val Ala Thr Met Gly Asp Ser Pro Gln		
1090	1095	1100
Ala Leu Ser Asp Arg Leu Cys Gly Ser Ala Thr Lys Val Trp Arg Gly		
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Gly Ala Glu Trp Thr Ala Glu Ala Phe Ala Arg Asn Gly Ala Ala Gly		
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Pro Ser Gln Ser Arg Leu Pro Val Thr Arg Asn Arg Ala Gln His Tyr		
1140	1145	1150
Ile Ala Lys Ile Trp Ala Thr Leu Thr Ile Ser Met Phe Tyr Lys Leu		
1155	1160	1165
Asp Val Glu Gly Met Glu Asn Leu Pro Pro Asn Ser Ser Pro Ala Ile		
1170	1175	1180
Tyr Val Ala Asn His Gln Ser Phe Leu Asp Ile Tyr Thr Leu Leu Thr		

1185                    1190                    1195                    1200  
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Arg Ile Ile

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35	40	45	
Ser Ile Tyr Gly Leu Asn Arg Ala Leu Ala Asp Val Ala Arg His Ser			
50	55	60	
Pro Ala Ala Ala Val Ser Arg Tyr Asn Arg Met Ala Arg Ala Gly Ala			
65	70	75	80
Asp Glu Val Thr Pro Asp Leu Cys Thr Tyr Gly Ile Leu Ile Gly Cys			
85	90	95	
Cys Cys Arg Ala Gly Arg Leu Asp Leu Gly Phe Ala Ala Leu Gly Asn			
100	105	110	
Val Ile Lys Lys Gly Phe Arg Val Glu Ala Ile Thr Phe Thr Pro Leu			
115	120	125	
Leu Lys Gly Leu Cys Ala Asp Lys Arg Thr Ser Asp Ala Met Asp Ile			

130	135	140
Val Leu Arg Arg Met Thr Glu Leu Gly Cys Ile Pro Asn Val Phe Ser		
145	150	155
Tyr Asn Asn Leu Leu Asn Gly Leu Cys Asp Glu Asn Arg Ser Gln Glu		
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Ala Leu Glu Leu Leu His Met Met Ala Asp Asp Arg Gly Gly Ser		
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Pro Pro Asp Val Val Ser Tyr Thr Val Ile Asn Gly Phe Phe Lys		
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Glu Gly Asp Ser Asp Lys Ala Tyr Ser Thr Tyr His Glu Met Leu Asp		
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Arg Gly Ile Leu Pro Asp Val Val Thr Tyr Ser Ser Ile Ile Ala Ala		
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Leu Cys Lys Gly Gln Ala Met Asp Lys Pro Trp Ser His Cys Lys Glu		
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Gly Arg Val Ile Glu Ser Glu Lys Leu Phe Asp Leu Met Val Arg Ile		
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Gly Val Lys Pro Asp Ile Ile Thr Tyr Ser Thr Leu Ile Asp Gly Tyr		
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Cys Leu Ala Gly Lys Met Asp Glu Ala Met Lys Leu Leu Ser Gly Met		
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Asn Gly Tyr Cys Lys Ile Ser Arg Met Glu Asp Ala Leu Val Leu Phe		
325	330	335
Lys Glu Met Glu Ser Ser Gly Val Ser Pro Asp Ile Ile Thr Tyr Asn		
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Ile Ile Leu Gln Gly Leu Phe Gln Thr Arg Arg Thr Ala Ala Ala Lys		
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Glu Leu Tyr Val Arg Ile Thr Glu Ser Gly Thr Gln Ile Glu Leu Ser		
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Thr Tyr Asn Ile Ile Leu His Gly Leu Cys Lys Asn Lys Leu Thr Asp		

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405		410	415
Glu Ala Arg Thr Phe Asn Ile Met Ile Asp Ala Leu Leu Lys Val Gly			
420		425	430
Arg Asn Asp Glu Ala Lys Asp Leu Phe Val Ala Phe Ser Ser Asn Gly			
435		440	445
Leu Val Pro Asn Tyr Trp Thr Tyr Arg Leu Met Ala Glu Asn Ile Ile			
450		455	460
Gly Gln Gly Leu Leu Glu Glu Leu Asp Gln Leu Phe Leu Ser Met Glu			
465		470	475
Asp Asn Gly Cys Thr Val Asp Ser Gly Met Leu Asn Phe Ile Val Arg			
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Glu Leu Leu Gln Arg Gly Val Val Val Val Val Ser Gly Glu Ser Ala			
500		505	510
Thr Thr Pro Pro Pro Thr Leu Lys Ile Leu Thr Cys Gly Ile Thr Val			
515		520	525
Asn Pro Phe Ser Lys Thr Cys Gly Ile Thr Val Asn Pro Phe Ser Lys			
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Pro Ile Val Gln Thr Gly Ala Cys Gly Gln Val Lys Glu Val Gly Lys			
545		550	555
Asn Ala Ser Glu Glu Arg Leu Ile Val Val Ser Ser Gln Glu Ile Pro			
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Lys Ala Ser Thr Leu Ala Glu Asn His Gln Leu Thr Thr Arg Leu Val			
595		600	605
Val Pro Ser Asn Lys Val Gly Cys Ile Leu Gly Glu Gly Lys Val			
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Ile Thr Glu Met Arg Arg Arg Thr Gly Ala Glu Ile Arg Val Tyr Ser			
625		630	635
Lys Ala Asp Lys Pro Lys Tyr Leu Ser Phe Asp Glu Glu Leu Val Gln			

645	650	655
His Ile Ser Leu Ile Leu Val Asp Arg His Ala Gly Arg Ala His Leu		
660	665	670
Leu Ser His Gln Leu Leu Thr Ala Ile Tyr Val Leu Val Leu Asn Arg		
675	680	685
Ser Ile Val Val Ala Glu Val Lys Asn Asn His Gly Thr Ala Ala Cys		
690	695	700
Trp Ala Leu Ala Ala Ile Ser Tyr Asn Arg Thr Asn Phe Ser Asp Leu		
705	710	715
720		
Leu Val Ser His Trp Phe Ile Ile Lys Ala Ser Val Phe Ser Phe Thr		
725	730	735
Leu Gly Leu Gln Asn Leu Arg Thr Gly Pro Ala His Asp Pro Tyr Thr		
740	745	750
Val Tyr Pro Val Glu Tyr Phe Ser Lys Arg Glu Tyr Pro Ser Gly Ser		
755	760	765
Ser Lys Val Ala Pro Ser Ala Ser Tyr Glu Arg Tyr Ala Ala Thr Thr		
770	775	780
Arg Leu Pro Asn Gly Glu Leu Pro Ser Ser Ile Ser Pro Gly Ala Asp		
785	790	795
800		
Tyr Met Ser Cys Arg Ser Tyr Leu Asp Gln Val Pro Thr Asp Arg Tyr		
805	810	815
Ser Asn Arg Val Thr Leu Gln Leu Gly Leu Ser Arg Ala Gly Asn Ser		
820	825	830
Asn Val Gln Gln Leu Gly Ile Thr Arg Ala Gly Asn Ser Asn Ala Tyr		
835	840	845
Asp Tyr Thr Glu Ala Ala Glu Gln Ile His Gly Arg Glu Asp Tyr Arg		
850	855	860
Arg Leu Ser Gly Leu Thr Gly Tyr Pro Gly Gly Ser Ser Asn Cys Gly		
865	870	875
880		
Phe Gln Ile Val Asn Trp Ser Leu Ser Leu Val Leu Val Ile Ser Gly		
885	890	895
Ala Arg Val Lys Leu His Glu Ala His Pro Gly Ser Ser Glu Ser Ile		

900	905	910
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Val Glu Ile Gln Gly Ile Pro Asp Gln Val Lys Ala Ala Gln Ser Leu		
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Leu Gln Gly Phe Ile Gly Ala Ser Ser Asn Ser Arg Gln Ala Pro Gln		
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Ser Ser Arg Met Ala His Tyr Phe		
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<211> 1737

<212> DNA

<213> Oryza sativa

<400> 26

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<400> 27

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Leu Val Gln Ala Leu Thr Gly Ala Ala Thr Ala Ala Ala His Arg

20				25				30						
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Leu Leu His Leu Leu Leu Arg Thr Ala Pro Pro Pro Pro Leu Pro Asp

35				40				45					
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Leu Val Ser Leu Ala Arg Trp Ser Arg Ala His Phe Arg Ala Pro Leu

50				55				60				
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Pro Leu Arg Leu His Gly Leu Leu Leu Ala Arg Leu Ala Ser Lys Gly

65				70				75				80
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Leu Tyr Pro Leu Leu Arg Ser Glu Leu His Val Leu Ala Ala Ala Arg

85				90				95			
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Leu His Ser Pro Ala Ser Ile Leu Arg Ala Leu Pro Ser Pro Ser Ala

100				105				110			
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Ser Ala Ser Ala Ser Thr Pro Leu Ile Ala Asp Met Leu Val Leu Ala

115				120				125			
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Leu Ala Arg Ala Ser Gln Pro Leu Arg Ala Tyr Asp Ala Phe Leu Leu

130				135				140			
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Ala Gly Glu Ser His Pro Arg His Arg Pro Ser Thr Ser Ser Val Asn

145				150				155			160
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Ala Leu Leu Ala Gly Leu Val Gly Ala Lys Arg Val Asp Leu Ala Glu

165				170				175		
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Lys Ala Phe Arg Ser Ala Leu Arg Arg Arg Val Ser Pro Asp Ile Tyr

180				185				190		
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Thr Phe Asn Thr Val Ile Ser Gly Leu Cys Arg Ile Gly Gln Leu Arg

195				200				205		
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Lys Ala Gly Asp Val Ala Lys Asp Ile Lys Ala Trp Gly Leu Ala Pro

210				215				220		
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 260 265 270

Gly Tyr Cys Lys Asn Ser Asn Thr Ala Ala Ala Val Arg Val Phe Glu  
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Glu Met Lys Gln Gln Gly Ile Ala Ala Ser Val Val Thr Tyr Asn Ser  
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Leu Ile Ser Gly Leu Cys Ser Glu Gly Lys Val Glu Glu Gly Val Lys  
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Leu Met Glu Glu Met Glu Asp Leu Gly Leu Ser Pro Asn Glu Ile Thr  
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Phe Gly Cys Val Leu Lys Gly Phe Cys Lys Lys Gly Met Met Ala Asp  
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Ala Asn Asp Trp Ile Asp Gly Met Thr Glu Arg Asn Val Glu Pro Asp  
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Val Val Ile Tyr Asn Ile Leu Ile Asp Val Tyr Arg Arg Leu Gly Lys  
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Met Glu Asp Ala Met Ala Val Lys Glu Ala Met Ala Lys Lys Gly Ile  
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Ser Pro Asn Val Thr Thr Tyr Asn Cys Leu Ile Thr Gly Phe Ser Arg  
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Ser Gly Asp Trp Arg Ser Ala Ser Gly Leu Leu Asp Glu Met Lys Glu  
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Lys Gly Ile Glu Ala Asp Val Val Thr Tyr Asn Val Leu Ile Gly Ala  
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Leu Cys Cys Lys Gly Glu Val Arg Lys Ala Val Lys Leu Leu Asp Glu  
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Met Ser Glu Val Gly Leu Glu Pro Asn His Leu Thr Tyr Asn Thr Ile  
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Ile Gln Gly Phe Cys Asp Lys Gly Asn Ile Lys Ser Ala Tyr Glu Ile  
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Arg Thr Arg Met Glu Lys Cys Arg Lys Arg Ala Asn Val Val Thr Tyr  
 500 505 510

Asn Val Phe Ile Lys Tyr Phe Cys Gln Ile Gly Lys Met Asp Glu Ala  
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Asn Asp Leu Leu Asn Glu Met Leu Asp Lys Cys Leu Val Pro Asn Gly  
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Ser Ser

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<211> 1365

<212> DNA

<213> Oryza sativa

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aaggagatgg aaagcaatgg tgttaatcct gatattatta catataacat aattctgcat 1200  
 ggtttatttc gcaccagaag aactgctgct gcaaaagaac tatatgccag gattaccgaa 1260  
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<210> 29

<211> 454

<212> PRT

<213> Oryza sativa

<400> 29

Met	Ala	Asp	Asp	Gly	Arg	Cys	Pro	Pro	Asp	Val	Val	Ser	Tyr	Asn	Thr
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20								25							

Thr	Tyr	His	Glu	Met	Leu	Asp	Arg	Arg	Val	Ser	Pro	Asp	Ala	Val	Thr
														45	
35							40								

Tyr	Asn	Ser	Ile	Ile	Ala	Ala	Leu	Ser	Lys	Ala	Gln	Ala	Met	Asp	Arg
														50	
50								55							60

Ala	Met	Glu	Val	Leu	Thr	Val	Met	Val	Met	Pro	Asn	Cys	Phe	Thr	Tyr
														65	
65							70								80

Asn	Ser	Ile	Met	His	Gly	Tyr	Cys	Ser	Ser	Gly	Gln	Ser	Glu	Lys	Ala
														85	
														90	

Ile	Gly	Ile	Phe	Arg	Lys	Met	Cys	Ser	Asp	Gly	Ile	Glu	Pro	Asp	Val
														100	
														105	

Val	Thr	Tyr	Asn	Ser	Leu	Met	Asp	Tyr	Leu	Cys	Lys	Asn	Gly	Lys	Cys
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									120					125	

Thr	Glu	Ala	Arg	Lys	Ile	Phe	Asp	Ser	Met	Val	Lys	Arg	Gly	Leu	Lys
														130	
														135	

Pro	Asp	Ile	Thr	Thr	Tyr	Gly	Thr	Leu	Leu	His	Gly	Tyr	Ala	Ser	Lys
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Gly	Ala	Leu	Val	Glu	Met	His	Asp	Leu	Leu	Ala	Leu	Met	Val	Gln	Asn
														165	
														170	

Gly	Met	Gln	Leu	Asp	His	His	Val	Phe	Asn	Ile	Leu	Ile	Cys	Ala	Tyr
														180	
														185	

Thr Lys Gln Glu Lys Val Asp Glu Val Val Leu Val Phe Ser Lys Met  
195 200 205

Arg Gln Gln Gly Leu Thr Pro Asn Ala Val Asn Tyr Arg Thr Val Ile  
210 215 220

Asp Gly Leu Cys Lys Leu Gly Arg Leu Asp Asp Ala Met Leu Asn Phe  
225 230 235 240

Glu Gln Met Ile Asp Lys Gly Leu Thr Pro Asn Val Val Val Tyr Thr  
245 250 255

Ser Leu Ile His Ala Leu Cys Thr Tyr Asp Lys Trp Glu Lys Ala Glu  
260 265 270

Glu Leu Ile Phe Glu Ile Leu Asp Gln Gly Ile Asn Pro Asn Ile Val  
275 280 285

Phe Phe Asn Thr Ile Leu Asp Ser Leu Cys Lys Glu Gly Arg Val Ile  
290 295 300

Glu Ser Lys Lys Leu Phe Asp Leu Leu Gly His Ile Gly Val Asn Pro  
305 310 315 320

Asp Val Ile Thr Tyr Ser Thr Leu Ile Asp Gly Tyr Cys Leu Ala Gly  
325 330 335

Lys Met Asp Gly Ala Met Lys Leu Leu Thr Gly Met Val Ser Val Gly  
340 345 350

Leu Lys Pro Asp Ser Val Thr Tyr Ser Thr Leu Ile Asn Gly Tyr Cys  
355 360 365

Lys Ile Asn Arg Met Glu Asp Ala Leu Ala Leu Phe Lys Glu Met Glu  
370 375 380

Ser Asn Gly Val Asn Pro Asp Ile Ile Thr Tyr Asn Ile Ile Leu His  
385 390 395 400

Gly Leu Phe Arg Thr Arg Arg Thr Ala Ala Ala Lys Glu Leu Tyr Ala  
405 410 415

Arg Ile Thr Glu Ser Gly Thr Gln Leu Glu Leu Ser Thr Tyr Asn Ile  
420 425 430

Ile Leu Met Asp Phe Ala Lys Thr Asn Ser Leu Met Met His Phe Gly  
435 440 445

Cys Phe Arg Thr Tyr Val  
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<210> 30  
<211> 1386  
<212> DNA  
<213> Oryza sativa

<400> 30  
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cgtggcaagg ggcgcacacat ctacggcttg aaccgcgccc tcgacgacgt cgccgcgtcac 180  
agccccgcgg ccgcgtgtc cgcgtacaac cgcatggccc gagccggcgc cgacgaggta 240  
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gacctcggtt tcgcggcctt gggcaatgtc attaagaagg gatttagagt ggaagccatc 360  
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atgcctgtatg atggaggtga ctgcacccacat gatgtggtgt tgtacaacac cgtcatcaat 600  
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caggggattt tgccagatgt tgtgacttac agctcttatta tcgctgcctt atgcaaggct 720  
caagctatgg acaaggccat ggaggtactt aacaccatgg ttaagaatgg tgtcatgcct 780  
aattgcagga catataatag tattgtgcac ggatattgct cttcaaggca gttgacagag 840  
gctattggat ttctcaaaat gatgtgcagt gatgggtcg aaccagatgt tggtaacttgc 900  
aacttgctga tggatttatct ttgcaagaac agaagatgca cggaaagctag aaagattttc 960  
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<210> 31  
<211> 461  
<212> PRT  
<213> Oryza sativa

<400> 31  
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Gln Gly Arg Gly Gly Arg Ala Gly Gly Asn Gly Ala Glu Asp Ala Arg  
20 25 30

His Val Phe Asp Glu Leu Leu Arg Arg Gly Lys Gly Ala Thr Ile Tyr			
35	40	45	
Gly Leu Asn Arg Ala Leu Asp Asp Val Ala Arg His Ser Pro Ala Ala			
50	55	60	
Ala Val Ser Arg Tyr Asn Arg Met Ala Arg Ala Gly Ala Asp Glu Val			
65	70	75	80
Thr Pro Asn Leu Tyr Thr Tyr Ser Val Leu Ile Gly Cys Cys Cys Arg			
85	90	95	
Ala Gly Arg Leu Asp Leu Gly Phe Ala Ala Leu Gly Asn Val Ile Lys			
100	105	110	
Lys Gly Phe Arg Val Glu Ala Ile Thr Phe Thr Pro Leu Leu Lys Gly			
115	120	125	
Leu Cys Ala Asp Lys Arg Thr Ser Asp Ala Met Asp Ile Val Leu Cys			
130	135	140	
Arg Met Thr Gln Leu Gly Cys Ile Pro Asn Val Phe Ser Cys Thr Ile			
145	150	155	160
Leu Leu Lys Gly Leu Cys Asp Glu Asn Arg Ser Gln Glu Ala Leu Glu			
165	170	175	
Leu Leu Gln Met Met Pro Asp Asp Gly Gly Asp Cys Pro Pro Asp Val			
180	185	190	
Val Leu Tyr Asn Thr Val Ile Asn Gly Phe Phe Lys Glu Gly Asp Pro			
195	200	205	
Asp Lys Ala Tyr Ala Thr Tyr His Glu Met Phe Asp Gln Gly Ile Leu			
210	215	220	
Pro Asp Val Val Thr Tyr Ser Ser Ile Ile Ala Ala Leu Cys Lys Ala			
225	230	235	240
Gln Ala Met Asp Lys Ala Met Glu Val Leu Asn Thr Met Val Lys Asn			
245	250	255	
Gly Val Met Pro Asn Cys Arg Thr Tyr Asn Ser Ile Val His Gly Tyr			
260	265	270	
Cys Ser Ser Gly Gln Leu Thr Glu Ala Ile Gly Phe Leu Lys Met Met			
275	280	285	

Cys Ser Asp Gly Val Glu Pro Asp Val Val Thr Cys Asn Leu Leu Met  
 290 295 300

Asp Tyr Leu Cys Lys Asn Arg Arg Cys Thr Glu Ala Arg Lys Ile Phe  
 305 310 315 320

Asn Ser Met Thr Lys Cys Gly Leu Lys Pro Asp Ile Thr Thr Tyr Cys  
 325 330 335

Thr Leu Leu Gln Gly Tyr Ala Thr Lys Gly Ala Leu Val Glu Met His  
 340 345 350

Asp Leu Leu Asp Leu Met Val Trp Asn Gly Ile Gln Pro Asn His His  
 355 360 365

Val Phe Asn Ile Leu Ile Cys Ala Tyr Ala Lys Gln Glu Lys Val Asp  
 370 375 380

Glu Ala Met Leu Val Phe Ser Lys Met Arg Gln Gln Gly Leu Ser Pro  
 385 390 395 400

Asn Ala Val Asn Tyr Arg Thr Val Ile Asp Val Leu Cys Lys Leu Gly  
 405 410 415

Arg Val Tyr Asp Ala Val Leu Thr Leu Lys Gln Met Ile Asn Glu Gly  
 420 425 430

Leu Thr Pro Asp Ile Ile Val Tyr Thr Pro Leu Ile His Gly Phe Cys  
 435 440 445

Thr Cys Asp Lys Trp Glu Lys Ala Glu Glu Leu Ile Phe  
 450 455 460

<210> 32

<211> 1521

<212> DNA

<213> Oryza sativa

<400> 32

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 ttcgacgaat tgctccggcg aggcatcgcc gctccgatcc gcagcttgaa cggcgctctc 180  
 gccgacgtcg cgccgcacaa ccccgccgccc gctgtgtccc gttcaaccg catggcacga 240  
 gctggtgcca gcatggtaac tcccaccgtg cacacctatg gcatcctcat cggctgctgc 300  
 tgcagtgcgg gccgcttaga cctcggttgc gcccgcgttgg gcatgtcgt taagaaggga 360  
 ttcagagtgg aaccatcat cttaatcct ctgctcaagg gcctctgtgc agacaagagg 420  
 acggacgacg caatggacat agtgctccgt ggaatgaccg agtcagctg cgtgccaaat 480

gtcttctccc acaccattat tctcaaggga ctctgtcatg agaacagaag ccaagaagct 540  
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 tacagcaccc tcacatcgatgg cctcttgaaa ggaggggatc cggacaaagc ctacgctaca 660  
 taccgtgaaa tgcttgaccg gaggatttg ccaaataatggtg tgatttacag ctccattatt 720  
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 aagaatggag ttacacccaa ttgcttcacg tatacttagtc ttgtgcatgg attttgctct 840  
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 ccaaataatggtg ttacttatag ctgcgttatg gactatctct gcaagaacgg aagatgcaca 960  
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 actgcaaaaag aattctatgt acagatttac aaaagtggca aaaaagatct tatagaacag 1440  
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<210> 33  
<211> 506  
<212> PRT  
<213> Oryza sativa

<400> 33  
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20 25 30

Gly Ala Glu Asp Ala Leu Asp Val Phe Asp Glu Leu Leu Arg Arg Gly  
35 40 45

Ile Gly Ala Pro Ile Arg Ser Leu Asn Gly Ala Leu Ala Asp Val Ala  
50 55 60

Arg Asp Asn Pro Ala Ala Val Ser Arg Phe Asn Arg Met Ala Arg  
65 70 75 80

Ala Gly Ala Ser Met Val Thr Pro Thr Val His Thr Tyr Gly Ile Leu  
85 90 95

Ile Gly Cys Cys Cys Ser Ala Gly Arg Leu Asp Leu Gly Phe Ala Ala  
100 105 110

Leu Gly His Val Val Lys Lys Gly Phe Arg Val Glu Pro Ile Ile Phe

115	120	125
Asn Pro Leu Leu Lys Gly Leu Cys Ala Asp Lys Arg Thr Asp Asp Ala		
130	135	140
Met Asp Ile Val Leu Arg Gly Met Thr Glu Leu Ser Cys Val Pro Asn		
145	150	155
160		
Val Phe Ser His Thr Ile Ile Leu Lys Gly Leu Cys His Glu Asn Arg		
165	170	175
Ser Gln Glu Ala Leu Glu Leu Leu His Met Met Ala Asp Asp Gly Gly		
180	185	190
Gly Cys Leu Pro Asn Val Val Ser Tyr Ser Thr Val Ile Asp Gly Leu		
195	200	205
Leu Lys Gly Gly Asp Pro Asp Lys Ala Tyr Ala Thr Tyr Arg Glu Met		
210	215	220
Leu Asp Arg Arg Ile Leu Pro Asn Val Val Ile Tyr Ser Ser Ile Ile		
225	230	235
240		
Ala Ala Leu Cys Lys Gly Gln Ala Met Asp Lys Ala Met Glu Val His		
245	250	255
Asp Arg Met Val Lys Asn Gly Val Thr Pro Asn Cys Phe Thr Tyr Thr		
260	265	270
Ser Leu Val His Gly Phe Cys Ser Ser Gly Gln Leu Thr Glu Ala Ile		
275	280	285
Lys Phe Leu Glu Lys Met Cys Ser Asn Gly Val Glu Pro Asn Val Val		
290	295	300
Thr Tyr Ser Ser Phe Met Asp Tyr Leu Cys Lys Asn Gly Arg Cys Thr		
305	310	315
320		
Glu Ala Arg Lys Ile Phe Asp Ser Met Val Lys Arg Gly Leu Lys Pro		
325	330	335
Asp Ile Thr Thr Tyr Ser Ser Leu Leu His Gly Tyr Ala Ile Glu Gly		
340	345	350
Ala Leu Val Glu Met His Gly Leu Phe Asp Leu Met Val Gln Ser Asp		
355	360	365
Met Gln Pro Asp His Tyr Val Phe Asn Thr Leu Ile Tyr Ala Ser Ala		

370	375	380
Lys Gln Gly Lys Val Asp Glu Ala Met Leu Val Phe Ser Lys Met Arg		
385	390	395
395		
Gln Gln Gly Leu Lys Pro Asn Cys Val Thr Tyr Ser Thr Leu Ile Asn		
405		
410		
415		
Gly Tyr Cys Lys Ile Thr Arg Met Glu Asn Ala Leu Ala Leu Phe Gln		
420	425	430
430		
Glu Met Val Ser Asn Gly Val Ser Pro Asn Phe Ile Thr Tyr Asn Ile		
435	440	445
445		
Met Leu Gln Gly Leu Phe Arg Thr Gly Arg Thr Ala Thr Ala Lys Glu		
450	455	460
460		
Phe Tyr Val Gln Ile Ile Lys Ser Gly Lys Lys Asp Leu Ile Glu Gln		
465	470	475
475		
480		
Gly Leu Leu Glu Glu Leu Asp Asp Leu Phe Leu Ser Met Glu Asp Asn		
485		
490		
495		
Asp Cys Ser Thr Val Ser Thr Pro Ala Cys		
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<210> 34  
<211> 1884  
<212> DNA  
<213> Oryza sativa
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<400> 34  
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gacgaattgc tccgtcgtgg cataccagat gtcttctcct acaatattct tctcaacggg 180  
ctgtgtgatg agaacagaag ccaagaagct ctcgagttac tgcacataat ggctgtatgt 240  
ggaggtgact gcccacctga tgtggtgtcg tacagcaccg tcatcaatgg cttcttcaag 300  
gagggggatc tggacaaaat gcttgaccag aggatttgc caaatgttgt gacctacaac 360  
tctattattg ctgcgtatg caaggctcaa actgtggaca aggccatgg ggtacttacc 420  
accatggta agagtggtgt catgcctgat tgcatgacat ataatagtat tgtgcattggg 480  
ttttgcctt cagggcagcc gaaagaggct attgtatttc tcaaaaagat ggcgcagtgtat 540  
ggtgtcgaac cagatgttgt tacttataac tcgctcatgg attatctttg caagaacgg 600  
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 aatttcatgt ttagggaact gttcagaga ggtgagataa ccagggctgg cacttacctt 1740  
 tccatgattt gatgagaagca ctttcctc gaagcatcca ctgcttcctt gtttataat 1800  
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<210> 35

<211> 627

<212> PRT

<213> Oryza sativa

<400> 35

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20														30

Ala	Glu	Asp	Ala	Arg	His	Val	Phe	Asp	Glu	Leu	Leu	Arg	Arg	Gly	Ile
35															45

Pro	Asp	Val	Phe	Ser	Tyr	Asn	Ile	Leu	Leu	Asn	Gly	Leu	Cys	Asp	Glu
50															60

Asn	Arg	Ser	Gln	Glu	Ala	Leu	Glu	Leu	Leu	His	Ile	Met	Ala	Asp	Asp
65															80

Gly	Gly	Asp	Cys	Pro	Pro	Asp	Val	Val	Ser	Tyr	Ser	Thr	Val	Ile	Asn
85															95

Gly	Phe	Phe	Lys	Glu	Gly	Asp	Leu	Asp	Lys	Met	Leu	Asp	Gln	Arg	Ile
100															110

Ser	Pro	Asn	Val	Val	Thr	Tyr	Asn	Ser	Ile	Ile	Ala	Ala	Leu	Cys	Lys
115															125

Ala Gln Thr Val Asp Lys Ala Met Glu Val Leu Thr Thr Met Val Lys  
 130                    135                    140

Ser Gly Val Met Pro Asp Cys Met Thr Tyr Asn Ser Ile Val His Gly  
 145                    150                    155                    160

Phe Cys Ser Ser Gly Gln Pro Lys Glu Ala Ile Val Phe Leu Lys Lys  
 165                    170                    175

Met Arg Ser Asp Gly Val Glu Pro Asp Val Val Thr Tyr Asn Ser Leu  
 180                    185                    190

Met Asp Tyr Leu Cys Lys Asn Gly Arg Cys Thr Glu Ala Arg Lys Ile  
 195                    200                    205

Phe Asp Ser Met Thr Lys Arg Gly Leu Lys Pro Asp Ile Thr Thr Tyr  
 210                    215                    220

Gly Thr Leu Leu Gln Gly Tyr Ala Thr Lys Gly Ala Leu Val Glu Met  
 225                    230                    235                    240

His Gly Leu Leu Asp Leu Met Val Arg Asn Gly Ile His Pro Asn His  
 245                    250                    255

Tyr Val Phe Ser Ile Leu Val Cys Ala Tyr Ala Lys Gln Glu Lys Val  
 260                    265                    270

Glu Glu Ala Met Leu Val Phe Ser Lys Met Arg Gln Gln Gly Leu Asn  
 275                    280                    285

Pro Asn Ala Val Thr Tyr Gly Thr Val Ile Asp Val Leu Cys Lys Ser  
 290                    295                    300

Gly Arg Val Glu Asp Ala Met Leu Tyr Phe Glu Gln Met Ile Asp Glu  
 305                    310                    315                    320

Gly Leu Arg Pro Asp Ser Ile Val Tyr Asn Ser Leu Ile His Ser Leu  
 325                    330                    335

Cys Ile Phe Asp Lys Trp Glu Lys Ala Glu Glu Leu Phe Leu Glu Met  
 340                    345                    350

Leu Asp Arg Gly Ile Cys Leu Ser Thr Ile Phe Phe Asn Ser Ile Ile  
 355                    360                    365

Asp Ser His Cys Lys Glu Gly Arg Val Ile Glu Ser Gly Lys Leu Phe  
 370                    375                    380

Asp Leu Met Val Arg Ile Gly Val Lys Pro Asp Ile Ile Thr Leu Gly  
385 390 395 400

Arg Phe Leu Gly Ser Ala Arg Arg Asp Tyr Ser Leu Phe Val Asn Ile  
405 410 415

Tyr Phe Ile Phe Thr Asn Met Ser Asn Thr Gly Asp Lys Glu Lys Glu  
420 425 430

Thr Pro Val Asn Thr Asn Gly Gly Asn Thr Ala Ser Asn Ser Ser Gly  
435 440 445

Gly Pro Phe Leu Gly Thr Tyr Asn Ile Ile Leu His Gly Leu Cys Lys  
450 455 460

Asn Lys Leu Thr Asp Asp Ala Leu Arg Met Phe Gln Asn Leu Cys Leu  
465 470 475 480

Met Asp Leu Lys Leu Glu Ala Arg Thr Phe Asn Ile Met Ile Asp Ala  
485 490 495

Leu Leu Lys Val Gly Arg Asn Asp Glu Ala Lys Asp Leu Phe Val Ala  
500 505 510

Phe Ser Ser Asn Gly Leu Val Pro Asn Tyr Trp Thr Tyr Arg Leu Met  
515 520 525

Ala Glu Asn Ile Ile Gly Gln Gly Leu Leu Glu Glu Leu Asp Gln Leu  
530 535 540

Phe Leu Ser Met Glu Asp Asn Gly Cys Thr Val Asp Ser Gly Met Leu  
545 550 555 560

Asn Phe Ile Val Arg Glu Leu Leu Gln Arg Gly Glu Ile Thr Arg Ala  
565 570 575

Gly Thr Tyr Leu Ser Met Ile Asp Glu Lys His Phe Ser Leu Glu Ala  
580 585 590

Ser Thr Ala Ser Leu Phe Ile Asp Leu Leu Ser Gly Gly Lys Tyr Gln  
595 600 605

Glu Tyr His Ile Phe Leu Pro Glu Lys Tyr Lys Ser Phe Ile Glu Ser  
610 615 620

Leu Ser Cys  
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<210> 36

<211> 1554

<212> DNA

<213> Oryza sativa

<400> 36

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<211> 517

<212> PRT

<213> Oryza sativa

<400> 37

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Cys	Cys	Cys	Arg	Ala	Gly	Arg	Leu	Asp	Leu	Gly	Phe	Ala	Ala	Leu	Gly
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260								265						270	
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Lys Glu Gly Asp Ser Asp Lys Ala Tyr Ser Thr Tyr His Glu Met Leu  
 355 360 365

Asp Arg Arg Ile Ser Pro Asn Val Val Thr Tyr Ser Ser Ile Ile Ala  
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 385 390 395 400

Thr Met Val Lys Asn Gly Val Met Pro Asp Cys Met Thr Tyr Asn Ser  
 405 410 415

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Thr Leu Lys Lys Met Arg Ser Asp Gly Val Glu Pro Asn Val Val Thr  
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Ala Arg Lys Ile Phe Asp Ser Met Thr Lys Arg Gly Leu Glu Pro Asp  
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Lys Tyr Leu Glu Lys  
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<211> 2784

<212> DNA

<213> Oryza sativa

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 35 40 45  
 Val Thr Glu Ser Glu Glu Asp Ala Ala Ala Val Gly Arg Asp Thr Arg  
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 Ala Pro Pro Ser Ile Gly Gly Ile Ala Arg Gly Ala Pro Arg Val Gly  
 65 70 75 80  
 Cys Asn Gly Gly Ala Ala Asp Asp Glu Glu Val Glu Arg Lys Ala  
 85 90 95  
 Arg Ala Val Ala Arg Ile Lys Leu Cys His Glu Leu Leu Arg Glu Arg  
 100 105 110  
 Arg Trp Arg Ala Met Arg Ala Ala Leu Ala Gln Leu Val Thr Glu Gln  
 115 120 125  
 Gly Ser Gly Ser Ala Ala Ala Leu Cys Asp Ile Leu Trp Asn Arg Phe  
 130 135 140  
 Arg Glu Cys Asp Ser Asn Gly Cys Val Trp Asp Ala Leu Ala Asn Ser  
 145 150 155 160  
 Tyr Ala Arg Ala Gln Met Val His Asp Ala Leu Tyr Val Leu Ser Lys  
 165 170 175  
 Met Ser Ser Leu Asn Met Gln Ile Ser Val Phe Thr Tyr Asp Ser Leu  
 180 185 190  
 Leu His Gly Leu Arg Met Thr Asp Val Ala Leu Glu Leu Phe Glu Glu

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Ile Asn Gly Leu Cys Lys Gln Asp Lys Val Gly Glu Ala Leu Ser Phe		
225	230	235
Leu Gln Glu Ala Arg Lys Glu Gly Lys Phe Lys Pro Leu Gly Met Thr		
245	250	255
Phe Asn Ile Leu Met Ser Ala Leu Cys Asn Trp Gly Phe Val Gln Ser		
260	265	270
Ala Lys Ser Phe Leu Cys Leu Met Leu Lys Tyr Gly Leu Val Pro Asp		
275	280	285
Arg Tyr Thr Phe Ser Thr Leu Ile His Gly Leu Cys Lys Val Gly Ser		
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Met Glu Glu Ala Leu Asp Leu Phe Glu Arg Val Thr Lys Glu Gly Met		
305	310	315
Glu Leu Glu Ile Val Thr Tyr Asn Ser Leu Ile Asn Gly Tyr Arg Leu		
325	330	335
Leu Gly Leu Thr Lys Glu Ile Pro Lys Ile Ile Gln Met Met Arg Gly		
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Gln Gly Val Glu Pro Asp Leu Val Thr Tyr Thr Ile Leu Ile Ala Gly		
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His Cys Glu Ser Gly Asp Val Glu Glu Gly Met Lys Val Arg Lys Asp		
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Val Leu Asp Gln Gly Leu Gln Leu Asn Ile Val Thr Tyr Ser Val Leu		
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Leu Asn Ala Leu Phe Lys Lys Gly Met Phe Cys Glu Ile Asp Asn Leu		
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Leu Gly Glu Ile Tyr Asn Ile Gly Leu Asp Met Asp Val Ile Ala Tyr		
420	425	430
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435	440	445
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465	470	475
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485	490	495
Pro Thr Asp Val Val Phe Tyr Asn Val Val Ile Asp Gly Tyr Ala Lys		
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Ala Gly Met His Pro Thr Ile Val Thr Cys Asn Ser Leu Leu Tyr Gly		
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Tyr Cys Lys Ile Gly Asp Leu Gln Leu Ala Glu Ser Tyr Phe Arg Ala		
545	550	555
Ile Gln Leu Ser Gly Leu Leu Pro Thr Ala Val Thr Tyr Thr Leu		
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Met Asp Ala Leu Ser Glu Ala Gly Glu Val Asn Thr Met Leu Ser Leu		
580	585	590
Phe Asp Glu Met Val Ala Lys Arg Ile Lys Ala Asn Ala Val Thr Tyr		
595	600	605
Ser Val Ile Val Lys Gly Leu Cys Lys Gln Leu Arg Phe Asp Glu Ala		
610	615	620
Ile Asn Val Leu Lys Asp Met Asp Ser Lys Gly Ile Asn Ala Asp Pro		
625	630	635
Ile Thr Tyr Asn Thr Leu Ile Gln Gly Phe Cys Glu Ser Glu Asn Val		
645	650	655
Gln Met Ala Phe His Ile His Asp Ile Met Leu Cys Arg Gly Leu Val		
660	665	670
Pro Thr Pro Val Thr Tyr Asn Leu Leu Ile Asn Val Leu Cys Leu Lys.		
675	680	685
Gly Lys Val Ile Gln Ala Glu Ile Leu Leu Glu Ser Leu Arg Glu Asn		
690	695	700
Gly Ile Lys Leu Arg Lys Phe Ala Tyr Thr Thr Leu Ile Lys Ala Gln		

705

710

715

720

Cys Ala Lys Gly Met Pro Ile Asn Ala Val Leu Leu Val Gly Lys Leu  
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Leu Asp Ala Gly Phe Glu Ala Ser Ile Glu Asp Phe Ser Ala Ala Ile  
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Asn Arg Leu Cys Lys Arg Gln Phe Ala Lys Glu Ala Phe Met Phe Val  
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Pro Ile Met Leu Ser Val Gly Ile Tyr Pro Asp Thr Gln Ile Tyr Cys  
 770 775 780

Val Leu Gly Arg Ala Leu Gln Lys Asn Ser Glu Leu Val Tyr Leu Pro  
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&lt;211&gt; 1779

&lt;212&gt; DNA

&lt;213&gt; Arabidopsis thaliana

&lt;400&gt; 40

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Arg Leu Arg Ser Gly Ile Val Asp Ile Lys Lys Asp Asp Ala Ile Ala  
 50 55 60

Leu Phe Gln Glu Met Ile Arg Ser Arg Pro Leu Pro Ser Leu Val Asp  
 65 70 75 80

Phe Ser Arg Phe Phe Ser Ala Ile Ala Arg Thr Lys Gln Phe Asn Leu  
 85 90 95

Val Leu Asp Phe Cys Lys Gln Leu Glu Leu Asn Gly Ile Ala His Asn  
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Ile Tyr Thr Leu Asn Ile Met Ile Asn Cys Phe Cys Arg Cys Cys Lys  
 115 120 125

Thr Cys Phe Ala Tyr Ser Val Leu Gly Lys Val Met Lys Leu Gly Tyr  
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Glu Pro Asp Thr Thr Phe Asn Thr Leu Ile Lys Gly Leu Phe Leu  
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Glu Gly Lys Val Ser Glu Ala Val Val Leu Val Asp Arg Met Val Glu

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Met Glu Glu Arg Asn Val Lys Ala Asp Val Phe Thr Tyr Ser Thr Ile		
210	215	220
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Ala Leu Leu Leu Lys Asp Met Val Ser Arg Glu Ile Val Pro Asn Val		
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325	330	335
Asn Arg Leu Ser Glu Ala Asn Asn Met Leu Asp Leu Met Val Arg Asn		
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Lys Cys Ser Pro Asp Ile Val Thr Phe Thr Ser Leu Ile Lys Gly Tyr		
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Cys Met Val Lys Arg Val Asp Asp Gly Met Lys Val Phe Arg Asn Ile		
370	375	380
Ser Lys Arg Gly Leu Val Ala Asn Ala Val Thr Tyr Ser Ile Leu Val		
385	390	395
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Gln Glu Met Val Ser His Gly Val Leu Pro Asp Val Met Thr Tyr Gly		

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465	470	475
Asp Ala Trp Asn Leu Phe Cys Ser Leu Pro Cys Lys Gly Val Lys Pro		
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Arg Asp Gly Asp Leu Thr Ala Ser Ala Lys Leu Ile Glu Glu Met Lys		
545	550	555
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580	585	590

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 <212> DNA  
 <213> Petunia sp.

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Cys Ser Ile Ser Val Lys Gly Asn Phe Gly Val Ser Asn Glu Phe Gln			
35	40	45	
Asn Val Lys Cys Leu Asp Asp Ala Phe Ser Leu Phe Arg Gln Met Val			
50	55	60	
Arg Thr Lys Pro Leu Pro Ser Val Ala Ser Phe Ser Lys Leu Leu Lys			
65	70	75	80
Ala Met Val His Met Lys His Tyr Ser Ser Val Val Ser Leu Phe Arg			
85	90	95	
Glu Ile His Lys Leu Arg Ile Pro Val His Glu Phe Ile Leu Ser Ile			
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Val Val Asn Ser Cys Cys Leu Met His Arg Thr Asp Leu Gly Phe Ser			
115	120	125	
Val Leu Ala Ile His Phe Lys Lys Gly Ile Pro Tyr Asn Glu Val Thr			
130	135	140	
Phe Thr Thr Leu Ile Arg Gly Leu Phe Ala Glu Asn Lys Val Lys Asp			
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Ala Val His Leu Phe Lys Lys Leu Val Arg Glu Asn Ile Cys Glu Pro  
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Asn Glu Val Met Tyr Gly Thr Val Met Asn Gly Leu Cys Lys Lys Gly  
180 185 190

His Thr Gln Lys Ala Phe Asp Leu Leu Arg Leu Met Glu Gln Gly Ser  
195 200 205

Thr Lys Pro Asn Thr Arg Thr Tyr Thr Ile Val Ile Asp Ala Phe Cys  
210 215 220

Lys Asp Gly Met Leu Asp Gly Ala Thr Ser Leu Leu Asn Glu Met Lys  
225 230 235 240

Gln Lys Ser Ile Pro Pro Asp Ile Phe Thr Tyr Ser Thr Leu Ile Asp  
245 250 255

Ala Leu Cys Lys Leu Ser Gln Trp Glu Asn Val Arg Thr Leu Phe Leu  
260 265 270

Glu Met Ile His Leu Asn Ile Tyr Pro Asn Val Cys Thr Phe Asn Ser  
275 280 285

Val Ile Asp Gly Leu Cys Lys Glu Gly Lys Val Glu Asp Ala Glu Glu  
290 295 300

Ile Met Arg Tyr Met Ile Glu Lys Gly Val Asp Pro Asp Val Ile Thr  
305 310 315 320

Tyr Asn Met Ile Ile Asp Gly Tyr Gly Leu Arg Gly Gln Val Asp Arg  
325 330 335

Ala Arg Glu Ile Phe Asp Ser Met Ile Asn Lys Ser Ile Glu Pro Asp  
340 345 350

Ile Ile Ser Tyr Asn Ile Leu Ile Asn Gly Tyr Ala Arg Gln Lys Lys  
355 360 365

Ile Asp Glu Ala Met Gln Val Cys Arg Glu Ile Ser Gln Lys Gly Leu  
370 375 380

Lys Pro Ser Ile Val Thr Cys Asn Val Leu Leu His Gly Leu Phe Glu  
385 390 395 400

Leu Gly Arg Thr Lys Ser Ala Gln Asn Phe Phe Asp Glu Met Leu Ser  
405 410 415

Ala Gly His Ile Pro Asp Leu Tyr Thr His Cys Thr Leu Leu Gly Gly  
 420 425 430

Tyr Phe Lys Asn Gly Leu Val Glu Glu Ala Met Ser His Phe His Lys  
 435 440 445

Leu Glu Arg Arg Arg Glu Asp Thr Asn Ile Gln Ile Tyr Thr Ala Val  
 450 455 460

Ile Asp Gly Leu Cys Lys Asn Gly Lys Leu Asp Lys Ala His Ala Thr  
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Phe Glu Lys Leu Pro Leu Ile Gly Leu His Pro Asp Val Ile Thr Tyr  
 485 490 495

Thr Ala Met Ile Ser Gly Tyr Cys Gln Glu Gly Leu Leu Asp Glu Ala  
 500 505 510

Lys Asp Met Leu Arg Lys Met Glu Asp Asn Gly Cys Leu Ala Asp Asn  
 515 520 525

Arg Thr Tyr Asn Val Ile Val Arg Gly Phe Leu Arg Ser Asn Lys Val  
 530 535 540

Ser Glu Met Lys Ala Phe Leu Glu Glu Ile Ala Gly Lys Ser Phe Ser  
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Pro Ser Ile Thr Arg Lys Met His Trp Ile Lys Leu His Ile Ala  
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<211> 1779

<212> DNA

<213> Petunia sp.

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 <211> 592  
 <212> PRT  
 <213> Petunia sp.

<400> 45

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Arg	Ser	Ile	Ser	Val	Lys	Gly	Asn	Phe	Gly	Val	Ser	Asn	Glu	Phe	Glu
35					40							45			

Asn	Val	Lys	Cys	Leu	Asp	Asp	Ala	Phe	Ser	Leu	Phe	Arg	Gln	Met	Val
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Arg	Thr	Lys	Pro	Leu	Pro	Ser	Val	Val	Ser	Phe	Ser	Lys	Leu	Leu	Lys
65				70				75				80			

Ala	Leu	Val	His	Met	Lys	His	Tyr	Ser	Ser	Val	Val	Ser	Leu	Phe	Arg
85								90				95			

Glu Ile His Lys Leu Arg Ile Pro Val His Glu Phe Ile Leu Ser Ile  
100 105 110

Val Val Asn Ser Cys Cys Leu Met His Arg Thr Asp Leu Gly Phe Ser  
115 120 125

Val Leu Ala Ile His Phe Lys Lys Gly Ile Pro Phe Asn Gln Val Ile  
130 135 140

Phe Asn Thr Leu Leu Arg Gly Leu Phe Ala Glu Asn Lys Val Lys Asp  
145 150 155 160

Ala Val His Leu Phe Lys Lys Leu Val Arg Glu Asn Ile Cys Glu Pro  
165 170 175

Asn Glu Val Met Tyr Gly Thr Val Met Asn Gly Leu Cys Lys Lys Gly  
180 185 190

His Thr Gln Lys Ala Phe Asp Leu Leu Arg Leu Met Glu Gln Gly Ser  
195 200 205

Thr Lys Pro Asn Thr Cys Ile Tyr Ser Ile Val Ile Asp Ala Phe Cys  
210 215 220

Lys Asp Gly Met Leu Asp Gly Ala Thr Ser Leu Leu Asn Glu Met Lys  
225 230 235 240

Gln Lys Ser Ile Pro Pro Asp Ile Phe Thr Tyr Ser Thr Leu Ile Asp  
245 250 255

Ala Leu Cys Lys Leu Ser Gln Trp Glu Asn Val Arg Thr Leu Phe Leu  
260 265 270

Glu Met Ile His Leu Asn Ile Tyr Pro Asn Val Cys Thr Phe Asn Ser  
275 280 285

Val Ile Asp Gly Leu Cys Lys Glu Gly Lys Val Glu Asp Ala Glu Glu  
290 295 300

Ile Met Arg Tyr Met Ile Glu Lys Gly Val Asp Pro Asp Val Ile Thr  
305 310 315 320

Tyr Asn Met Ile Ile Asp Gly Tyr Gly Leu Arg Gly Gln Val Asp Arg  
325 330 335

Ala Arg Glu Ile Phe Asp Ser Met Ile Asn Lys Ser Ile Glu Pro Asn  
340 345 350

Ile Ile Ser Tyr Asn Ile Leu Ile Asn Gly Tyr Ala Arg Gln Lys Lys  
 355 360 365

Ile Asp Glu Ala Met Gln Val Cys Arg Glu Ile Ser Gln Lys Gly Leu  
 370 375 380

Lys Pro Ser Ile Val Thr Cys Asn Val Leu Leu His Gly Leu Phe Glu  
 385 390 395 400

Leu Gly Arg Thr Lys Ser Ala Gln Asn Phe Phe Asp Glu Met Leu Ser  
 405 410 415

Ala Gly His Ile Pro Asp Leu Tyr Thr His Cys Thr Leu Leu Gly Gly  
 420 425 430

Tyr Phe Lys Asn Gly Leu Val Glu Glu Ala Met Ser His Phe His Lys  
 435 440 445

Leu Glu Arg Arg Arg Glu Asp Thr Asn Ile Gln Ile Tyr Thr Ala Val  
 450 455 460

Ile Asp Gly Leu Cys Lys Asn Gly Lys Leu Asp Lys Ala His Ala Thr  
 465 470 475 480

Phe Glu Lys Leu Pro Leu Ile Gly Leu His Pro Asp Val Ile Thr Tyr  
 485 490 495

Thr Ala Met Ile Ser Gly Tyr Cys Gln Glu Gly Leu Leu Asp Glu Ala  
 500 505 510

Lys Asp Met Leu Arg Lys Met Glu Asp Asn Gly Cys Leu Ala Asp Asn  
 515 520 525

Arg Thr Tyr Asn Val Ile Val Arg Gly Phe Leu Arg Ser Asn Lys Val  
 530 535 540

Ser Glu Met Lys Ala Phe Leu Glu Glu Ile Ala Gly Lys Ser Phe Ser  
 545 550 555 560

Phe Glu Ala Ala Thr Val Glu Leu Leu Met Asp Ile Ile Ala Glu Asp  
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Pro Ser Leu Leu Asn Met Ile Pro Glu Phe His Arg Asp Asn Lys Lys  
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<212> DNA  
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<210> 47  
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<220>  
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<400> 47  
tttatgatac atggatttca acgac 25

<210> 48  
<211> 25  
<212> DNA  
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<223> Description of Artificial Sequence: Primer

<400> 48  
tgaaaatgac aatcgtaaca gaaaa 25

<210> 49  
<211> 24  
<212> DNA  
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<223> Description of Artificial Sequence: Primer

<400> 49  
aacatttcctc cagacattat taca 24

<210> 50  
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<212> DNA  
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<220>  
<223> Description of Artificial Sequence: Primer

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24

<210> 51  
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<212> DNA  
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<223> Description of Artificial Sequence: Primer

<400> 51  
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27

<210> 52  
<211> 31  
<212> DNA  
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<220>  
<223> Description of Artificial Sequence: Primer

<400> 52  
gaattcactt tgctctcacg ataaaactaag a

31